



## **A COMPREHENSIVE ASSESSMENT OF ANALYTICAL TECHNIQUES FOR QUANTIFYING THE NUCLEOSIDE ANALOGUE REVERSE TRANSCRIPTASE INHIBITOR (NRTIS) STAVUDINE**

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### **Abstract**

HIV/AIDS should always be treated for the remainder of their lives with influential life-saving drugs such as nucleoside reverse transcriptase inhibitors, non-nucleoside reverse transcriptase inhibitors, and protease inhibitors. HIV/AIDS remains a major global public health problem, having claimed the lives of 40.1 million [33.6-48.6 million] people to date. In 2021, 650 000 [510 000-860 000] people may have died from HIV-related causes, and 1.5 million [1.1-2.0 million] will have become infected. HIV infection has no cure exists. There is currently no cure for HIV/AIDS. Once infected, your body is unable to rid itself of it. There are, however, innumerable medications that can control HIV and prevent complications. These drugs are known as antiretroviral therapy (ART). Based on their molecular mechanism and resistance profiles, these drugs are separated into six distinct classes: (1) Reverse transcriptase inhibitors predicated on nucleoside aptamers. Just before to 1996, there had been few antiretroviral treatment options for HIV-1 infection. Prophylaxis against common opportunistic pathogens and management of AIDS-related illnesses composed the major part of HIV-1 clinical management. The development of inhibitors of reverse transcriptase and protease, two of HIV-1's three essential enzymes, and the introduction of drug regimens that combined these agents to increase overall efficacy and durability of therapy reinvented the treatment of HIV-1 infection in the mid-1990s. A timeline for both the development and approval of antiviral drugs for human use.

**Keywords:** 'analysis' 'NRTIs' and STA

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## 1. Introduction

IUPAC name on stavudine as 1-[ (2R,5S) -5-(hydroxymethyl)-2,5-dihydrofuran-2-yl] -5-methyl-1,2,3,4-tetrahydropyrimidine-2,4-dione. Formula Molecular Its racemic mixture C<sub>10</sub>H<sub>12</sub>N<sub>2</sub>O<sub>4</sub>, also termed as 2',3'-didehydro-3'-deoxythymidine, is a white, crystalline solid or powder with little to no smell. Stavudine appears to be 83 mg/mL dissolved in water and 30 mg/mL accessible in propylene glycol at 23° C.

Approaches such as emission spectra, chromatography, critical assessment, capillary electrophoresis, and chemometrically assisted methods can be used to calculate the amount of ketones existing in dosage forms and biological fluids, etcetera. Figure 2 is a summary of the analytical methods used to figure out STA levels. Figure 3 shows how the number of ways to test for STA changed from 1978 to 2022.

This objective seeks to simplify, contextualise, and discuss the many analytical procedures that can be used to measure STA in a variety of compositions and biological matrices. This will fulfil assignment criteria (Figure 4). Volumetric analysis, optical

methods, chromatographic, electro analysis tools, linear electrochemical techniques, bioanalytical methods, and chemometric analysis were frequent studies. Capillary electrophoretic and electro analytical tests are many other.

### Volumetric Approaches

#### Evaluation via Titrimetry

In the field of science, titration is implemented as a quantitative method of analysis. How much of a renowned chemical must be introduced to a solution whose concentration has been established in order to validate the reaction The method was evaluated in light of the ICH recommendations

### Integrating Optical Approaches

#### Spectroscopic techniques in the UV/visible area

A spectrophotometer is an external device of light that somehow a sample emanates at a multitude of wavelengths. Measurement tool that is extremely exact. Spectrophotometry is a testing tool that is designed when other, more complex and costlier methods, like GLC or HPLC, are inaccessible. These speedy, minimal, and simple methods come with lots of pros

“Table 1 :Spectrophotometry”

met hod	matrices	solvent	lambda max	linearity	lod	loq	ref
UV	bulk and pharmaceutical dosage forms	N/A	250nm	4-20 mcg / ml.	N/A	N/A	1
UV	bulk and pharmaceutical dosage forms	phosphate buffer pH 6.8	266 nm	3-24 µg/mL	N/A	N/A	2
UV	bulk and pharmaceutical dosage forms	Methanol, Distilled water	250 nm	2-20 µg / ml	0.45	1.35	1
UV	bulk and pharmaceutical dosage forms	Bromate-bromide mixture	A520 B610 C550nm	A 0.125–1.75 B 1.0–10.0 C 1.0–9.0	A 0.03 B 0.15 C0.11	A 0.11 B 0.44 C 0.34	3

### Spectrophotometry combined

analyte	method	matrices	solvent	lambda max	linearity	lod	loq	r e f
(STA), (LMV) and (NVP)	UV spectroscopy method	tablet formulations	Phosphate buffer pH – 6.8	t 266, 271 and 315 nm	10 to 30 µg / ml	N/A	N/A	4
STA and chloranilic acid	uv spectroscopy method	bulk and dosage forms.	1,4-dioxan	526 nm	N/A	N/A	N/A	5
METH OD A								

STA	uv spectroscopy method	bulk and pharmaceutical formulations	1,2-naphthaquinone-4-sulfonic acid	464 nm	2.0-12.0 µg/mL	N/A	N/A	6
METHOD B								
STA	uv spectroscopy method	bulk and pharmaceutical formulation	1,10-phenanthroline	480 nm	5.0-30.0 µg/mL	N/A	N/A	
(NVP), LMV and STA	HPLC and spectrophotometric methods	tablet/capsule	N/A	300, 285, and 270 nm		N/A	N/A	7
NVP, LMV, STA	UV visible spectrophotometer	bulk and dosage forms.	Water 0.01 N HCl pH 4.5 Acetate buffer pH 6.8 phosphate buffer	N/A	N/A	N/A	N/A	8
LMV, NVP, STA	HPLC and UV spectrophotometric	pharmaceutical dosage forms	HPLC grade methanol, acetonitrile and isopropyl alcohol					9
METHOD 1 HPLC						0.17 ± 0.07, 0.03 ± 0.01, 0.12 ± 0.1	0.52 ± 0.22, 0.09 ± 0.05, 0.37 ± 0.05	
METHOD 2 UV						0.19, 0.29, 0.25	0.19, 0.29, 0.26	
STA, LMV, NVP	UV spectroscopy, reverse phase HPLC, and HPTLC	tablets	sodium phosphate buffer, acetonitrile, with pH adjusted to 3.5 using phosphoric acid	265 nm	s 0.8–6.4 g/ml (r = 0.996), 4–32g/ml (r = 0.9962) and 5.33–42.64g/ml (r = 0.9843)	N/A	N/A	10

### Chromatographic protocols

#### Optimal Liquid Chromatography

It is the most extensively used chromatographic separation technique medication compounds. Lc is an effective strategy and for its sensitivity, persistence, and efficiency. Sridhar, N. Y. et.al used

UPLC for analysis of LMV, NVP,STA, using methanol as solvent and column used is acquity x-bridge column 9(50 x4.6mm, 3.5 um) using phosphate buffer of ph2.2 is better compared to other reported method

“Table 2: Chromatography combined”

analyte	method	matrices	mobile phase	stationary phase	detection	flow rate	linearity	LOD	LOQ	ref
STA, LMV, NVP	UPLC	TABLET FORMULATION	gradient elution of 0.1% perchloric acid in water adjusted the pH 2.6 with dil sodium hydroxide solution and acetonitril	Acquity UPLC HSS T3 (2.1 × 100 mm), 1.8 μm column	UV detection	0.2 mL/min	values of correlation coefficients greater than 0.995	%w/w for LA-Imp-1 to, LA-Imp-6, ST-Imp-1 to ST-Imp-3, NE-Imp-1, to NE-Imp-2 was 0.010, 0.010, 0.010, 0.011, 0.013, 0.009, 0.024, 0.020, 0.019, 0.015, 0.010 and 0.011	%w/w for LA-Imp-1 to, LA-Imp-6, to ST-Imp-3, NE-Imp-1, to NE-Imp-2 is 0.020, 0.019, 0.020, 0.021, 0.026, 0.018, 0.049, 0.040, 0.039, 0.030, 0.021 and 0.022	11
STA		CAP SULFES								12
STA	RR-LC	bulk drug	water and methanol in the ratio of 85:15 (v/v)	C18	UV detection	0.5 mL/min	he correlation coefficient obtained was greater than 0.999	N/A	.500 μg mL <sup>-1</sup>	8
STA	Isocratic, simultaneous	bulk drug	mixt. of Et acetate with 4% (vol./vol.) of ethylene glycol	common silica column	phaseseparation	n/a	N/A	N/A	N/A	13

	us nor mal - pha se LC sep arat ion				io n						
zidovudine (AZT), LMV, didanosine (ddA) and its administered form (ddl), stavudine (d4T) and hivud (ddC)	elec troch ro mat ogr aph y	re ve rse trans cript ase inhi bitor s in a pool of endo geno us nucl eosi des		$\beta$ - cyclo dextri n- bond ed silica	U V D E T E C TI O N	N/ A	5 to 25 gm L-1	0.5 gmL-1 (ddI) and 0.9 gmL-1 (AZT)	ranged from 1.7 gmL-1 for ddl to 3.2 gmL-1 for AZT	1 4	
STA + LMV + NVP	Thin Layer Chro mato gra phy and Self - Rep orte d Ad her ence									1 5	
AZT, STA,LMV,N VP	Mic ella r Ele ctro kin etic Cap illar y Chr	caps ules	A buffer consisting of 5 mM sodium tetraborate at pH 9.8, containing 50 mM SDS, 30% methanol and 5% ethanol	c18	ph as e se pa rat ion	N/ A	N/ A	N/A	0.037, 0.051, 0.029 and 0.028 mg/L	1 6	

	om ato gra phy											
LMV, STA, tenofovir, AZT and efavirenz	mic ella r liqu id chr om ato gra phy	TAB LET FOR MU LAT ION	Four aq. micellar mobile phases buffered at pH 7 sodium dodecyl sulfate as the tensioactive, and 1- propanol or 1- pentanol as the org. modifier	C18 apola r colu mn	U V de te cti on set at 21 4 n m	1 m L m i n - 1	NO TE MP ER AT UR E CO NT RO L	0.5 and 50 pp m (r2 > 0.9 995 )	lower than 0.25 ppm	N/A	1 7	
LMV, NVP,STA	UP LC	TAB LET S	METHANOL	acqui ty x- bridg e colu mn 9(50 x4.6 mm, 3.5 um) using phos phate buffe r og ph2.2	U V de te cti on w as pe rf or m ed at 26 6 n m	1 . 0 m L / m i n - 1	30°	3.2- 19. 2, 4.4- 26. 4, and 8.8- 52. 8 µg/ min -1	N/A	N/A	1 8	
abacavir sulphate, valaciclovir hydrochloride , acyclovir sodium, ganciclovir sodium , didanosine, efavirenz, indinavir sulphate, LMV, NVP anhydrous, zalcitabine and AZT , ritonavir AND STA	mic ella r liqu id chr om ato gra phy	table ts	SDS, which was buffered with disodium hydrogenphos phate/sodium dihydrogenph osphate 10 mM at pH 7, and lastly 1- propanol, 1- butanol or 1- pentanol was added to achieve the desired concentration	n/ a	n / a	n/a	f 0- 10 µg mL -1	m 6 to 30 ng	10s criterion	1 9		

LMV,STA	Gas chromatography	liposomal formulations	n <sub>2</sub> carrier gas	BP 624 column	GC 17A Shimadzu with FID	N/A	N/A	0.9988	100 ppb	N/A	20
amivudine, STA, and NVP	micellar electrochromatography	pharmaceutical formulations	buffer consisting of pH 9.8 10 mM sodium tetraborate, 100 mM sodium dodecyl sulfate (SDS) and 15% 2-propanol.	a 75 µm i.d. uncoated fused-silica capillary	N/A	N/A	N/A	20–200, 5–50 and 25–250 µg/mL	0.6, 0.8, and 0.6 µg/mL	1.8, 2.0, and 1.6 µg/mL-1	21

“Table 3: C HPLC Single Component”

analyte	method	matrices	mobile phase	stationery phase	detection	flow rate	column	linearity	LOD	LOQ	ref
Lamivudine and Stavudine	RPLC	tablet dosage form	0.1M disodium hydrogen phosphate anhyd. buffer: methanol, adjusted to pH 5 with glacial acetic acid in the ratio of 50:50, vol./vol	REVERSED PHASE BDS HYPERSIL C18 COLUMN	UV detector at 238 nm	flow rate of 1 mL/min	N/A	good (r <sup>2</sup> > 0.999)	N/A	N/A	22

$\beta$ -stavudine	R P- H P L C	TA BL ET dosa ge for m	0.01 mmol·L-1 ammoniu m acetate soln.- acetonitril e (96.5:3.5) and 0.01 mmol·L-1 ammoniu m acetate soln.- acetonitril e (75:25) with gradient elution	SUPELC OSIL LC- 18-DB column (4.6 mm × 250 mm, 5 $\mu$ m) with Agilent 1100 liq. chromato g	U V D ET E C TI O N 25 4 N / A	2 5 ° C .	0.51-26 $\mu$ g·mL-1 (r = 1.000) for $\beta$ - stavudine, 0.13-27 $\mu$ g·mL-1 (r = 1.000) for thymine, 0.50-25 $\mu$ g·mL-1 (r = 1.000) for $\beta$ thymidine and 1.7-6.3 $\mu$ g·mL-1 (r = 1.000) for 5- O'-benzoyl- stavudine.	2. 5 · m L- 1	N/ A	23
stavudine	R P- H P L C	bulk and phar mac euti cal dosa ge for ms	0.01M ammoniu m acetate buffer:met hanol (60: 40).	Venusil XBP C- 18, 5 $\mu$ m column in iso cratic mode	uv det ect io n 26 5 n m in	1. 0 / m in	a m b l i e n t f 25-75 mcg / ml			24
STAVUDINE	R P- H P L C	bulk and caps ule dosa ge for ms	methanol: acetate buffer pH 6.0 (40:60)	A Phenomen ex Gemini C- 18, 5 $\mu$ m column having 250 × 4.6 mm i.d. in isocratic mode	U V D ET E C TI O N A T 26 5 N M	1. 0 m / N in	N / A 25-75 mcg/mL			25
stavudine	H P L C	caps ules	methanol- water (15:85)	Attach C18 column (150 mm × 4.6 mm, 5 $\mu$ m)	U V D ET E C TI O N A T 26 6 N M	N / A	N / A 9.63-77.04 $\mu$ g/mL (r = 1.0000)			26



STAVUDINE	H P L C	bulk and phar mac euti cal dosa ge for ms	acetonitril e and 0.05M potassium dihydroge n phosphate (pH 4.2) in the ratio of 50:50	RPC-18 column using an isocratic HPLC system	U V D E T E C T I O N A T 26 6 N M	1 m L/ m in	N / A	0.1-140 µg/ml	N/ A	N/ A	27
STAVUDINE	H P L C	phar mac euti cals or susp ecte d prod ucts		C18 column	U V D E T E C T I O N				N/ A	N/ A	28
METHOD A			Ammoniu m acetate buffer 0.025 M pH 4.0 AND METHAN OL	YMC pack ODS-AM, 250 × 4.6 mm, 5m	λ = 27 0 n m	1. 0 m l/ m in	a m b i e n t				
METHOD B			Potassium phosphate buffer 50 mM pH 5.65 AND acetonitril e	Symmetry C18 250 × 4.6, 5m	λ = 26 0 n m	1. 5 m l/ m in	3 0 ° C				
Zalcitabine (ddC), lamivudine (3TC), didanosine (ddI), stavudine (d4T), carbovir (CBV), zidovudine (AZT), tenofovir (PMPA) (tenofovir diisopropyl fumarate, TDF),	H P L C	PH AA RM AC EU TIC AL S	acetic acid- hydroxyla mine buffer (ionic strength 5 mM, pH 7)- acetonitril e elution gradient	Atlantis dC18 column	U V at 26 0 n m	N / A	A M B I E N T	30-10,000 ng/mL plasma concn	N/ A	30- 90 ng/ mL con cn	29
stavudine	H P L C as	PH AR MA CE UTI	water- methanol in the ratio of 90:10	C18 column	U V D E T E	N / A	N / A	25-500 µg mL-1	N/ A	N/ A	30

		SA Y M E T H O D	CA L DO SA GE FO RM			C T I O N A T 2 6 5 N M																	
																							358
STAVUDINE		R P- H P L C	TA BL ET	water- methanol (34:66)	ODS C18 column	U V D E T E C T I O N 2 6	N /	N /	8-600 µg mL- 1 (r = 0.999 9)	N/ A	N/ A											31	

"Table 4: HPLC Combined"

analyte	method	matrix	mobile phase	stationary phase	detection	flow rate	column	linearity	lod	iq	ref
LM V, STA and efavi renz	RP- HPLC	bulk and pharmaceutical formulation	0.2% Triethylamine: Acetonitrile (40:60 pH: 2.0).	reversed- phase L10 column (ZORBA X SB CN Column, 5µ, 150 mm × 4.6 mm)	uv detection at 258 nm	0 .6 mL /min	am bi ent				32

LM V, STA and Nevi rapin ein	RP- HPLC	pha rma ceut ical dos age for ms	0.1% OPA:Aceto nitrile(50:50)	Zorbax C8(250 mm × 4.6mm; 5 µm)	PDA detector at 290nm	N / A	37.5- 112.5µg/ mL (r2 = 0.999), 7.5- 22.5µg/ mL (r2 = 0.999), 50- 150µg/m L (r2 = 0.999)	0.146, 0.170 5 and 1.215	0 .4 8 6 , 0 .5 6 8 2 a n d 4 .0 4 9	33
LM V, STA	RP- HPLC	Tab let dos age for m	MeOH, acetonitrile, and 0.05 M phosphate buffer (adjusted to pH 4.5 with ortho- phosphoric acid) at a ratio of 60:20:20 vol./vol./v	C 18 column grace smart RP18 (250 × 4.6 mm, 5 µm)	ASPD detector at 254 nm	1 .0 m L / m i n/ a	10-602 2 µg/mL (r = 0.9992) for lamivudi ne and 10-60 µg/mL (r = 0.999) for stavudin e	0.16µ g/mL and 0.14µ g/mL	0 .4 9 µ g / m L a n d 0 .4 1 µ g / m L	34
STA ,LM V	RP- HPLC	pha rma ceut ical dos age for ms	HPLC grade water and methanol in the ratio of 90:10 vol./vol.	reverse phase C18, 4.5µ (250 × 4.6 mm)	UV DETEC TION 257nm	1 .0 m L / m i n A	20-100 µg/spot	3.52µ g/ml/0 .825µ g/ml	1 0 .6 1 µ g / m l / 2 .	35

									5 0 1 μ g / m l	
STA ,LM V	RP- HPLC	tabl et dos age for m by RP- HP LC	ammonium formate buffer (pH 3.5) and acetonitrile in the ratio of 15:85 vol./vol.	Phenome nex C18 column (250 × 4.6 mm id, 5 μm particle size utilizing Shimadz u HPLC LC- 10AT VP)	UV- VISIBL E AND PDA DETEC TOR- AT266N M	1 . 0 M / M N / N A	1 - 15 μg/ml for Stavudin e and 2 – 30 μg/ml for Lamivud ine	0.096 3μg/m 1,0.41 01μg/ ml	0 . 2 9 1 7 μ g / m l , 1 . 2 4 2 8 μ g / m l	36
LM V,S TA	RP- HPLC	TA BL ET DO SA GE FO RM S	methanol, acetonitrile and 0.05 M phosphate buffer (adjusted to pH 4.5 with ortho- phosphoric acid) at a ratio of 60:20:20 vol./vol./v	C 18 column grace smart RP18 (250 × 4.6 mm, 5 μm) in isocratic mode	ASPD detector at 254 nm	1 . 0 M / M N / N A	10-602 2 μg/mL (r = 0.9992) for lamivudi ne and 10-60 μg/mL (r = 0.999) for stavudin e.	0.16 μg/mL and 0.14 μg/ml	0 . 4 9 μ g / m L  a n d  0 . 4 1 μ g / m L	37

LM V,S TA	RP- HPLC	tabl et dos age for m	94: 6 v/v (buffer: acetonitrile)	hypersil ODS C18 150×4.6 mm i.d, 5µm size was fixed	270mm	N / A	N / A	37.5 to 225 ppm and 10 to 60 ppm	N/A	N / A	38
LM V,S TA	RP- HPLC	tabl et dos age for m	0.1 M disodium hydrogen phosphate anhyd. buffer: methanol, adjusted to pH 5 with glacial acetic acid in the ratio of 50:50, vol./vol	reversed phase BDS HYPERS IL C18 column	238nm	1 . 0 m l/ m i n	N / A	15- 19(µg/ml ) ,3- 18(µg/ml )	N/A	N / A	39
LM V,S TA	RP- HPLC	co mbi ned dos age for ms	acetonitrile-0.02 M ammonium acetate buffer (pH 4.5) in a gradient elution mode	Kromasil C8 (250 mm × 4.6 mm i.d.; 5 µm) column	265 nm	1 . 0 m l/ m i n	N / A	2.5-50 µg/mL for 3TC and 0.5- 10 µg/mL for d4T	0.82 µg/m L and 0.33 µg/m L	2 . 5 ( µ g / m l ) , 0 . 5 ( µ g / m l )	40
LM V,S TA	HPLC method develop ment and validatio n	bul k and co mbi ned tabl et dos age for m	Phosphate buffer pH 2.5:methanol (80:20 vol./vol.)	symmetr y C-18, Thermo (100 × 4.6), i.d, and 3.5 µm particle size in isocratic mode	UV detector dual i.e. wavelen gth of 262 nm for stavudin e and 271 nm for lamivudi ne	0 . 8 M L / M I N	N / A	25-125 µg/mL			41

LM V,S TA, NVP	RP- HPLC	co mbi ned dos age for m	mixt. of phosphate buffer:acetonitril e (pH-3.5, adjusted with 1% Glacial acetic acid) in the ratio of 85:15% vol./vol	A C8 Intersil column [4.6 (i.d.) × 250 mm, 10 µm]	266.00 nm	1 .0 M L / M N I / N A	15-75 µg/ml, 4- 20 µg/ml and 20- 100 µg/ml	15µg/ ml, 4µg/m l and 20µg/ ml	4 5 µ g / m l , 1 2 µ g / m l a n d 6 0 µ g / m l	42
(NV P), LM V and STA	Compari son of HPLC & spectrop hotomet ric methods	sing le tabl ets/ cap sule s	15mM phosphate buffer pH 7.5 and acetonitrile (80:20 v/v)	C18 column	UV detector set at 260 nm.	1 .0 M L / M I / N N	6.25(µg/ ml),12.5( µg/ml),2 5.0(µg/m l),50.0(µ g/ml)	N/A	N / A	7
LM V,S TA	RP- HPLC	TA BL ET S	methanol: 0.1% wt./vol. of ammonium acetate, adjusted to pH 3.8 with glacial acetic acid in the ratio of 15:85, vol./vol.	Sunfire C18 250 × 4.6 mm, 5 µm particle size column in isocratic mode	UV detection at 266 nm	1 .2 M L / M I / N A	75-225 µg/mL and 20- 60 µg/mL	N/A	N / A	43
LM V,S TA	RP- HPLC	tabl et dos age for m	methanol and water (80:20 vol./vol.)	reversed- phase C- 18 SYMME TRY column	UV detection at 266nm	N / n/ A a	0.999,0.9 99	0.9µg 0.8 µg	2 .8 µ g / m l a n d 2 .5 µ g / m l	44

										µg	
STAV LMV	HPLC	human plasma	10 mM acetate buffer pH 6.5 and acetonitrile	C18 column	UV detection at 265 nm			50-3,000 ng/mL for stavudine and 50-5,000 ng/mL for lamivudine			45
LAM, AZT and abacavir	RP-HPLC	tablet dosage form	0.01 M potassium dihydrogen ortho-phosphate (pH 3.0) and methanol (55:45 vol./vol.)	HiQ Sil C 18 V column	272 nm	0.8 mL/min	N/A	5-250 µg/mL for both zidovudine and abacavir and 5-140 µg/mL for lamivudine	N/A	N/A	46
STAV	HPLC	BU LK DR UG	isocratic elution with methanol/water (20:80)	LiChrospher C18 column (250 × 4.6, 5 µm)	UV detection at 266 nm			0.5-5.0 µg/mL	N/A	0.021 µg thyminine/mL and 0.134 µg	47

										thymidine / mL	
STA .LM V	RP- HPLC	co mbi ned dos age for m	acetonitrile- methanol-water at 10:1:1 ratio	isocratic HPLC with a single tunable absorban ce detector and $\mu$ - Bondapa ck C18 column	280 nm						48
STA .LM V,N VP	RP- HPLC	mul tico mp one nt pha rma ceut ical s			UV detection at 265 nm				0.1- 75.0 $\mu$ g/m L-1, 0.6- 375.0 $\mu$ g/m L-1, and 0.8- 500.0 mg/m L-1		49
LM V,S TA, NVP	HPLC	TA BL ET S	phosphate buffer pH 3.0 with acetonitrile (75:25),	LiChrosp her ODS column	UV DETEC TION AT 266NM	1 .o m l/ 3 m 0 i o n C	15-135 $\mu$ g/mL (lamivud ine), 4- 36 $\mu$ g/mL (stavudin e), and 20-180 $\mu$ g/mL (nevirapi ne)	N/A	N / A	50	



LA M,S TA, NVP	HPLC	anti retr ovir al fixe d dos e co mbi nati ons	mobile phase (A) comprising of 80% of 10 mM acetate buffer adjusted to pH 3.5 with glacial acetic acid and 20% MeOH, and mobile phase (B) comprising of 50% acetonitrile with 50% iso-Pr alc	C-18 column	UV DETEC TION 270 nm	0 . 6 M L / M N / I / N A	5-100 g ml-1, 5- 100 g ml-1, 5- 100 g ml-	N/A	N / A	51
LM V,S TA	first derivativ e spectrop hotomet ry and high perform ance liquid chromat ography	anti retr ovir al fixe d dos e co mbi nati ons						0.13g/ m	0 . 4 0 g / m	51
MET HO D 1	UV- spectrop hotomet ry				UV DETEC TION AT 280 and 300 nm		2-20 g/ml, 2- 14 g/ml			
MET HO D 2	HPLC		MeOH:water (20:80) as the mobile phase	reverse phase column	UV DETEC TION 270 nm		1-10 g/ml, 1- 10 g/ml			
LM V,S TA, NVP	RP- HPLC and ultraviol et spectrop hotomet ric method	pha rma ceut ical dos age for ms	mobile phases	reversed- phase C- 18 SYMME TRY column	uv detection		2.5-20, 2-20 and 4-30 g/ml			9
hplc meth od								0.17 ± 0.07 ,0.03 ± 0.01, 0.12 ± 0.1	0 . 5 2 ± 0 . 2 2	

									0 · 0 9 ± 0 · 0 5 0 · 3 7 ± 0 · 0 5	
uv meth od								0.19 0.29 0.25	0 · 5 8 0 · 9 2 0 · 7 6	
			Lamivudine: Methanol:water (70:30)		270 nm	0 · 7 5 M L / M I N				
			Stavudine : Methanol:water (20:80)		270 nm	0 · 8 0 M L / M I N				

			Nevirapine :Solvent A: methanol:water (20:80) 0.18 27 5 Solvent B: acetonitrile:isopr opyl alcohol (50:50)		27 nm	0 . 1 8 M L / M I N				
STA ,LM V,N VP	UV spectros copy, reverse phase HPLC, and HPTLC	tabl ets			266, 271, and 315 nm					52
uv meth od							0.8–6.4 g/ml (r = 0.996), 4– 32g/ml (r = 0.9962) and 5.33– 42.64g/m l (r = 0.9843)			
rp hplc			20 mM sodium phosphate buffer (contg. 8 mM 1- octanesulfonicac id sodium salt):acetonitrile (4:1, vol./vol.) with pH adjusted to 3.5 using phosphoric acid	C18- ODS- Hypersil (5 μm, 250 mm × 4.6 mm) column in isocratic mode			10– 50g/ml (r = 0.9968), 50– 250g/ml (r = 0.9999) and 66.6– 333.2g/m l (r = 0.9996)			

hptlc			chloroform:methanol (9:1, vol./vol.)	precoated plate of silica gel 60 F254	densitometric absorbance mode at 265 nm		10–50g/ml (r = 0.997), 50–250g/ml (r = 0.998) and 66.6–333.2g/ml (r = 0.9993)		
STA, LM, V, N, VP	rp- hplc	tablets	0.01M sodium octane sulfonate buffer (pH 2.8) and methanol (60: 40 vol./vol.)	inertsil ODS - 3V (250mm × 4.6mm, 5u)	UV detection 270 nm		7.5 µg/mL to 30.0 µg/mL, 28.0 µg/mL to 112.0 µg/mL and 37.5 µg/mL to 150.0 µg/mL		53
zalci tabine, LM, V, didanosine, STA, AZT, Abacavir	liquid chromatography		human blood plasma	new Polarity dC C18 silica column	1.0ml/min	n/a	0.781760, .078, 1.087960, .044, 1.653360, .022, 1.396060, .025, 1.228160, .033, 1.871960, .035and 2.145260, .011	n/a	100 ng/ml 54

“Table 5: HPTLC”

analyte	method	matrices	mobile phase	stationery phase	detection	flow rate	linearity	limit of detection	ref
sulfamet hoxazole and trimetho	hptlc	combination							55

prim, LMV,ST A,NVP		tabl ets								
STA,NV P	norm al- phas e HPT LC meth od	com bina tion tabl ets	ethylacetate, methanol, toluene and concd. ammonia (38.7:19.4:38.7:3.2, v:v:v)	silica gel 60F 254 plates	densito metric detecti on 254 nm		0. 28 4, 0. 27 6 an d 0. 32 7	N/ A	N / A <sup>56</sup>	
STA, LMV,N VP	UV spect rosc opy, rever se phas e HPL C, and HPT LC	TA BLE TS								52
	uv- spect rosc opy				266, 271, and 315 nm		0. 8– 6. 4 g/ m l (r = 0. 99 6) , 4– 32 g/ m l (r = 0. 99 62 ) an d 5. 33 – 42			

						.6 4g / m l (r = 0. 98 43 )			
						10 – 50 g/ m l (r = 0. 99 68 ) , 50 – 25 0g / m l (r = 0. 99 99 ) an d 66 .6 – 33 3. 2g / m l (r = 0. 99 96 )			
	RP- HPL C	20 mM sodium phosphate buffer (contg. 8 mM 1- octanesulfonicacid sodium salt):acetonitrile (4:1, vol./vol.) with pH adjusted to 3.5 using phosphoric acid		C18- ODS- Hyper sil (5 µm, 250 mm × 4.6 mm) column					

						10 – 50 g/ m l (r = 0. 99 7) , 50 – 25 0g / m l (r = 0. 99 8) an d 66 .6 – 33 3. 2g / m l (r = 0. 99 93 )			
	HPT LC		chloroform:methanol (9:1, vol./vol.)	preco ated plate of silica gel 60 F254	densito metric absorba nce mode at 265 nm				
LMV, STA	HPT LC		toluene:n hexane:tetrahydrofuran (7:1.5:1.5 vol./vol.)	Silica gel 60 F254 plate	uv detecti on at 253nm	2. 5- 7. 4u g, 0. 5- 1. 4u g			57
hptlc sin									

STA	HPT LC assay meth od	bulk drug and mul tatio n	toluene-methanol- chloroform-acetone (7.0:3.0:1.0:1.0, vol./vol./vol./vol.)	TLC silica gel 60 F 254 on alumi nium, 10- × 20-cm (cut from a 20- × 20- cm) 200- µm thickn ess colum n	Densito metric analysi s at 270 nm	30 - 10 00 ng /s po t	10 ng/ spo t	3 0 n g / p o t	58
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STA-STAVUDINE,LMV-LAMIVUDINE,NVP-NEVIRAPINE

### Hyphenation

To optimize the efficiency between both methods, experts often adopt coupled chromatographic and spectral strategies. Chromatography can then be used to isolate a pure or substantially pure fraction of a chemical component from the a mixture. If you really need to identify something precise, you could use spectroscopy, which is a technique that uses either benchmark spectra or a catalog of spectra to generate selected information. Compain et.al reported analysis of AZT, STA, zalcitabine, didanosine, LMV and abacavir using liquid chromatographic/tandem mass spectrometric assay with linearity 0.5-200 ng ml-1 is the better than other reported method

“Table 6: Hyphenation”

ana lyte	meth od	e xt ra ct io n m e d i c a l	extr acti on sol ven t	mobile phase	stationery phase	det ecti on	f l o w r a t e	I S	linearity	lod	loq	Ref
ST A	HPL C- UV	li q. - li q. e xt n	tert -Bu Me ther	10mM ammonium acetate pH 3.5, methanol, acetonitrile and glacial acetic acid	Hypercil, C18 column	UV det ect or at 26 5n m	H y d r o c h l o r o l o r i t h i z i		0.05 to 2µg/mL	N/A	N/A	59



								d e				
ST A	Liquid chromatography-tandem mass spectrometric assay	electrolytic	ethyl acetate and pH 6 phosphate buffer (50 mmol/L)	methanol-water 1% ammonia (50 : 50 : 0.5, v/v/v)	Zorbax Extend C18 column, SecurityGuard C18 guard column			$\beta$ -D-glucopyranoside	5.00-1000 ng/mL	N/A	5.00 ng/mL	<sup>58</sup>
(AZT), LMV, STA, (NVP), (EFV), (LPV) (RTV)	LC-MS/MS	liquid chromatography	Ethyl acetate	methanol/0.1% formic acid aq. soln. (80:20)	Agilent ZORBAX Eclipse XDB-C18 column	electrospray ionization (ESI) source, and multiple reaction monitoring (MRM) mode to perform detection						<sup>60</sup>
(LMV),	LC-MS/MS	protein	1.0 mL	A (0.1% formic acid in water): B	Eclipse XDB-C18 (150 mm ×	triplicate	0.5	20-3200 µg/L for 3TC,	AZT 0.25 (µg-L-	AZT 1 (µg-		<sup>61</sup>

(ST A), (A ZT) , (EF V), (N VP ) , (LP V/ RT V)		n p t n. a n d l i q. - l i q. e x t n	ethyl acet ate	(0.1% formic acid in methanol) (20:80, v/v)	4.6 mm, 5mm) analytical column	rup ole ma ss spe ctr om ete r, AP I- 32 00	M L / M I N	m i s a r t a n	d4T, AZT, 40-6400 µg/L for EFV and NVP, 62.5- 10000 µg/L for LPV, and 12.5- 2000 µg/L for RTV	1),3TC 0.5 (µg·L- 1),d4t 1.0 (µg·L- 1),EFV 1.0 (µg·L- 1),NVP 0.2 (µg·L-1), LPV 0,015 (µg·L-1), RTV 0.008 (µg·L-1)	L- 1),3 TC 2 (µg· L- 1), D4t 3 (µg· L- 1),E FV 2 (µg· L- 1),N VP 0.5 (µg· L- 1),L PV 0.05 (µg· L- 1),R TV 0.01 5 (µg· L-1)	
ST A	LC/ MS/ MS				Reverse phase micro bore column with ion pairing	mu ltip le rea cti on mo nit ori ng	0 . 3 m / m i n	2 H A T P	n/a	9.2,5.6f mol/10 6 cell and 3.1fmo;/ 10 6 cells for d4T- mp,D4t- dp, D4t- tp	0.30 pmo l,0.2 4pm ol,0. 14p mol for d4T- MP, d4T- DP ,D4t -tp	62
ST A and DN S	CE- ESI- MS/ MS					trip le qua dru pol e ele ctr osp ray (E S) MS (M					50 mg/ L (dd ATP , dAT P) to 110 mg/ L (d4T TP,	63

						S/ MS )					dTT P)	
AZ T, ST A, zal cita bin e, did ano sin e, LM V and aba cav ir	liqui d chro mato graph ic/tan dem mass spect rome tric assay	s ol id - ph as e e xt n	Me OH	two mixed solutions A and B, where A was composed of 5% MeOH and 95% ultrapure water and B was composed of 40% acetonitrile (ACN) and 60% ultrapure water	Zorbax Stable Bond C18 column followed by MS/MS anal				0.5-200 ng ml-1		LO Q for the NR TIs was set at 0.5 ng ml1 for plas ma sam ples and 0.02 5 ng per pelle t for intra cellu lar sam ples	64
(ST A), (A ZT) and (EF V)	liqui d chro mato graph y- tande m mass spect rome try			NH <sub>4</sub> OAc	Supelco LC- 18-DB (3.3 mm x 3.0 mm, 3.0 m ID) chromatogra phic column equipped with Supelco Discovery C-18 (3.0 mm) guard column (Supelco, St. Louis, MO, USA)	mu ltip le rea cti on mo nit ori ng (M R M)						65
(A ZT) (ST A)	CZE- MS/ MS			formic acidammoni a (pH 10, 50 mM ionic strength)	fused-silica capillary of 47 (or 67) cm50 m ID375 m OD	uv det ecti on	a t 2 5 4					66

(AZT) (ST A), (LMV), (DNS)	capillary electrophoresis-ionization mass spectrometry				fused-silica capillary of 37 cm× 75 μm i.d × 375 μm o.d	uv detection at 254nm				2.5 ppb for AZT and 20 ppb for d4T, 2 ppb for ddA and 5 ppb for 3TC	67
STA	liquid chromatography/tandem mass spectrometry			solvent A comprised DMH (10MM) AMMONIUM FORMATE (3MM)BPH 11.5, SOLVENT B comprised DMH (20mm) and amonium formate (6mm)/acetonitril 1:1 (v/v)	SMT-C18 O.d. 5micrometer 100 A (150x2.1mm) column	multiple reaction monitoring (MRM) mode	[2.8-8.1] - ATP			d4T-TP was 138 fmol per 7 mL blood (9.8 fmol per 106 cells)	62
STAS	capillary electrophoresis-ionization tandem mass spectrometry			0.5% formic acid added to methanol-water (95:5V/V)	fused-silica separation capillary has 70 cm350 mm I.D.3150 mmO.D. geometrical dimensions	uv detection at 254nm				2 μg l <sup>-1</sup> in a formic acid ammonia buffer (pH 2.5, 10 mM ionic strength)	68

## 2. Discussion

STA has been utilized in drug manufacturing, UV/VIS spectroscopy, and HPLC since 1976. STA insolubility makes bioanalytical or capillary electrophoretic studies tricky. Sample solution consisted MeOH and ACN. Recent advances in STA determination have been hindered by the need to upgrade sophisticated equipment to strengthen

sensitivity and tackle issues such the cost-effective use of organic solvent in sample preparation.

## 3. Conclusion

This research is aimed at spectrophotometric and spectrofluorometric chromatographic characterization of STA in both standalone and in combination with other drugs, following its

evolution and development through time. Liquid chromatography is frequently used for both solitary and combined STA analysis. Though there are established protocols for determining and managing STA levels, most procedures still do not adhere to environmentally benign principles. Therefore, efforts will be made to create biological matrices and dosage forms that limit negative impacts on the environment. As a result, less potentially harmful organic effluents are needed

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