



## Kynurenic Acid Attenuates Ischemia Reperfusion Induced Acute Kidney Injury: A Comprehensive Review

Mandeep Pundir<sup>1\*</sup>, Baldev Singh<sup>2</sup>, Kamaljeet Kaur<sup>3</sup>, Gagandeep Singh<sup>1</sup>, Deeksha Kumari<sup>1</sup>, MD Rehan<sup>1</sup>

1. Faculty of Pharmacy, Desh Bhagat University, Mandigobindgarh, Punjab.

2. Bahra Institute of Pharmacy, Patiala, Punjab

3. KC College of Pharmacy, Nawanshahr, Punjab

For Correspondence: [pundirmandeep87@gmail.com](mailto:pundirmandeep87@gmail.com)

---

### Abstract

Kynurenic acid was the first tryptophan metabolite to be isolated and characterized in mammals. It is synthesized from kynurenine which is catalysed by kynurenineaminotransferase and is found in mammalian liver, kidney, intestine, and also in rat brain. The extracellular kynurenic acid is synthesized in astrocytes and is released in the extracellular space. Kynurenic acid restores the renal functions as they have antioxidant property. Along with this they are also reported to antagonize N-Methyl D-Aspartate receptors which gets activated during the ischemia reperfusion injury thereby making the drug (Kynurenic acid) a new approach for the treatment of acute kidney injury where the major cause of renal dysfunctioning is ischemia reperfusion. Also this drug can be considered for future study keeping the very important aspect in mind that NMDA receptors are abundantly found in kidney and the drug actively work by blocking both the NMDAR glycine binding site and alpha-7-nicotine receptor.

**Keywords** Kynurenic acid, Acute Kidney Injury, NMDA, tryptophan, Ischemia Reperfusion Injury

---

### Introduction

Acute kidney injury (AKI) is defined as an abrupt deterioration in renal functioning typified by decreased glomerular filtration rate (GFR) and tubular dysfunction. (Bellomo *et al.*, 2004). According to the Acute Kidney Injury Network, AKI is defined as a decrease in kidney function that is accompanied by a rise in serum creatinine of more than 0.3 mg/dL or 50% in just 48 hours. (Mehta *et al.*, 2007). AKI is more common in elderly people, those with diabetes, hypertension, vascular disease, and those who already have renal impairment. AKI accounts for about 1% of hospital admissions, of which more than 7% worsen to cause further harm. AKI's long-term effects include the accelerated onset of end-stage renal disease and chronic kidney disease. AKI is most frequently caused by sepsis and shock in intensive care units. The mortality rate from sepsis is 20–50%, and it might reach 60% when there are hemodynamic changes. (Uchino *et al.*, 2005).

## Causes of AKI

AKI has three general categories of causes: pre renal, intrinsic renal, and post renal. The pre renal cause, which accounts for 40–70% of renal disorders, mostly includes decreased blood supply to the kidneys. (Hou *et al.*, 1983; Kaufman *et al.*, 1991). In 10 to 50 percent of kidney injury cases, the intrinsic renal failure is brought on by the direct renal parenchymal damage. (Kaufman *et al.*, 1991; LianoandPascual, 1996). 10% of renal patients have blocked urine flow as the post-renal cause of AKI. (LianoandPascual, 1996).

### ***Pre renal***

As pre and post glomerular arteriolar resistance fluctuates over a large range of mean arterial pressure, the renal blood flow and GFR remain essentially constant. The pre-glomerular arteriolar vasodilation caused by prostaglandins and nitric oxide and the post-glomerular arteriolar vasoconstriction caused by angiotensin I are the two key factors in renal autoregulation. AKI results from any perturbation in this equilibrium. The main pre-renal causes of AKI are atherosclerotic cardiovascular disease, renal hypoperfusion brought on by volume depletion, hypotension, or renal artery stenosis.(Hilton, 2006).

### ***Intrinsic renal***

Diseases that impact the glomeruli, tubules, vasculature, or interstitium are the primary causes of intrinsic acute renal failure. In sepsis, which is accompanied by many organ damage, it is frequently complex and prevalent. (Mehta *et al.*, 2004). Nephrotoxins, heavy metals, crystallisation of uric acid and oxalate, and deposition of these substances are additional significant causes of intrinsic renal failure.

### ***Post renal***

Renal stones, issues with the bladder, and ureteral obstruction are the main causes of post renal failure. (Hilton, 2006). The main causes of renal failure are listed in table 1.

**Table 1. Various causes of AKI**

- 
- **Pre renal**
    - Hypotension
    - Reduced effective circulating volume
    - Renal artery stenosis
    - Drugs: angiotensin converting enzyme inhibitors, selective cyclo-oxygenase 2 inhibitors
  - **Intrinsic**
    - Glomerulonephritis
    - Systemic lupus erythematosus
    - Thrombotic disease
    - Ischemic acute tubular necrosis (ATN)
    - Nephrotoxic ATN

- Myeloma cast nephropathy
- Drugs: aminoglycosides, radio-contrast media, heavy metals
- Uric acid and oxalate crystal deposition

➤ **Post renal**

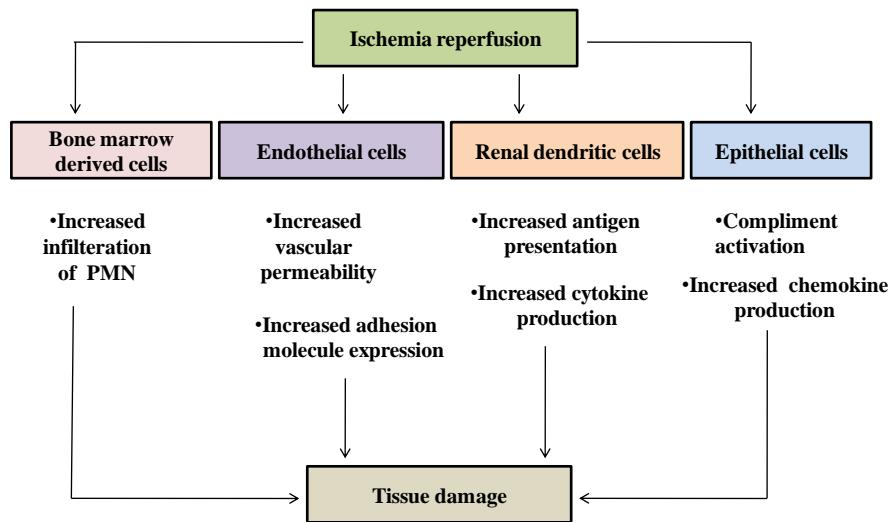
- Renal papillary necrosis
  - Kidney stones
  - Bladder tumor
  - Carcinoma of cervix
  - Prostatic hypertrophy
  - Ureteral obstructions
  - Pelvic malignancy
- 

### **Renal ischemia reperfusion injury**

Ischemia reperfusion injury (IRI) is a condition of decreased metabolite washout and tissue oxygen deprivation. The restoration of blood flow to the ischemic tissue is known as reperfusion. (Star, 1998). Even while tissues are receiving new nutrients and oxygen, reperfusion itself causes serious damage.(Bonventre, 1993). The renal IRI is seen in diseases such hydronephrosis, partial nephrectomy, cardiopulmonary bypass, sepsis, and renal transplantation, all of which are linked with severe morbidity and death. (Desai *et al.*, 2005; Harnendezet *et al.*, 2008). Vasoconstriction, tubular and glomerular damage, as well as other processes, are all part of the complicated process known as renal IRI.(Bird *et al.*, 1988). As a result of the production of various reactive oxygen species (ROS) by renal ischemia, including superoxide radicals ( $O_2\cdot$ ), hydrogen peroxide, and hydroxyl radicals ( $OH\cdot$ ), neutrophils accumulate and lytic enzymes are released, resulting in increased oxidative stress and tissue damage. (Paller *et al.*, 1984; Bonventre, 1993). The ROS are involved in intracellular and intercellular communication under normal circumstances.(Nose, 2000). In renal IRI, the  $O_2$  ion released during IRI combines with NO to create peroxynitrite (ONOO), which causes cellular damage by causing DNA strands to break and protein tyrosine residues to nitrate. The ONOO also produces OH, which is extremely harmful and worsens the cellular injury.(Beckman *et al.*, 1990). Additionally, by nitrating and deactivating antioxidant enzymes like superoxide dismutase, ONOO increases the kidney damage. (MacMillan-Crowet *et al.*, 1996). Additionally, oxidative stress decreases the bioavailability of  $NO\cdot$ , which causes vasoconstriction and microvascular thrombosis. (Gryglewskiet *et al.*, 1986). This is clear from the fact that during the early stages of renal IRI,  $O_2$  increases prior to inducible nitric oxide synthase (iNOS) producing large concentrations of NO. (Araujo and Welch, 2006). Given that ONOO $\cdot$  scavenger ebselen has been shown to offer significant protection in models of ischemia renal failure, it has been established that ONOO $\cdot$  plays a key role in renal IRI. (Noiri *et al.*, 2001). The production of ROS during reperfusion is linked to tissue injury. Lipid peroxidation is accelerated by the produced peroxy radicals, which take hydrogen atoms from lipids and cause a chain reaction.(Noguchi and Niki, 1998). The resulting lipid peroxides change membrane permeability and interfere with cellular processes

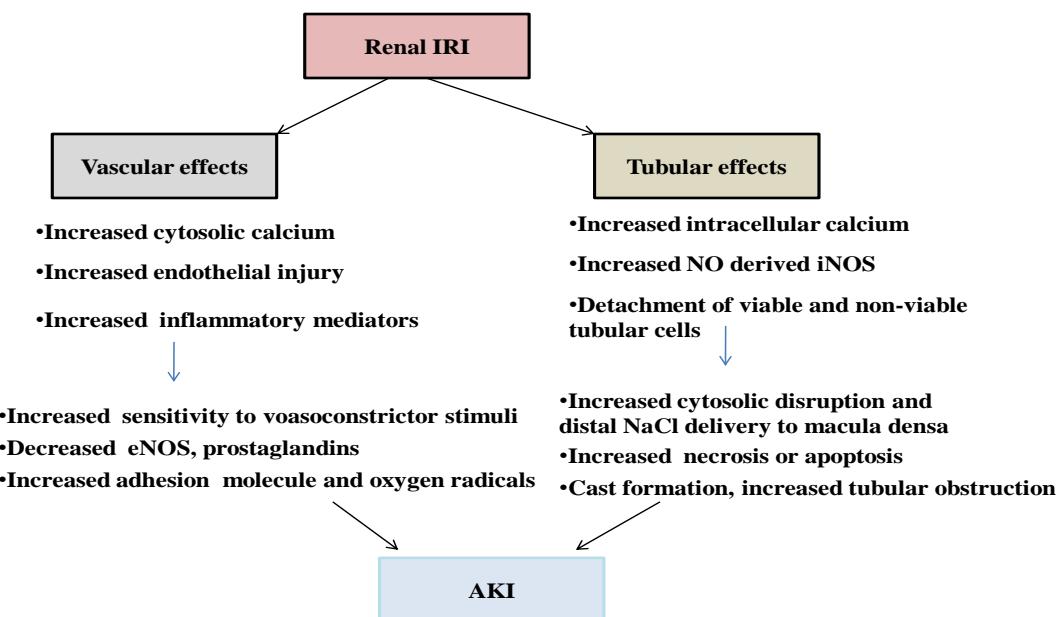
like membrane-bound ion pumps. Advanced glycosylation end products and extracellular signal-regulated kinase (ERK) are involved in the renal IRI-induced oxidative stress, which causes tissue damage.(Bas *et al.*, 2009; Mansouri *et al.*, 2011).An imbalance between endothelial NOS (eNOS) and iNOS expression in renal IRI, which increases iNOS and significantly contributes to the pathogenesis of renal IRI, occurs.(Goligorsky *et al.*, 2002, 2004).Apoptosis is caused by the activation of caspase-3 and mitogen-activated protein kinase by the enzyme nicotinamide adenine dinucleotide phosphate oxidase. Additionally, profibrotic factors including nuclear factor kappa B and transforming growth factor- activated by renal IRI promote the production of monocyte chemoattractant protein-1 (MCP-1) and interleukin-8 (IL-8). (Morigi *et al.*, 2002; Tang *et al.*, 2003).MCP-1 is a powerful inducer of the recruitment of macrophages, which is thought to increase oxidative stress and have a role in the inflammation of the tubulointerstitial tissue.(Chow *et al.*, 2007).

In the renal IRI, physiologically active mediators such bradykinin, histamine, platelet activating factor, and pro-inflammatory cytokines like IL-1,6 and tumour necrosis factor (TNF-) are produced and released.(Couser, 1998; Chatterjee *et al.*, 1999; Dahaandvan Kooten, 2000;Thurman, 2007).Because dialysis significantly benefits from removing inflammatory mediators from plasma, their significance in renal inflammation has been demonstrated. (InthornandHoffmann, 1996; Montoliu, 1997).Additionally contributing to renal inflammation are vasoactive substances like NO and arachidonic acid metabolites such cysteinyl leukotrienes. (Lieberthal, 1998; Patel *et al.*, 2004).Also, pro-inflammatory proteins including vascular cell adhesion molecule-1 (VCAM-1) and intracellular adhesion molecule-1 (ICAM-1) are explicitly involved in renal IRI. (Muller *et al.*, 1996; Molitoris and Marrs, 1999;Burne-Taney and Rabb, 2003).Through a highly specialised transport system that uses intracellular energy, the proximal tubular cells enable the movement of ions, water, and chemicals through the cell layer. This energy is disrupted during renal IRI, which causes adenosine triphosphate (ATP) levels to drop. This dysfunction of the Na<sup>+</sup>K<sup>+</sup>-ATPase pump causes cell enlargement, intracellular disturbance, and ultimately cell death.(LieberthalandLevine, 1996;PadanilamandLewington,1999; Sheridan and Bonventre, 2000).Endothelial damage causes increased production of cell adhesion molecules such ICAM-1 on damaged endothelial cells, which further obstructs capillary and post capillary venules, resulting in endothelial damage and increasing microvascular permeability. (Friedewald and Rabb, 2004).Finally, many IRI-related events that resulted in tissue injury are outlined. in figure 1.

**Figure 1. Mechanisms of IRI induced tissue damage.**

### Mechanisms of AKI

AKI is caused by a variety of vascular and tubular causes. The vascular factor comprises endothelial damage, leukocyte infiltration, and inflammatory mediators as well as renal vasoconstriction. Renal ischemia raises endothelins, which contribute to vasoconstriction and a lower GFR, while decreasing eNOS and vasodilatory prostaglandins. Inhibition of these culprits has proven protection against AKI, and overexpression of adhesion molecules has been observed in the outer medullary congestion. The tubular factors include tubuloglomerular feedback that is increased, tubular cast development, tubular blockage, and back-leak of glomerular filtrate.(Schrieret al., 2004). Additionally, the excretion of epithelial tubule cells and proximal tubule brush border membranes into urine (Thadhani et al., 1996). Reduced GFR results from the cast formation of viable and non-viable tubular cells, apoptotic, viable, and necrotic tubule epithelial cells, brush boundary membranes, and extracellular matrix, including fibronectin. (Kribbenet al., 1999). During renal damage, it is seen that the caspase, caspase-1 (an inflammatory caspase), and caspase-3 (an executioner caspase) cysteine protease pathways are activated. Both a rise in cytosolic calcium and a decline in calpastatin activate calpain. The inflammatory cytokine IL-8 is then stimulated by calpain's activation of caspase-1, and caspase-3 cleaves calpastatin to induce apoptosis. Since proximal sodium reabsorption is hampered and distal sodium chloride supply to the macula densa is ultimately increased by the intracellular rise in calcium, this further reduces GFR. (Schrieret al., 2004). Figure 2 provides a summary of the numerous tubular and vascular variables that contribute to AKI.

**Figure 2.**Mechanisms involved in IRI induced AKI.

### N-Methyl-D-Aspartate Receptors (NMDAR)

The most sensitive agonist for the NMDAR class of ionotropic glutamate receptors is NMDA.(Hollmann and Heinemann, 1994).The only receptors that have an agonist site that is responsive to both glutamate and glycine are NMDARs. (Johnson and Ascher, 1987).When NMDAR is activated, an ion channel opens, allowing calcium ions to enter the cell and act as a second messenger, changing various brain functions as synaptic plasticity, excitotoxicity, and memory. Subunits of the NMDAR include NR1, NR2A, NR2B, NR2C, NR3A, and NR3B.(Avshalumov and Rice, 2002; Leunig et al., 2004; Isaac et al., 2007).Among these, NR2A and NR2B are discovered to differ from NR1 and NR3B due to their extended C-terminal lengths. Along with one or two NR2 or NR3 subunits, the NR1 subunit is a constant component of the NMDAR. Glycine is bound to the NR1 subunit of NMDAR, while glutamate is bound to the NR2 subunit. (Laube et al., 1997; Anson et al., 1998).Multiple indicators point to the presence of two NR1 and two NR2 subunits in the NMDAR complex. (Laube et al., 1998; Sobolevsky et al., 2009).Glycine binding modules are created when the NR3 subunit and NR1 subunit form a complex. (Chatterton et al., 2002).Four homologous pore-forming subunits combine to produce the tetrameric NMDAR complex. The subunits are made up of the C-terminal domain (CTD), the transmembrane domain (TMD), the agonist binding domain (ABD), and the vast extracellular N-terminal domain (NTD). The ABD permits glutamate or glycine to bind, the NTD takes part in subtype-specific assembly and regulation, and the TMD also has a re-entrant loop that filters the ion selectivity. The Y-shaped structure of the receptor is revealed by the crystal structure, with

TMD at the base, NTD at the top, and ABD in the middle.(Mayer, 2011).NMDAR activity is inhibited by the NR3A and B subunit. The body has multiple NMDAR isoforms, and the functional characteristics are determined by the differential expression of NR2 subunits and the splicing of NR1 transcripts. The membrane domain is in charge of the receptor's high calcium permeability and unitary conductance. Each subunit contains a cytoplasmic domain in addition. (Sobolevsky *et al.*, 2009).

### Functions of NMDAR

The NMDAR performs a variety of neurophysiological functions in the body. Strong depolarization and glutamate release are necessary for NMDAR to function synaptically, enabling the integration and decoding of incoming synaptic activity. (Seeburget *et al.*, 1995).High calcium ion influx allows the receptor to translate the synaptic input pattern into a long-lasting change in synaptic strength. (Mayer *et al.*, 1984; Nowak *et al.*, 1984).According to the long-term potentiation (LTP) phenomenon, which Bliss and Lomo first described in 1973, an intense burst of synaptic input causes excitatory synapses to become stronger over time. (Bliss and Lomo, 1973).The NMDAR plays a crucial part in the hippocampus area's production of LTP. Along with NMDAR, which creates the action potential, voltage-gated sodium and calcium channels also play a role in the active conductance of dendrites. (Spruston *et al.*, 1995;Stuart *et al.*, 1997).NMDARs are investigated for their function in ischemic brain injury in addition to their roles in cognition and memory. (Rothman and Olney, 1986).On a chronic model of inflammation and neuropathic pain, NMDAR antagonist has anti-nociceptive effects. (Quet *et al.*, 2009).In monoarthritic rats, NMDAR in the spinal cord modifies the expression of NOS isoforms.(Infante *et al.*, 2007).Additionally, glutamate-mediated protection involves inducing COX-2, which reduces neuronal mortality brought on by excitotoxicity, leading to an upregulation of COX-2 expression in neurons via the glutamatergic route. (Strauss and Marini, 2002).For many neurological conditions, including stroke, hypoxia, ischemia, head trauma, Huntington's, Parkinson's, and Alzheimer's diseases, epilepsy, neuropathic pain, mood disorders, alcoholism, and schizophrenia, the NMDAR is a significant therapeutic target. (Quet *et al.*, 2009; Dhanwan *et al.*, 2011; Gonda, 2012; Moojen *et al.*, 2012; Tayebet *et al.*, 2012). Additionally, they have had limited clinical success due to their poor efficacy and undesirable side effects, which include catatonia, ataxia, nightmares, hallucinations, and memory problems.

### Pharmacology of NMDAR

Glutamate functions as a powerful NMDA agonist at the NMDAR's particular binding site. Another such agonist that binds selectively to NMDARs is homoquinolinate, a derivative of the endogenous agonist quinolate. (Grimwood *et al.*, 2002).L-aspartate and sulfur-containing amino acids like homocysteate, in addition to glutamate and quinolate, act as agonists at NMDAR. (Hollmann and Heinemann, 1994; Kew and Kemp, 2005).Numerous substances operate at the glycine binding site and are NMDAR agonists. One of the most powerful agonists of the glycine binding site is D-serine produced from neurons.(Kartvelishvily *et al.*, 2006; Panatier *et al.*, 2006).One such substance, 1-aminocyclopropane carboxylic acid, functions as a high affinity full

agonist and a low affinity competitive antagonist, respectively, at the glycine and glutamate sites. (Nahum-Levy *et al.*, 1999). In astrocytes, kynurenic acid is produced endogenously and is thought to regulate NMDAR in vivo. (Poeggeler *et al.*, 2007). The kynurenic acid is an antagonist that competes with the NMDAR and nicotinic alpha-7-acetylcholine receptor for the glycine site. (Erhard *et al.*, 2009).

### **Role of NMDAR in peripheral organs including kidney**

NMDAR has been found in a variety of peripheral organs, including cells from the pancreas, the urogenital tract, the microvasculature, and even lymphocytes and megakaryocytes. (Yang *et al.*, 2008). Additionally involved in gastric motility, the NMDAR is expressed by both osteoblasts and osteoclasts. (Chen *et al.*, 1998; Watanabe *et al.*, 2008). Additionally, NMDAR receptors are found on taste buds, and blocking them increases hunger and food intake. (Dingledine and Conn, 2000; Covasa *et al.*, 2003). They are crucial to the growth of the embryo. Additionally, NMDAR is expressed in the lung tissues, and when activated, it worsens lung damage. (Shen *et al.*, 2010).

In addition to other peripheral organs, the kidney's existence of NMDAR is widely known. (Ma *et al.*, 2002; Tanaka *et al.*, 2004; de Araujo *et al.*, 2005). The glomeruli, brush boundary membrane, and outer medulla of the nephron all contain the NMDAR. In the brain, the nNOS colocalizes with NMDAR and is notably expressed in renal macula densa cells. (Deng *et al.*, 2002). NMDAR agonists and antagonists are both thought to protect the kidneys from IRI-induced AKI in rats. The NMDAR agonist's protective impact is achieved by its vasodilatory effect in response to constitutive NOS, which mediates vasodilation. (Deng *et al.*, 2002). In IRI-induced AKI, the NMDAR antagonist is also said to be renoprotective. (Yang *et al.*, 2008). Calcium influx and overload, increased ROS production, and COX-2 overexpression are all caused by NMDAR activation. (Biber *et al.*, 1981; Choi, 1992; Monyer *et al.*, 1992; Conn and Pin, 1997; Tian *et al.*, 2008). Additionally, NMDAR activation uses the endothelin-NO route to cause cell damage. (Leung *et al.*, 2004).

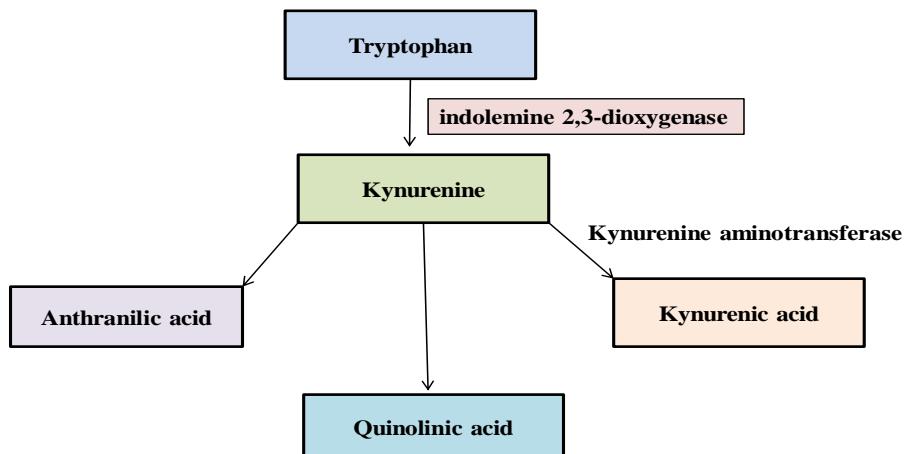
### **Various target sites of NMDAR**

The NMDAR has a variety of target sites, including sites for glutamate binding and sites for glycine and polyamine modulation. It is widely known that binding of glycine and glutamate results in calcium influx and channel opening. A non-essential amino acid is glutamate. Glutamate is the name for the carboxylate anion and the salts of glutamic acid. It is an essential component of cellular metabolism and is crucial for the body's removal of extra or waste nitrogen. It is among the most prevalent excitatory neurotransmitters in vertebrates' nervous systems. The significance of glutamate receptors in synaptic plasticity demonstrates its importance for cognitive processes in the brain, including memory and learning. At glutamatergic synapses in the hippocampus, neocortex, and other regions of the brain, the LTP is a type of plasticity. Excitotoxicity brought on by glutamate is a result of the ischemic cascade and is linked to stroke, amyotrophic lateral sclerosis, lathyrism, autism, mental retardation, and

Alzheimer's disease.(Hyndet *al.*, 2004; Sapolsky, 2005). Additionally, it lessens the antinociception that morphine causes and is known to cause hyperalgesia.(Srivastava *et al.*, 1995). With the arrival of calcium, glutamate or NMDA binding causes the channel to open. The glutamate binding site's competitive antagonists, 2-amino-5-phosphopentanoate (AP-5) and 2-amino-7-phosphonoheptanoic acid (AP-7) work to counteract glutamate's effects on NMDARs. (van den Boset *al.*, 1992; Abizaidet *al.*, 2006).

### Kynurenic Acid as a treatment drug

The first tryptophan metabolite to be identified and characterised in animals was kynurenic acid. (Ellinger, 1904; Homer, 1914). It is extensively researched in mammalian liver, kidney, gut, as well as rat brain. It is synthesised from kynurenine, which is catalysed by kynurenine aminotransferase (figure 3).(Minatogawa *et al.*, 1974; Okuno *et al.*, 1980).



**Figure 3.**Synthesis of kynurenic acid.

Astrocytes produce extracellular kynurenic acid, which is then discharged into the extracellular space.(Poeggeler *et al.*, 2007). Poor blood-brain barrier translocation from peripheral to central nervous system (Swartzet *al.*, 1990; Fukuiet *al.*, 1991). However, by irreversible transmission of kynurenine, its immediate precursor, kynurene, can successfully penetrate the blood-brain barrier to create a dose-dependent increase in kynurenic acid content. (Robotka *et al.*, 2008). Due to its ability to block both the alpha-7-nicotine receptor and the NMDAR glycine binding site, kynurenic acid functions as a neuroprotectant.(Hilmaset *al.*, 2001). Additionally, it reduces the release of TNF- from mononuclear cells treated with lipopolysaccharide and functions as a chemokine by regulating leukocyte endothelial contacts in the vasculature under

physiologically relevant circumstances.(Wang *et al.*, 2006).Because of the antagonistic interaction between kynurenic acid and the glutamate system in the amygdale, kynurenic acid has an effect on the epileptogenic process.(Szyndleret *al.*, 2012).It has been discovered to be crucial in regulating brain plasticity and cognition.(Potter *et al.*, 2010).Kynurenic acid treatment intracerebrally or systemically reduces quinolinic acid-induced seizures in mice and rats. (Lapin, 1976,1980).It lessens the rat brain's excitotoxic neuronal damage. (Foster *et al.*, 1984).The kynurenic acid produced by glial cells has a high rate of turnover and greatly builds up with ageing. (Moroniet *al.*, 1988).In the rat locus coeruleum, extracellular kynurenic acid concentration increases have behavioural effects such as decreased locomotor activity, mild analgesia, seizure control, and the protection of excitotoxic neuronal damage. (Carpenedoet *al.*, 1994; Nemeth *et al.*, 2004).In both in vitro and in vivo models of focal or generalised ischemia, kynurenic acid has been shown to lessen post-ischemic brain damage. (Cozziet *al.*, 1999; Carpenedoet *al.*, 2002).Additionally, it lessens neurodegeneration in various transgenic models of the diseases Huntington and Alzheimer. (Zwillingetal., 2011).The kynurenic acid reduces the number of fatalities in the septic shock model and possesses direct antioxidant properties. (Lugohuitronet *al.*, 2011).

### Future Prospect of Kynurenic Acid

It is seen that increased urinary glycolysis intermediates and alters purine and tryptophan catabolism in mice with histopathological AKI. These pathways are potential diagnostic and therapeutic targets for postoperative AKI in this high-risk cohort(Davidson, J.A et al., 2022). Modest to severe AKI following newborn heart surgery is related to changes in the blood metabolome, particularly major changes in the purine, methionine, and kynurenine/nicotinamide metabolism. Hence there can be scope for metabolic investigations in the evaluation of lower-stage injury since some infants with moderate AKI showed comparable metabolic changes(Davidson, J.A et al., 2021).Animal research and human observational studies of AKI and CKD have both revealed dysregulated Kynurenic pathway (KP) metabolites. The magnitude and direction of modifications in the KP depend on the aetiology of the injury in AKI. From the onset of the disease to its advanced stages, KP metabolites are altered in CKD, including uremia and its related vascular effects. The activation of the KP and redirection to certain sub-branches are currently being studied for the treatment of various illnesses, especially in light of the immunomodulatory capabilities of some KP metabolites. Further knowledge of the KP may aid in the development of biomarkers and specialised treatments for specific kidney diseases (Wee, H.N et al., 2021)

### Conclusion

The KYN pathway is a promising target in kidney disease prevention and treatment. Although many questions remain to be answered, future studies should explicitly explain the role of the KYN pathway in the pathogenesis of renal disorders, especially AKI. Searching for novel agents

modulating KYN pathway activity may contribute to the introduction of new drugs for kidney diseases and significantly improve patient prognosis

## References

- Abizaid, A., Liu, Z., Andrews, Z., Shanabrough, M., Borok, E., Elsworth, J., Roth, R., Sleeman, M., Picciotto, M., Tschop, M., Gao, X. and Horvath, T. (2006). Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *The Journal of Clinical Investigation* **116**:3229-39.
- Aebi, H., Wyss, S.R., Scherze, B. and Skvaril, F. (1974). Heterogeneity of erythrocyte catalase II. Isolation and characterization of normal and variant erythrocyte catalase and their subunit. *European Journal of Biochemistry* **17**:307-18.
- Agus, Z.S. (1999). Hypomagnesemia. *Journal of the American Society of Nephrology* **10**:1616-22.
- Annetta, M.G., Iemma, D., Garisto, C., Tafani, C. and Proietti, R. (2005). Ketamine: new indications for an old drug. *Current Drug Targets* **6**:789-94.
- Anson, L.C., Chen, P.E., Wyllie, D.J., Colquhoun, D. and Schoepfer, R. (1998). Identification of amino acid residues of the NR2A subunit that control glutamate potency in recombinant NR1/NR2A NMDARs. *The Journal of Neuroscience* **18**:581-89.
- Araujo, M. and Welch, W.J. (2006). Oxidative stress and nitric oxide in kidney function. *Current Opinion in Nephrology and Hypertension* **15**:72-77.
- Avshalumov, M.V. and Rice, M.E. (2002). NMDA receptor activation mediates hydrogen peroxide-induced pathophysiology in rat hippocampal slices. *Journal of Neurophysiology* **87**:2896-903.
- Barbosa, F.T., Barbosa, L.T., Jucá, M.J. and Cunha, R.M. (2010). Applications of magnesium sulfate in obstetrics and anesthesia. *Revista Brasileira De Anestesiologia* **60**:104-10.
- Bas, M., Tugcu, V., Kemahli, E., Ozbek, E., Uhri, M., Altug, T. and Tasci, A.I. (2009). Curcumin prevents shock-wave lithotripsy-induced renal injury through inhibition of nuclear factor kappa-B and inducible nitric oxide synthase activity in rats. *Urological Research* **37**:159-64.
- Beckman, J.S., Beckman, T.W., Chen, J., Marshalland, P.A. and Freeman, B.A. (1990). Apparent hydroxyl radical production by peroxy nitrite: implications for endothelial injury from nitric oxide and superoxide. *Proceedings of the National Academy of Sciences USA* **87**:1620-24.
- Bellé, N.A., Dalmolin, G.D., Fonini, G., Rubin, M.A. and Rocha, J.B. (2004). Polyamines reduces lipid peroxidation induced by different pro-oxidant agents. *Brain Research* **1008**:245-51.
- Bellomo, R., Ronco, C., Kellum, J.A., Mehta, R.L. and Palevsky P. (2004). Acute Dialysis Quality Initiative workgroup. Acute renal failure-definition, outcome measures, animal models, fluid therapy and information technology needs: the Second International Consensus Conference of the Acute Dialysis Quality Initiative(ADQI) Group. *Critical Care* **8**:R204-12.

- Benveniste, M. and Mayer, M.L. (1993). Multiple effects of spermine on *N*-methyl-d-aspartic acid receptor responses of rat cultured hippocampal neurones. *The Journal of Physiology* **464**:131-63.
- Berlese, D.B., Sauzem, P.D., Carati, M.C., Guerra, G.P., Stiegemeier, J.A., Mello, C.F. and Rubin, M.A. (2005). Time-dependent modulation of inhibitory avoidance memory by spermidine in rats. *Neurobiology of Learning and Memory* **83**:48-53.
- Beutler, E., Duron, O. and Kelly, B.M. (1963). Improved method for the determination of blood glutathione. *The Journal of Laboratory and Clinical Medicine* **61**:882-88.
- Biber, J., Stieger, B., Haase, W. and Murer, H. (1981). A high yield preparation for rat kidney brush border membranes. Different behaviour of lysosomal markers. *Biochimie et Biophysica Acta* **647**:169-76.
- Bird, J.E., Milhoan, K. and Wilson, C.B. (1988). Ischemic acute renal failure and antioxidant therapy in the rat: the relaxation between glomerular and tubular dysfunction. *The Journal of Clinical Investigation* **81**:1630-38.
- Bliss, T.V.P. and Lomo, T. (1973). Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *The Journal of Physiology* **232**:331-56.
- Bonventre, J.V. (1993). Mechanism of ischemic acute renal failure. *Kidney International* **43**: 1160-78.
- Bradley, P.P., Priebat, D.A., Christensen, R.D. and Rothstein, G. (1982). Measurement of cutaneous inflammation: estimation of neutrophil content with an enzyme marker. *Journal of Investigative Dermatology* **78**:206-09.
- Burne-Taney, M.J. and Rabb, H. (2003). The role of adhesion molecules and T cells in ischemic renal injury. *Current Opinion in Nephrology and Hypertension* **12**:85-90.
- Campos, S.B., Silva, J.C. and Seguro, A.C. (2001). Hypomagnesemia potentiates postischemic acute renal failure. *Journal of the American Society of Nephrology* **12**:A4057.
- Carpenedo, R., Chiarugi, A., Russi, P., Lombardi, G., Carla, V., Pellicciari, R., Mattoli, L. and Moroni, F. (1994). Inhibitors of kynurenine hydroxylase and kynureinase increase cerebral formation of kynurenic acid and have sedative and anticonvulsant activities. *Neuroscience* **61**:237-44.
- Carpenedo, R., Meli, E., Peruginelli, F., Pellegrini-Giampietro, D.E. and Moroni, F. (2002). Kynurenine 3-mono-oxygenase inhibitors attenuate post-ischemic neuronal death in organotypic hippocampal slice cultures. *Journal of Neurochemistry* **82**:1465-71.
- Carvalho, F.B., Mello, C.F., Marisco, P.C., Tonello, R., Girardi, B.A., Ferreira, J., Oliveira, M.S. and Rubin, M.A. (2012). Spermidine decreases  $\text{Na}^+ \text{K}^+$ ATPase activity through NMDA receptor and protein kinase G activation in the hippocampus of rats. *European Journal of Pharmacology* **684**:79-86.
- Cebere, A., Cebers, G., Wägner, A. and Liljequist, S. (2002). Spermidine attenuates the inhibitory effect of ethanol on NMDA-induced neurotoxicity. *Naunyn Schmiedebergs Archives of Pharmacology* **366**:117-22.

- Cerda, J., Bagga, A., Kehr, V. and Chakravarthi, R.M. (2008). The contrasting characteristics of acute kidney injury in developed and developing countries. *Nature Clinical Practice Nephrology* **4**:138-53.
- Chatterjee, P.K., Hawksworth, G.M. and McLay, J.S. (1999). Cytokine stimulated nitric oxide production in human renal proximal tubule and its modulation by natriuretic peptides: a novel immunomodulatory mechanism? *Experimental Nephrology* **7**:438-48.
- Chatterjee, P.K., Zacharowski, K., Cuzzocrea, S., Otto, M. and Thiemermann, C. (2000). Inhibitors of poly (ADP-ribose) synthetase reduce ischemia-reperfusion injury in the anesthetized rat *in vivo*. *The Journal of the Federation of American Societies for Experimental Biology Journal* **14**:641-51.
- Chatterton, J.E., Wobuluyi, M., Premkumar, L.S., Takahashi, H., Talantova, M., Shin, Y., Cui, J., Tu, S., Sevarino, K.A., Nakanishi, N., Tong, G., Lipton, S.A. and Zhang, D. (2002). Excitatory glycine receptors containing the NR3 family of NMDAR subunits. *Nature* **415**:793-98.
- Chenu, C., Serre, C.M., Raynal, C., Burt-Pichat, B. and Delmas, P.D. (1998). Glutamate receptors are expressed by bone cells and are involved in bone resorption. *Bone* **22**:295-99.
- Choi, D.W. (1992). Excitotoxic cell death. *Journal of Neurobiology* **23**:1261-76.
- Chow, F.Y., Nikolic-Paterson, D.J., Ma, F.Y., Ozols, E., Rollins, B.J. and Tesch, G.H. (2007). Monocyte chemoattractant protein-1-induced tissue inflammation is critical for the development of renal injury but not type 2 diabetes in obese db/db mice. *Diabetologia* **50**:471-80.
- Clayden, E.C. (1971). *Practical section cutting and staining*. Churchill Livingstone, London. 115.
- Coca, S.G., Yusuf, B., Shlipak, M.G., Garg, A.X. and Parikh, C.R. (2009). Long-term risk of mortality and other adverse outcomes after acute kidney injury: a systematic review and meta-analysis. *American Journal of Kidney Diseases* **53**:961-73.
- Coderre, T.J. and Van Empel, I. (1994). The utility of excitatory amino acid (EAA) antagonists as analgesic agents. I. Comparison of the antinociceptive activity of various classes of EAA antagonists in mechanical, thermal and chemical nociceptive tests. *Pain* **59**:345-52.
- Conn, P.J. and Pin, J.P. (1997). Pharmacology and functions of metabotropic glutamate receptors. *Annual Review of Pharmacology and Toxicology* **37**:205-37.
- Cotman, C.W. and Iversen, L.L. (1987). Excitatory amino acids in the brain: focus on NMDA receptors. *Trends in Neurosciences* **10**:263-65.
- Couser, W.G. (1998). Pathogenesis of glomerular damage in glomerulonephritis. *Nephrology Dialysis Transplantation* **13**:10-15.
- Covasa, M., Ritter, R.C. and Burns, G.A. (2003). Cholinergic neurotransmission participates in increased food intake induced by NMDA receptor blockade. *American Journal of Physiology Regular Integrative and Comparative Physiology* **285**:641-48.
- Cozzi, A., Carpenedo, R. and Moroni, F. (1999). Kynurenone hydroxylase inhibitors reduce ischemic brain damage: studies with (m-nitrobenzoyl)-alanine (mNBA) and 3, 4-

- dimethoxy-[*N*-4-(nitrophenyl) thiazol-2yl]-benzenesulfonamide (Ro 61–8048) in models of focal or global brain ischemia. *Journal of Cerebral Blood Flow and Metabolism* **19**:771-77.
- Crespi, F., Lazzarini, C., Andreoli, M. and Vecchiato, E. (2000). Voltametric and functional evidence that *N*-methyl-D-aspartate and substance P mediated rat vascular relaxation via nitrogen monoxide release. *Neuroscience Letters* **287**:219-22.
- Cromhout, A. (2003). Ketamine: its use in emergency department. *Emergency Medicine* **15**:155-59.
- Daha, M.R. and van Kooten, C. (2000). Is the proximal tubular cell a proinflammatory cell? *Nephrology Dialysis Transplantation* **15**:41-43.
- Davidson, J.A., Robison, J., Khailova, L., Frank, B.S., Jaggers, J., Ing, R.J., Lawson, S., Iguidbashian, J., Ali, E., Trece, A. and Soranno, D.E., 2022. Metabolomic profiling demonstrates evidence for kidney and urine metabolic dysregulation in a piglet model of cardiac surgery-induced acute kidney injury. *American Journal of Physiology-Renal Physiology*, **323**(1), pp.F20-F32.
- Davidson, J.A., Frank, B.S., Urban, T.T., Twite, M., Jaggers, J., Khailova, L. and Klawitter, J., 2021. Serum metabolic profile of postoperative acute kidney injury following infant cardiac surgery with cardiopulmonary bypass. *Pediatric Nephrology*, **36**(10), pp.3259-3269.
- Dawson, N., Morris, B.J. and Pratt, J.A. (2011). Subanaesthetic Ketamine Treatment Alters Prefrontal Cortex Connectivity With Thalamus and Ascending Subcortical Systems. *Schizophrenia Bulletin*. DOI:10.1093/schbul/sbr144.
- deAraujo, M., Andrade, L., Coimbra, T.M., Rodrigues, A.C. Jr. and Seguro, A.C. (2005). Magnesium supplementation combined with *N*-acetylcysteine protects against postischemic acute renal failure. *Journal of the American Society of Nephrology* **16**:3339-49.
- Deng, A., Valdivielso, J.M., Munger, K.A., Blantz, R.C. and Thomson, S.C. (2002). Vasodilatory N-Methyl-D-Aspartate Receptors Are Constitutively Expressed in Rat Kidney. *Journal of the American Society of Nephrology* **13**:1381-84.
- Desai, M.M., Gill, I.S., Ramani, A.P., Spaliviero, M., Rybicki, L. and Kaouk, J.H. (2005). The impact of warm ischemia on renal function after laparoscopic partial nephrectomy. *British Journal of Urology International* **95**:377-83.
- Dhawan, J., Benveniste, H., Luo, Z., Nawrocky, M., Smith, S.D. and Biegon, A. (2011). A new look at glutamate and ischemia: NMDA agonist improves long-term functional outcome in a rat model of stroke. *Future Neurology* **6**:823-34.
- Di mari, J.F., Davis, R. and Safirsteinrl. (1999). Mapkactivation determines renal epithelial cell survival during oxidative injury. *American Journal of Physiology* **277**:f195-203.
- Dingledine, R. and Conn, P.J. (2000). Peripheral Glutamate Receptors: Molecular Biology and Role in Taste Sensation. *The Journal of Nutrition* **130**:1039S-42S.
- Ellinger, A. (1904). Die entstehung der kynurensaure. *Zeitschrift für Physikalische Chemie* **43**:325-37.

- Erhardt, S., Olsson, S.K. and Engberg, G. (2009). Pharmacological manipulation of kynurenic acid: potential in the treatment of psychiatric disorders. *CNS Drugs* **23**:91-101.
- Fandrey, J., Rob, P.M. and Jelkmann, W. (1991). Theophylline and magnesium inhibit the contraction elicited with ciclosporin and angiotensin II in mesangial cell cultures. *Nephron* **57**:94-98.
- Faraci, F.M. and Breese, K.R. (1993). Nitric oxide mediates vasodilatation in response to activation of N-methyl-D-aspartate receptors in brain. *Circulation Research* **72**:476-80.
- Folin, O. and Denis, W. (1912). A new (colorimetric) method for the determination of uric acid in blood. *The Journal of Biological Chemistry* **13**:469.
- Foster, A., Vezzani, A., French, E.D. and Schwarcz, R. (1984). Kynurenic acid blocks neurotoxicity and seizures induced in rats by the related brain metabolite quinolinic acid. *Neuroscience Letters* **48**:273-78.
- Friedewald, J.J. and Rabb, H. (2004). Inflammatory cells in ischemic acute renal failure. *Kidney International* **66**:486-91.
- Fukui, S., Schwartz, R., Rapoport, S.I., Takada, Y. and Smith, Q.R. (1991). Blood-brain barrier transport of kynurenines: implications for brain synthesis and metabolism. *Journal of Neurochemistry* **56**:2007-17.
- Getova, D.P. and Doncheva, N.D. (2011). Effects of ketamine on memory and nociception in rats. *Folia Medica (Plovdiv)* **53**:53-59.
- Goligorsky, M.S., Brodsky, S.V. and Noiri, E. (2002). Nitric oxide in acute renal failure: NOS versus NOS. *Kidney International* **61**:855-61.
- Goligorsky, M.S., Brodsky, S.V. and Noiri, E. (2004). NO bioavailability, endothelial dysfunction, and acute renal failure: new insights into pathophysiology. *Seminar in Nephrology* **24**:316-23.
- Gonda, X. (2012). Basic pharmacology of NMDA receptors. *Current Pharmaceutical Design* **18**:1558-67.
- Gonzalez-Cadavid, N.F., Ryndin, I., Vernet, D., Magee, T.R. and Rajfer, J. (2000). Presence of NMDA receptor subunits in the male lower urogenital tract. *Journal of Andrology* **21**:566-78.
- Grimwood, S., Wafford, K.A., Macaulay, A. and Hutson, P.H. (2002). N-Methyl-d-aspartate receptor subtype-selectivity of homoquinolinate: an electrophysiological and radioligand binding study using both native and recombinant receptors. *Journal of Neurochemistry* **82**:794-800.
- Gryglewski, R.J., Palmer, R.M. and Moncada, S. (1986). Superoxide anion is involved in the breakdown of endothelium-derived vascular relaxing factor. *Nature* **320**:454-56.
- Gupta, A., Devi, L.A. and Gomes, I. (2011). Potentiation of  $\mu$ -opioid receptor-mediated signaling by ketamine. *Journal of Neurochemistry* **119**:294-302.
- Hallak, M. (1998). Effect of parenteral magnesium sulfate administration on excitatory amino acid receptors in the rat brain. *Magnesium Research* **11**:117-31.

- Headley, P.M., Parsons, C.G. and West, D.C. (1987). Therole of Nmethylaspartate receptors in mediating responses of rat and cat spinal neurones to defined sensory stimuli. *The Journal of Physiology (London)* **385**:169-88.
- Hernandez, D.J., Roberts, W.B., Miles-Thomas, J., Magheli, A., Saha, S., Schaeffer, E.M., Racusen, L.C. and Allaf, M.E. (2008). Can ischemic preconditioning ameliorate renal ischemia-reperfusion injury in a single-kidney porcine model? *Journal of Endourology* **22**:2531-36.
- Hilmas, C., Pereira, E.F., Alkondon, M., Rassoulpour, A., Schwarcz, R. and Albuquerque, E.X. (2001). The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: physiopathological implications. *The Journal of Neuroscience* **21**:7463-73.
- Hilton, R. (2006). Acute renal failure. *British Medical Journal* **333**:786-90.
- Himmelseher, S. and Durieux, M.E. (2005). Revising a dogma: ketamine for patients with neurological injury? *Anesthesia and Analgesia* **101**:524-34.
- Hollmann, M. and Heinemann, S. (1994). Cloned glutamate receptors. *Annual Review of Neuroscience* **17**:31-108
- Homer, A. (1914). The constitution of kynurenic acid. *The Journal of Biological Chemistry* **17**:509-18.
- Hou, S.H., Bushinsky, D.A., Wish, J.B., Cohen, J.J. and Harrington, J.T. (1983). Hospital-acquired renal insufficiency: a prospective study. *The American Journal of Medicine* **74**:243-48.
- Hsu, C.Y., Chertow, G.M., McCulloch, C.E., Fan, D., Ordonez, J.D. and Go, A.S. (2009). Nonrecovery of kidney function and death after acute on chronic renal failure. *Clinical Journal of American Society of Nephrology* **4**:891-98.
- Hynd, M., Scott, H.L. and Dodd, P.R. (2004). Glutamate-mediated excitotoxicity and neurodegeneration in Alzheimer's disease. *Neurochemistry International* **45**:583-95.
- Igarashi, K. and Kashiwagi, K. (2000). Polyamines: mysterious modulators of cellular functions. *Biochemical and Biophysical Research Communications* **271**:559-64.
- Igarashi, K. and Kashiwagi, K. (2010). Modulation of cellular function by polyamines. *The International Journal of Biochemistry Cell Biology* **42**:39-51.
- Infante, C., Díaz, M., Hernández, A., Constandil, L. and Pelissier, T. (2007). Expression of nitric oxide synthase isoforms in the dorsal horn of monoarthritic rats: effects of competitive and uncompetitive N-methyl-D-aspartate antagonists. *Arthritis Research and Therapy* **9**:R53.
- Inthorn, D. and Hoffmann, J.N. (1996). Elimination of inflammatory mediators by hemofiltration. *The International Journal of Artificial Organs* **19**:124-26.
- Isaac, J., Tōgel, F.E. and Westenfelder, C. (2007). Extent of glomerular tubularization is an indicator of the severity of experimental acute kidney injury in mice. *Nephron Experimental Nephrology* **105**:e33-40.

- Ishani, A., Xue, J.L., Himmelfarb, J., Eggers, P.W., Kimmel, P.L., Molitoris, B.A. and Collins, A.J. (2009). Acute kidney injury increases risk of ESRD among elderly. *Journal of the American Society of Nephrology* **20**:223-28.
- Jaffe, M. (1886). Über den niederschlag, welchen pikrinsäure in normalem ham erzeugt und übereineneuereaktion des kreatinins. *Zeitschrift Fur Physikalische Chemie* **10**:391-400.
- Johnson, J.W. and Ascher, P. (1987). Glycine potentiates the NMDA response in cultured mouse brain neurons. *Nature* **325**:529-31.
- Kaeberlein, M. (2009). Spermidine surprise for a long life. *Nature Cell Biology* **11**:1277-78.
- Kalantar-Zadeh, K., Brennan, M.L. and Hazen, S.L. (2006). Serum myeloperoxidase and mortality in maintenance hemodialysis patients. *American Journal of Kidney Diseases* **48**:59-68.
- Kalman, S. (2002). Cytokines and growth factors in renal injury. *Journal of Turkish Society of Nephrology* **11**:140-43.
- Kartvelishvily, E., Shleper, M., Balan, L., Dumin, E. and Wolosker, H. (2006). Neuron-derived d-serine release provides a novel means to activate N-methyl-d-aspartate receptors. *The Journal of Biological Chemistry* **281**:14151-62.
- Kato, R. and Foex, P. (2002). Myocardial protection by anesthetic agents against ischemia-reperfusion injury: an update for anesthesiologists. *Canadian Journal Anaesthesia* **49**:777-91.
- Kaufman, J., Dhakal, M., Patel, B. and Hamburger, R. (1991). Community-acquired acute renal failure. *American Journal of Kidney Diseases* **17**:191-98.
- Kew, J.N. and Kemp, J.A. (2005). Ionotropic and metabotropic glutamate receptor structure and pharmacology. *Psychopharmacology (Berlin)* **179**:4-29.
- Khan, M.J., Seidman, M.D., Quirk, W.S. and Shivapuja, B.G. (2000). Effects of kynurenic acid as a glutamate receptor antagonist in the guinea pig. *European Archives of Otorhinolaryngology* **257**:177-81.
- Kim, Y., Cho, H.Y., Ahn, Y.J., Kim, J. and Yoon, Y.W. (2012). Effect of NMDA NR2B antagonist on neuropathic pain in two spinal cord injury models. *Pain* **153**:1022-29.
- Kiss, C., Ceresoli-Borroni, G., Guidetti, P., Zielke, C.L., Zielke, H.R. and Schwarcz, R. (2003). Kynurene production by cultured human astrocytes. *Journal of Neural Transmission* **110**:1-14.
- Knyihár-Csillik, E., Toldi, J., Mihály, A., Krisztin-Péva, B., Chadaide, Z., Németh, H., Fenyo, R. and Vécsei, L. (2007). Kynurenone in combination with probenecid mitigates the stimulation-induced increase of c-fosimmunoreactivity of the rat caudal trigeminal nucleus in an experimental migraine model. *Journal of Neural Transmission* **114**:417-21.
- Kobayashi, M., Sugiyama, H., Wang, D.H., Toda, N., Maeshima, Y., Yamasaki, Y., Masuoka, N., Yamada, M., Kira, S. and Makino, H. (2005). Catalase deficiency renders remnant kidneys more susceptible to oxidant tissue injury and renal fibrosis in mice. *Kidney International* **68**:1018-31.

- Kramer, A.H. (2012). Early ketamine to treat refractory status epilepticus. *Neurocritical Care* **16**:299-305.
- Krawisz, J.E., Sharon, P. and Stenson, W.F. (1984). Quantitative assay for acute intestinal inflammation based on myeloperoxidase activity. Assessment of inflammation in rat and hamster models. *Gastroenterology* **87**:1344-50.
- Kribben, A., Edelstein, C.L. and Schrier, R.W. (1999). Pathophysiology of acute renal failure. *Journal of Nephrology* **12**:S142-51.
- Krizbai, I.A., Deli, M.A., Pestena'cz, A., Siklo's, L., Szabo, C.A., Andra's, I. and Joo,F. (1998). Expression of glutamate receptors on cultured cerebral endothelial cells. *Journal of Neuroscience Research* **54**: 814-19.
- Lapin, I.P. (1976). Depressor effect of kynurenone and its metabolites in rats. *Life Sciences* **19**:1479-84.
- Lapin, I.P. (1980). Experimental studies on kynurenone as neuroactive tryptophan metabolites: past, present and future. *Trends in Pharmacological Sciences* **1**:410-13.
- Laube, B., Hirai, H., Sturqess, M., Betz, H. and Kuhse, J. (1997). Molecular determinants of agonist discrimination by NMDAR subunits: analysis of the glutamate binding site on the NR2B subunit. *Neuron* **18**:493-503.
- Laube, B., Kuhse, J. and Betz, H. (1998). Evidence for a tetrameric structure of recombinant NMDARs. *The Journal of Neuroscience* **18**:2954-61.
- Lee, C.Y., Jan, W.C., Tsai, P.S. and Huang, C.J. (2011). Magnesium sulfate mitigates acute lung injury in endotoxemia rats. *The Journal of Trauma* **70**:1177-85.
- Leung, J.C., Marphis, T., Craver, R.D. and Silverstein, D.M. (2004). Altered NMDA receptor expression in renal toxicity: protection with a receptor antagonist. *Kidney International* **66**:167-176.
- Liano, F. and Pascual, J. (1996). Epidemiology of acute renal failure: a prospective, multicenter, community-based study. *Kidney International* **50**:811-18.
- Lieberthal, W. (1998). Biology of ischemic and toxic renal tubular injury: role of nitric oxide and the inflammatory response. *Current Opinion in Nephrology and Hypertension* **7**:289-95.
- Lieberthal, W. and Levine, J.S. (1996). Mechanisms of apoptosis and its potential role in renal tubular epithelial cell injury. *American Journal of Physiology* **271**:F477-88.
- Loix, S., De Kock, M. and Henin, P. (2011). The anti-inflammatory effects of ketamine: state of the art. *Acta Anaesthesiologica Belgica* **62**:47-58.
- Lombardi, G., Dianzani, C., Miglio, G., Canonico, P.L. and Fantozzi, R. (2001). Characterization of ionotropic glutamate receptors in human lymphocytes. *British Journal of Pharmacology* **133**: 936-44.
- Lugo-Huitron, R., Blanco-Ayala, T., Ugalde-Muniz, P., Carrillo-Mora, P., Pedraza-Chaverri, J., Silva-Adaya, D., Maldonado, P.D., Torres, I., Pinzón, E., Ortiz-Islas, E., López, T., García, E., Pineda, B., Torres-Ramos, M., Santamaría, A. and La Cruz, V.P. (2011). On the antioxidant properties of kynurenic acid: Free radical scavenging activity and inhibition of oxidative stress. *Neurotoxicology and Teratology* **33**:538-47.

- Luscher, C. and Malenka, R.C. (2012). NMDA Receptor-Dependent Long-Term Potentiation and Long-Term Depression (LTP/LTD). *Cold Spring Harbor Perspectives in Biology* DOI:10.1101/cshperspect.a005710.
- Ma, M.C., Huang, H.S. and Chen, C.F. (2002). Impaired renal sensory responses after renal ischemia in the rat. *Journal of the American Society of Nephrology* **13**:1872-83.
- MacMillan-Crow, L.A., Crow, J., Kerby, J., Beckman, J.S. and Thompson, J.A. (1996) Nitration and inactivation of manganese superoxide dismutase in chronic rejection of human renal allografts. *Proceedings of the National Academy of Sciences USA* **93**:11853-58.
- Madeo, F., Eisenberg, T., Büttner, S., Ruckenstein, C. and Kroemer, G. (2010). Spermidine :A novel autophagy inducer and longevity elixir. *Autophagy* **6**:160-62.
- Malis, C.D. and Bonventre, J.V. (1986). Mechanism of calcium potentiation of oxygen free radical injury to renal mitochondria. A model for postischemic and toxic mitochondrial damage. *The Journal of Biological Chemistry* **261**:14201-08.
- Mansouri, E., Panahi, M., Ghaffari, M.A. and Ghorbani, A. (2011). Effects of grape seed proanthocyanidin extract on oxidative stress induced by diabetes in rat kidney. *Iranian Biomedical Journal* **15**:100-06.
- Mathew, S.J., Shah, A., Lapidus, K., Clark, C., Jarun, N., Ostermeyer, B. and Murrough, J.W. (2012). Ketamine for treatment-resistant unipolar depression: current evidence. *CNS Drugs* **26**:189-204.
- Matsuyama, M., Nakatani, T., Hase, T., Kawahito, Y., Sano, H., Kawamura, M. and Yoshimura, R. (2004). The expression of cyclooxygenases and lipoxygenases in renal ischemia-reperfusion injury. *Transplantation Proceedings* **36**:1939-42.
- Mauskop, A. and Altura, B.M. (1998). Role of magnesium in the pathogenesis and treatment of migraines. *Clinical Neurosciences* **5**:24-27.
- Mayer, M.L. (2011). Emerging models of glutamate receptor ion channel structure and function. *Structure* **19**: 1370-80.
- Mayer, M.L., Westbrook, G.L. and Guthrie, P.B. (1984). Voltage-dependent block by Mg of NMDA responses in spinal cord neurons. *Nature* **309**:261-63.
- Mehta, R.L., Kellum, J.A., Shah, S.V., Molitoris, B.A., Ronco, C., Warnock, D.G. and Levin, A. (2007). Acute Kidney Injury Network: report of an initiative to improve outcomes in acute kidney injury. *Critical Care* **11**:R31.
- Mehta, R.L., Pascual, M.T., Soroko, S., Savage, B.R., Himmelfarb, J., Ikizler, T.A., Paganini, E.P. and Chertow, G.M. (2004). Spectrum of acute renal failure in the intensive care unit: the PICARD experience. *Kidney International* **66**:1613-21.
- Minatogawa, Y., Noguchi, T. and Kido, R. (1974). Kynurenone pyruvate transaminase in rat brain. *Journal of Neurochemistry* **23**:271-72.
- Molitoris, B.A. and Marrs, J. (1999). The role of cell adhesion molecules in ischemic acute renal failure. *The American Journal of Medicine* **106**:583-92.
- Montoliu, J. (1997). Clearance of inflammatory mediators through continuous renal replacement therapy. *Blood Purification* **15**:305-08.

- Monyer, H., Sprengel, R., Schoepfer, R., Herb, A., Higuchi, M., Lomeli, H., Burnashev, N., Sakmann, B. and Seeburg, P.H. (1992). Heteromeric NMDA receptors:molecular and functional distinction of subtypes. *Science* **256**:1217-21.
- Moojen, V.K., Damiani-Neves, M., Bavaresco, D.V., Pescador, B.B., Comim, C.M., Quevedo, J. and Boeck, C.R. (2012). NMDA preconditioning prevents object recognition memory impairment and increases brain viability in mice exposed to traumatic brain injury. *Brain Research* **1466**:82-90.
- Morigi, M., Macconi, D., Zoja, C., Donadelli, R., Buelli, S., Zanchi, C., Ghilardi, M. and Remuzzi, G. (2002). Protein overload-induced NFkappaB activation in proximal tubular cells requires H<sub>2</sub>O<sub>2</sub> through a PKC-dependent pathway. *Journal of the American Society of Nephrology* **13**:1179-89.
- Moroni, F., Russi, P., Carla, V. and Lombardi, G. (1988). Kynurenic acid is present in the rat brain and its content increases during development and aging processes. *Neuroscience Letters* **94**:145-50
- Morris, D.R. and Levenson, C.W. (2012). Ion channels and zinc: mechanisms of neurotoxicity and neurodegeneration. *Journal of Toxicology* **2012**:785647.
- Moss, D.W and Henderson, A.R. (1986). Enzymes. In: Burtis, C.A. and Ashwood, E.R. (eds). *Tietz Textbook of Clinical Chemistry*. pp 735-896. Philadelphia, Saunders Co.
- Muir, K.W. (1988). New experimental and clinical data on the efficacy of pharmacological magnesium infusions in cerebral infarcts. *Magnesium Research* **11**:43-56.
- Muller, G.A., Muller, C.A. and Markovic-Lipkovski, J. (1996). Adhesion molecules in renal diseases. *Renal Failure* **18**:711-24.
- Nahum-Levy, R., Fossum, L.H., Skolnick, P. and Benveniste, M. (1999). Putative partial agonist 1-aminocyclopropanecarboxylic acid acts concurrently as a glycine-site agonist and a glutamate-site antagonist at N-methyl-d-aspartate receptors. *Molecular Pharmacology* **56**:1207-18.
- Naka, Y., Roy, D.K., Smerling, A.J., Michler, R.E., Smith, C.R., Stern, D.M., Oz, M.C. and Pinsky, D.J. (1995). Inhaled nitric oxide failsto confer the pulmonary protection provided by distal stimulation of the nitric oxide pathway at the level of cyclic guanosine monophosphate. *The Journal of Thoracic and Cardiovascular Surgery* **110**:1434-40.
- Nash, K., Hafeez, A. and Hou, S. (2002). Hospital-acquired renal insufficiency. *American Journal of Kidney Diseases* **39**:930-36.
- Nemeth, H., Robotka, H., Kis, Z., Rozsa, E., Janaky, T., Somlai, C., Marosi, M., Farkas, T., Toldi, J. and Vécsei, L. (2004). Kynurenone administered together with probenecid markedly inhibits pentylenetetrazol-induced seizures. An electrophysiological and behavioural study. *Neuropharmacology* **47**:916-25.
- Nichans, W.G. and Samuelson, B. (1968). Formation of malondialdehyde from phospholipids arachidonate during microsomal lipid peroxidation. *European Journal of Biochemistry* **6**:126-30.

- Nikolaev, M.V., Magazanik, L.G. and Tikhonov, D.B. (2012). Influence of external magnesium ions on the NMDA receptor channel block by different types of organic cations. *Neuropharmacology* **62**:2078-85.
- Noguchi, N. and Niki, E. (1998). Dynamics of vitamin E action against LDL oxidation. *Free Radical Research* **28**:561-72.
- Noiri, E., Nakao, A., Uchida, K., Tsukahara, H., Ohno, M., Fujita, T., Brodsky, S. and Goligorsky, M.S. (2001). Oxidative and nitrosative stress in acute renal ischemia. *American Journal of Physiology* **281**:F948-57.
- Nose, K. (2000). Role of reactive oxygen species in the regulation of physiological functions. *Biological and Pharmaceutical Bulletin* **23**:897-903.
- Nowak, L., Bregestovski, P., Ascher, P., Herbet, A. and Prochiantz, A. (1984). Magnesium gates glutamate-activated channels in mouse central neurons. *Nature* **307**:462-65.
- Okuno, E., Minatoaawa, Y., Nakamura, M., Kamado, N., Nakanishi, J., Makino, M. and Kido, R. (1980). Crystallization and characterization of human liver kynurenine-glyoxylateaminotransferase. *The Journal of Biochemistry* **189**:581-90.
- Padanilam, B.J. and Lewington, A.J. (1999). Molecular mechanisms of cell death and regeneration in acute ischemic renal injury. *Current Opinion in Nephrology and Hypertension* **8**:15-19.
- Paller, M.S., Hoidal, J.R. and Ferris, T.F. (1984). Oxygen free radicals in ischemic acute renal failure in the rat. *The Journal of Clinical Investigation* **74**:1156-64.
- Panatier, A., Theodosis, D.T., Mothet, J.P., Touquet, B., Pollegioni, L., Poulain, D.A. and Oliet, S.H. (2006). Glia-derived d-serine controls NMDA receptor activity and synaptic memory. *Cell* **125**:775-84.
- Patel, N.S.A., Cuzzocrea, S., Chatterjee, P.K., Di Paola, R., Sautebin, L., Britti, D. and Thiemer, C. (2004). Reduction of renal ischemia-reperfusion injury in 5-lipoxygenase knockout mice and by the 5-lipoxygenase inhibitor zileuton. *Molecular Pharmacology* **66**:220-27.
- Pirinen, E., Gylling, H., Itkonen, P., Yaluri, N., Heikkinen, S., Pietilä, M., Kuulasmaa, T., Tusa, M., Cerrada-Gimenez, M., Pihlajamäki, J., Alhonen, L., Janne, J., Miettinen, T.A. and Laakso, M. (2010). Activated polyamine catabolism leads to low cholesterol levels by enhancing bile acid synthesis. *Amino acids* **38**:549-60.
- Poeggeler, B., Rassoulpour, A., Wu, H.Q., Guidetti, P., Roberts, R.C. and Schwarcz, R. (2007). Dopamine receptor activation reveals a novel, kynureneate-sensitive component of striatal N-methyl-D-aspartate neurotoxicity. *Neuroscience* **148**:188-97.
- Potter, M.C., Elmer, G.I., Bergeron, R., Albuquerque, E.X., Guidetti, P., Wu, H.Q. and Schwarcz, R. (2010). Reduction of endogenous kynurenic acid formation enhances extracellular glutamate, hippocampal plasticity, and cognitive behavior. *Neuropsychopharmacology* **35**:1734-42.

- Qu, X.X., Cai, J., Li, M.J., Chi, Y.N., Liao, F.F., Liu, F.Y., Wan, Y., Han, J.S. and Xing, G.G. (2009). Role of the spinal cord NR2B-containing NMDA receptors in the development of neuropathic pain. *Experimental Neurology* **215**:298-307.
- Robotka, H., Sas, K., Agoston, M., Rozsa, E., Szenasi, G., Gigler, G., Vecsei, L. and Toldi, J. (2008). Neuroprotection achieved in the ischaemic rat cortex with L-kynurenesulphate. *Life Sciences* **23**:915-19.
- Rock, D.M. and Macdonald, R.L. (1992). The polyamine spermine has multiple actions on N-methyld-aspartate receptor single-channel currents in cultured cortical neurons. *Molecular Pharmacology* **41**:83-88.
- Rothman, S.M. and Olney, J.W. (1986). Glutamate and the pathophysiology of hypoxic-ischemic brain damage. *Annals of Neurology* **19**:105-11.
- Safirsteinrl. (1994). Gene expression in nephrotoxic and ischemic acute renal failure. *Journal of the American Society of Nephrology* **4**:1387-95.
- Safirsteinrl. and Bonventre, J. (1995). Molecular response to ischemic and nephrotoxic acute renal failure. Schlondorff, D. and Bonventre, J. (eds). *Molecular Nephrology*. pp 839-54. New York, Marcel Dekker, inc.
- Salman, A.E., Dal, D., Salman, M.A., Iskit, A.B. and Aypar, U. (2005). The effect of ketamine on acute muscular ischaemia reperfusion in rats. *European Journal of Anaesthesiology* **22**:712-16.
- Santangelo, F., Witko-Sarsat, V., Dru' eke, T. and Descamps-Latscha, B. (2004). Restoring glutathione as a therapeutic strategy in chronic kidney disease. *Nephrology Dialysis Transplantation* **19**:1951-55.
- Sapolsky, R. (2005). *Biology and Human Behavior: The Neurological Origins of Individuality*. pp 19-20. The Teaching Company, Chantilly.
- Saunders, J.A., Gandal, M.J. and Siegel, S.J. (2012). NMDA antagonists recreate signal-to-noise ratio and timing perturbations present in schizophrenia. *Neurobiology of Disease* **46**:93-100.
- Schilstrom, B., Konradsson-Geuken, A., Ivanov, V., Gertow, J., Feltmann, K., Marcus, M.M., Jardemark, K. and Svensson, T.H. (2011). Effects of S-citalopram, citalopram, and R-citalopram on the firing patterns of dopamine neurons in the ventral tegmental area, N-methyl-D-aspartate receptor-mediated transmission in the medial prefrontal cortex and cognitive function in the rat. *Synapse* **65**:357-67.
- Schrier, R.W., Wang, W., Poole, B. and Mitra, A. (2004). Acute renal failure: definitions, diagnosis, pathogenesis, and therapy. *The Journal of Clinical Investigation* **114**:5-14.
- Seburg, P.H., Burnashev, N., Kohr, G., Kuner, T., Sprengel, R. and Monyer, H. (1995). The NMDAR channel: molecular design of a coincidence detector. *Recent Progress in Hormone Research* **50**:19-34.
- Seguro, A.C., Araujo, M., Seguro, F.S., Rienzo, M., Magaldi, A.J. and Campos, A.C. (2003). Effects of hypokalemia and hypomagnesemia on zidovudine (AZT) and didanosine (DDI) nephrotoxicity in rats. *Clinical Nephrology* **59**:267-72.

- Shechter, M., Sharir, M., Labrador, M.J., Forrester, J., Silver, B. and BaireyMerz, C.N. (2000). Oral magnesium therapy improves endothelial function in patients with coronary artery disease. *Circulation* **102**:2353-58.
- Shen, L., Li, L., She, H., Yue, S., Li, C. and Luo, Z. (2010). Inhibition of pulmonary surfactants synthesis during N-methyl-D-aspartate-induced lunginjury. *Basic and Clinical Pharmacology and Toxicology* **107**:751-57.
- Sheridan, A.M. and Bonventre, J.V. (2000). Cell biology and molecular mechanisms of injury in ischemic acute renal failure. *Current Opinion in Nephrology and Hypertension* **9**:427-34.
- Skilling, S.R., Smullin, D.H., Beitz, A.J. and Larson, A.A. (1992). Extracellular amino acid concentrations in the dorsal spinalcord of freely moving rats following veratridine and nociceptive stimulation. *Journal of Neurochemistry* **51**:127-32.
- Sobolevsky, A.I., Rosconi, M.P. and Gouaux, E. (2009). X-ray structure, symmetry and mechanism of an AMPA-subtype glutamate receptor. *Nature* **462**:745-56.
- Spentzas, T., Shapley, R.K., Aguirre, C.A., Meals, E., Lazar, L., Rayburn, M.S., Walker, B.S. and English, B.K. (2011). Ketamine inhibits tumor necrosis factor secretion by RAW264.7 murine macrophages stimulated with antibiotic-exposed strains of community-associated, methicillin-resistant *Staphylococcus aureus*. *Bio Med Central Immunology* **25**:12:11.
- Spruston, N., Schiller, Y., Stuart, G. and Sakmann, B. (1995). Activity-dependent action potential invasion and calcium influx into hippocampal CA1 dendrites. *Science* **268**:297-300.
- Srivastava, R.K., Gombar, K.K., Kaur, A.H. and Khosla, P. (1995). Attenuation of morphine-induced antinociception by L-glutamic acid at the spinal site in rats. *Canadian Journal of Anaesthesia* **42**:541-46.
- Star, R.A. (1998). Treatment of acute renal failure. *Kidney International* **54**:1817-31.
- Strauss, K.I. and Marini, A.M. (2002). Cyclooxygenase-2 inhibition protects cultured cerebellar granule neurons from glutamate-mediated cell death. *Journal of Neurotrauma* **19**:627-38.
- Stuart, G., Spruston, N., Sakmann, B. and Häusser, M. (1997). Action potential initiation and backpropagation in neurons of the mammalian CNS. *Trends in Neurosciences* **20**:125-31.
- Swartz, K.J., During, M.J., Freese, A. and Beal, M.F. (1990). Cerebral synthesis and release on kynurenic acid and endogenous antagonist of excitatory amino acid receptors. *The Journal of Neuroscience* **10**:2965-73.
- Szyndler, J., Maciejak, P., Turzyn'ska, D., Sobolewska, A., Walkowiak, J. and Płaz'nik, A. (2012). The effects of electrical hippocampal kindling of seizures on amino acids and kynurenic acid concentrations in brain structures. *Journal of Neural Transmission* **119**:141-49.
- Tanaka, T., Nangaku, M., Miyata, T., Inagi, R., Ohse, T., Ingelfinger, J.R. and Fujita, T. (2004). Blockade of calcium influx through L-type calcium channels attenuates mitochondrial injury and apoptosis in hypoxic renal tubular cells. *Journal of the American Society of Nephrology* **15**:2320-33.

- Tang, S., Leung, J.C.K., Abe, K., Chan, K.W., Chan, L.Y.Y., Chan, T.M. and Lai, K.L. (2003). Albumin stimulates interleukin-8 expression in proximal tubular epithelial cells in vitro and in vivo. *The Journal of Clinical Investigation* **111**:515-27.
- Tayeb, H.O., Yang, H.D., Price, B.H. and Tarazi, F.I. (2012). Pharmacotherapies for Alzheimer's disease: beyond cholinesterase inhibitors. *Pharmacology and Therapeutics* **134**:8-25.
- Thadhani, R., Pascual, M. and Bonventre, J.V. (1996). Acute renal failure. *The New England Journal of Medicine* **334**:1448-60.
- Thurman, J.M. (2007). Triggers of inflammation after renal ischemia/reperfusion. *Clinical Immunology* **123**:7-13.
- Thurnau, G.R., Kemp, D.B. and Jarvis, A. (1987). Cerebrospinal fluid levels of magnesium in patients with preeclampsia after treatment with intravenous magnesium sulfate: a preliminary report. *American Journal of Obstetrics and Gynecology* **157**:1435-38.
- Tian, J., Kim, S.F., Hester, L. and Snyder, S.H. (2008). S-nitrosylation/activation of COX-2 mediates. *Proceedings of the National Academy of Sciences* **105**:10537-40.
- Tibbles, L.A. and Woodgett, J.R. (1999). The stress-activated protein kinase pathways. *Cellular and molecular life sciences* **55**:1230-54.
- Uchino, S., Kellum, J.A., Bellomo, R., Doig, G.S., Morimatsu, H., Morgera, S., Schetz, M., Tan, I., Bouman, C., Macedo, E., Gibney, N., Tolwani, A. and Ronco, C (2005). Acute renal failure in critically ill patients: a multinational, multicenter study. *The Journal of American Medical Association* **294**:813-18.
- van den Bos, R., Charria Ortiz, G.A. and Cools, A.R. (1992). Injections of the NMDA-antagonist D-2-amino-7-phosphonoheptanoic acid (AP-7) into the nucleus accumbens of rats enhance switching between cue-directed behaviours in a swimming test procedure. *Behavioural Brain Research* **48**:165-70
- Wang, H.D., Pango, P.J., Du, Y., Cayatte, A.J., Quinn, M.T., Brecher, P. and Cohen, R.A. (1998). Superoxide anion from the adventitia of the rat thoracic aorta inactivates nitric oxide. *Circulation Research* **82**:810-18.
- Wang, J., Simonavicius, N., Wu, X., Swaminath, G., Reagan, J., Tian, H. and Ling, L. (2006). Kynurenic acid as a ligand for orphan G protein-coupled receptor GPR35. *The Journal Biological Chemistry* **281**:22021-28.
- Watanabe, K., Kanno, T., Oshima, T., Miwa, H., Tashiro, C. and Nishizaki, T. (2008). Vagotomy upregulates expression of the N-methyl-D-aspartate receptor NR2D subunit in the stomach. *Journal of Gastroenterology* **43**:322-26.
- Wee, H.N., Liu, J.J., Ching, J., Kovalik, J.P. and Lim, S.C., 2021. The kynureine pathway in acute kidney injury and chronic kidney disease. *American Journal of Nephrology*, 52(10-11), pp.771-787.
- Wybenga, D.R., Di Glorio, J. and Pileggi, V.J. (1971). Manual and automated methods for urea nitrogen measurement in whole serum. *Clinical Chemistry* **17**:891-95.
- Xiao, W.H. and Bennett, G.J. (1994). Magnesium suppresses neuropathic pain responses in rats via a spinal site of action. *Brain Research* **666**:168-72.

- Yamamoto, T. and Yakih, T.L. (1992). Spinal pharmacology ‘of thermal hyperesthesia induced by constriction of sciatic nerve: excitatory amino acid antagonists. *Pain* **49**:121-28.
- Yang, C.C., Chien, C.T., Wu, M.H., Ma, M.C. and Chen, C.F. (2008). NMDA receptor blocker ameliorates ischemia-reperfusion-induced renal dysfunction in rat kidneys. *American Journal of Physiology Renal Physiology* **294**:F1433-40.
- Zwilling, D., Huang, S.Y., Sathyasaikumar, K.V., Notarangelo, F.M., Guidetti, P., Wu, H.Q., Lee, J., Truong, J., Andrews-Zwilling, Y., Hsieh, E.W., Louie, J.Y., Wu, T., Scearce-Levie, K., Patrick, C., Adame, A., Giorgini, F., Moussaoui, S., Laue, G., Rassoulpour, A., Flik, G., Huang, Y., Muchowski, J.M., Masliah, E., Schwarcz, R. and Muchowski, P.J. (2011). Kynurenine 3-monooxygenase inhibition in blood ameliorates neurodegeneration. *Cell* **145**:863-74.