



TECHNIQUES FOR ARSENIC ANALYSIS, REMOVAL AND ITS BIOMEDICAL APPLICATIONS

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Abstract

In the realm of the environment, a great deal of research has been done on the poisonous and damaging impacts of heavy metals including arsenic, mercury, and others. Due to environmental pollution and the bioaccumulation of certain heavy metals in the food chain, they adversely impact human health despite their presence in low concentrations (10 ppm). Recent research studies suggest that exposure to arsenic during various life stages leads to gut microbial dysbiosis and is linked to immune dysfunction, altered lipid metabolism, and neurobehavioral damage. Therefore, it is important to detect, analyze and remove it from the environment to reduce its direct effect on human health.

Keywords: heavy metals, neurobehavioral damage, immune dysfunction, bioaccumulation

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1. TECHNIQUES FOR ARSENIC ANALYSIS

In current scenario various analytical techniques such as hydride generation atomic fluorescence spectrometry (HG-ASF), Inductively coupled plasma atomic emission spectroscopy (ICP-AES), inductively coupled plasma-mass-spectrometry (ICP-MS), hydride generation atomic absorption spectroscopy (HGAAS), graphite furnace atomic absorption spectroscopy (AAS), and fluorescence spectrometry have been used for the detection of heavy metals including As (Le et al., 2000). These instruments can detect low levels of arsenic, however, highly expensive and sophisticated instrumentation with a well-established laboratory set up is required for effective determination. They are also time consuming, not easily accessible, and are not suitable for on-site analysis. Moreover, with these methods, the cost of analyses can be as high as 8–10 USD per sample (Hu et al., 2012). Now-a-days the focus of researchers is on development of simple, portable and economical sensors with rapid and reliable Arsenite detection/analysis in environmental samples, especially in developing countries and in areas with insufficient infrastructure and technical facilities.

Electrochemical methods are advantageous as they are cost-effective, timely, and easy to implement. Various electroanalytical tools such as cyclic voltammetry (CV), linear sweep voltammetry (LSV), Anodic stripping voltammetry (ASV), differential pulse voltammetry (DPV) and square wave ASV (SWASV) can be employed to detect low concentrations of As with good sensitivity and reliability (Shen et al., 2017). Furthermore, stripping voltammetry techniques are adaptable for successful and rapid field screening, high accuracy, and enhanced sensitivity and demonstrate a very low detection limit (Cavicchioli et al., 2004). Employing the highly sensitive stripping voltammetry technique, researchers have detected As to a low

detection limit and significant sensitivity using various electrode materials under optimized experimental conditions (Forsberg et al., 1975; Rahman et al., 2010; Yang et al., 2016).

Arsenic detection by electrochemical methods is investigated using mercury electrodes (Wang et al., 2013) such as hanging mercury drop electrodes (HMDE) because they provide a wider potential window for redox reaction of various metals and regeneration of a clean surface is quite easier via simply creating a new mercury drop. However, because of obvious toxicity considerations from mercury, it is very difficult to discard the used mercury and clean the whole electrochemical setup after each measurement (Kwang-Seok Yun et al., n.d.). Furthermore, various metals such as gold (Au), silver (Ag), and Hg could not be potentially detected on HMDE. Therefore, they have been significantly replaced by a number of solid electrodes such as platinum (Forsberg et al., 1975) silver, boron-doped diamond electrode (A. O. Simm et al., 2005), gold microwire electrode (Liu et al., 2014), gold microdisk (Simm et al., 2004) and iridium oxide-modified boron-doped diamond electrodes (Salimi et al., 2004). Recently, researchers investigated the platinum NP-modified glassy carbon electrode for electrochemical sensing of arsenic in 1 M aqueous HClO₄ electrolyte solution. After optimization, a lower detection limit (LOD) of 2.1 ppb was obtained (Dai and Compton, 2006). Further, an iridium-modified boron-doped diamond electrode was generated by ion implantation method and exploited directly for electrochemical detection of arsenic to an LOD of 1.5 ppb. The aforementioned sensor was favorably able to selectively sense increased arsenic in tap water containing a substantial amount of various other elements as well (Ivandini et al., 2006). However, these materials are expensive, thus making As^{III} detection not feasible for long-term applications.

The successful large-scale implication of the electrochemical sensors mainly depends on careful designing and fabrication of desired materials, which can facilitate adequate accumulation and subsequent oxidation of the analyte from its surface under optimized electrochemical conditions. Among many other solid electrode-based sensors, gold (Au)-centered materials have demonstrated enhanced electrochemical activity because of their advanced characteristic features in the field of catalysis, electroanalysis, and nanoscale devices (A. Simm et al., 2005). They have excellent electronic, optical, and electrical characteristics, which merely depend on the surface morphology and size of Au particles (Feeney and Kounaves, 2000).

Therefore, microstructured/nanostructured Au electrodes have been exploited largely for applications in the detection of arsenic in water. They are superior to the commercially available metal electrodes because of the presence of more electroactive sites, much enhanced electron-transfer rate and favorable electrochemical kinetics (Welch et al., 2004).

Gold-based nanoscale materials can be fabricated by chemical synthesis, ultraviolet light or electron beam irradiation and electrochemical methods (Tan et al., 2002). Electrochemical methods are much facile and easy to use relative to other methods (Fukushima et al., 2003). Gold nanoparticles can be electrodeposited on the electrode using 0.1 mM HAuCl_4 solution, which show good As sensing at parts per billion levels (Dai et al., 2004). Gold nanoelectrodes also enable simultaneous detection of As, Hg, and Cu (Joya and de Groot, 2016). Furthermore, gold NP-modified indium tin oxide (ITO)-coated glass electrodes were prepared by direct electrodeposition method from 0.5 M H_2SO_4 solution containing HAuCl_4 solution. The Au-based films were nanostructured and detected As to an LOD

of 5.0 ppb using the LSV technique (Babar et al., 2019).

However, it is a tedious method, and during deposition, subsequent accumulation of Au ions on cathodic sites also poses a challenge. To address this issue, separators are employed, which make the system more complex and thereby increase the cost for analyses (Joya and de Groot, 2016). The gold nanotextured electrode (Au/GNE) assemblage is exceedingly stable with promising reproducibility maintaining highly active nanoscale surface features by repeating the analyses several times and allows for very reliable, selective, and highly sensitive detection of arsenic using CV and SWASV. This method is also successfully employed in real water samples for arsenic analysis. In the complex system containing Cu^{2+} , Ni^{2+} , Fe^{2+} , Pb^{2+} , Hg^{2+} and other ions, Au/GNE is amazingly applicable for highly selective and sensitive detection of As in water. Next, the experiments are under process to scale up the Au/GNE-based electrochemical sensors for real-time applications owing to their simple fabrication and assembly, high reproducibility and robustness, and electro analytical performance for arsenic sensing (Babar et al., 2019).

2. REMOVAL OF ARSENIC FROM WATER

Removal of arsenic is very important and is the focus of several researchers, industries, environmental groups (Alka et al., 2021). The removal of arsenic from water can be achieved through different mechanisms, methods, technologies including adsorption, chemical precipitation, ion exchange, phytoremediation, electro kinetic techniques and membrane technology. Recent research focuses on different traditional and emerging technologies for the improvement of complex resources active in the elimination of arsenic and other substantial metals. Recently, the number of studies on arsenic

removal techniques have been continuously increasing.

A. Nanomaterials as adsorbents

Adsorption has been a commonly used technique for water treatment as early as 4000 BCE. Cutting-edge Egypt, adsorption was used for the coloring of silk fibers, cotton fibers and some plant and animal fibers, in addition to decoloration of beverages and diet. Adsorption finds several applications in day-to-day life. In adsorption, the materials existing in a fluid stage are collected or adsorbed on the solid stage by physical/chemical adsorption, followed by their subsequent removal from the liquid, as a mass transfer. Arsenic removal by adsorption is economically feasible and very efficient, moreover, it does not require use of chemical additives. It is easy to use and applicable in areas lacking skilled manpower, moreover it does not require consistent electricity supply. The quantitative removal efficiencies reported for arsenate and arsenite remediation have been as high as >95% (M. Kumar et al., 2019; R. Kumar et al., 2019; Ratna Kumar et al., 2004).

Nanomaterials show significant benefits in the adsorption of arsenic from water because of greater superficial adsorption movement and great reactivity. The adsorbent nanomaterials showing excellent removal efficiency and less maintenance cost, particularly for removal of low concentration of heavy metals, highlight the use of adsorption as an encouraging method in arsenic removal from water. Presently, nanomaterial including carbon materials, metal oxides, metal-organic framework and chitosan have been used for removing arsenic from water (Alka et al., 2021; Liu et al., 2015, 2018). The unique hollow structure of carbon nanotubes, their large specific surface area, high porosity, and rapid transport of water make them ideal candidates as adsorbents for As elimination from water (Addo Ntim and Mitra, 2011; Dehghani et al., 2015). Vadahanambi et al. designed 3D graphene carbon nanotube-iron oxide for the elimination of arsenic

from water. Iron-oxide nanoparticles shows significant adsorption of arsenic from water (Vadahanambi et al., 2013). Andjelkovic et al. also fabricated 3D graphene-iron oxide nanoparticle aerogel for elimination of arsenic from water (Andjelkovic et al., 2015). Iron-oxide covered carbon nanotubes for elimination of arsenic from water (Ma et al., 2018) but the nanotubes failed to show effective adsorption because of hydrophobic shells, unfortunate dispensability, and absence of functional groups.

B. Ion-exchange technology

Ion-exchange is a physicochemical process employed for the elimination of arsenic from the environment. In this technique, the ions were retained electrostatically on the solid surface and exchanged from the solution with ions having similar charge (Katsoyiannis and Zouboulis, 2006). Ion exchange is an effective method for adsorption and is mainly applied to decrease water hardness. It is also employed for excerpt pollutants like nitrate, arsenate, chromate and selenite ions from water (Al-jubouri and Holmes, 2020). The United States Environmental Protection Agency (EPA) has suggested definite ion-exchange materials, particularly chloride form, aimed to arsenic elimination (Jadhav et al., 2015). The ion-exchange resins remove the arsenic via mechanism describes ion-exchange resins that are filled by chloride ions in an conversation spot wherever water contaminated (Shankar et al., 2014). A study described the elimination of arsenic from water through ion-exchange resins. Hence these ion exchanges resins can be used for the elimination of toxic arsenic from water. Some factors that affect the removal of arsenic include the entire liquefied items, arsenic concentration, kind of resin used for removal and competing ions (Karakurt et al., 2019; Sarkar and Paul, 2016). Rivero et al. also demonstrated the removal of arsenic from water by using resin in a hybrid ion electro dialysis process.

C. Membrane technology

The membrane is an extensively recognized technology for the filtration of water and is one of the highest well-organized effective methods of arsenic removal with a potential to remove 96% of contaminated As from portable and groundwater. The technique is more effective for removing pollutants and cost-effective at the same time as the process requirements are minimal. Another advantage of this method is that it does not involve any chemical usage (Gonzalez et al., 2019; Ungureanu et al., 2015). Recent reports suggest that several forms of membranes are employed in the elimination of arsenic from water systems with their applications in technologies such as nanofiltration, microfiltration, ultra-filtration, and reverse osmosis (Gonzalez et al., 2019; Pramod et al., 2020). However, Pramod et al. used the combination of microfiltration and heterogeneous Fenton method for the elimination of arsenic from water (Pramod et al., 2020).

D. Chemical precipitation

Chemical precipitation is a method which uses sulfides, ferric salts, calcium and magnesium salts and other chemicals for the elimination of As from water. The chemicals assist in eliminating arsenic by changing dissolved form to its lesser soluble form. Precipitation by ferric arsenate and calcium arsenate is the most useful method for removal of arsenic in wastewater (Long et al., 2019). Chemical precipitation has been also used to treat arsenic and calcium from gold mining waste using two-stage nanofiltration (Sarankumar et al., 2020). Di Iorio et al. developed magnetite nanoparticles for the removal of arsenic from wastewater (Di Iorio et al., 2019).

E. Phytoremediation

Phytoremediation is a widely recognized technique which uses plants for the removal of contaminants. The major advantage of this technique is limited nutrient requirement, lesser maintenance and its role in ecological sustainability (Manoj et al., 2020). Phytoremediation is performed using plants by wide root system, great

acceptance of toxicants and fast growing rate.

3. BIOMEDICAL APPLICATIONS OF ARSENIC

Being considered as one of the oldest poisons, arsenic is too recognized to consume a miracle effect for management several illnesses such as cancer, ulcers, malaria and bubonic plague (Zhao et al., 2021). Around 2,000 years ago, Chinese and Greek healers used arsenic for the treatment of major diseases from syphilis to cancer. From the 20th century, the discovery of chemotherapy and antibiotics led to the abandonment of arsenic based treatments (Zhu et al., 2002). Literature review suggests that these materials have been used as chemotherapeutics in leukemia and cancers due to their anti-proliferative and pro-apoptotic properties (Dilda and Hogg, 2007; Ettliger et al., 2019; Kim et al., 2017; Tian et al., 2020; Yoon et al., 2016; Yu et al., 2020; Zhang et al., 2016). The therapeutic success further encouraged researchers to explore arsenic as a potential future solution for other types of cancers. For example, In a study the effectiveness of a combination of arsenic trioxide and L-buthionine-sulfoximine against advanced solid tumors was assessed. More detailed results of the tumor in figure 1 indicate the difference between the treatment and control group. The As₂O₃ (arsenite) shows cytotoxic effect by the generation of reactive oxygen species (ROS) followed by inhibition of radical scavenging that would enhance the therapeutic efficacy (Maeda et al., 2004). Multiple nanotechnology-based therapies are under development for the effective delivery of As. However, Zhang et al. developed core-shell nanoparticulate arsenic trioxide for effective treatment of solid tumors through the facile route (Zhang et al., 2016). pH-sensitive (zeolitic imidazolate framework-8 based) nanoparticles were developed for effective delivery of arsenic trioxide (Zhang et al., 2016). The anti-tumor activity of sodium meta-arsenite in glioblastoma through

advanced Akt activities was assessed. Briefly, to estimate the anti-tumor activity of sodium meta-arsenite (dose 2 mg/kg and 5 mg/kg), the tumor was evaluated for 21 days in mice injected with U87-MG orthotopic xenograft tumor. The results showed a important decrease in growth of tumor with reduced Akt stimulation as shown in figure 2. The effective amounts of sodium meta-arsenite employed in this case have remained described to be non-toxic (Lee et al., 2020). Arsenic trioxide is used as an anticancer agent traditionally; it demonstrates a important healing

outcome against severe promyelocytic leukemia (Zhu et al., 2002). Recently, the U.S. FDA approved arsenic trioxide as the first-line management against acute promyelocytic leukemia. The mechanism of arsenic trioxide triggering both apoptosis and differentiation of leukemic cells, in a way similar to that of retinoic acid (Antman, 2001; Leu and Mohassel, 2009; Mathews et al., 2001). The result of arsenic trioxide on cervical cancer and reported an increased apoptosis by 3-fold related to control group was observed (L. Zhang et al., 2020).

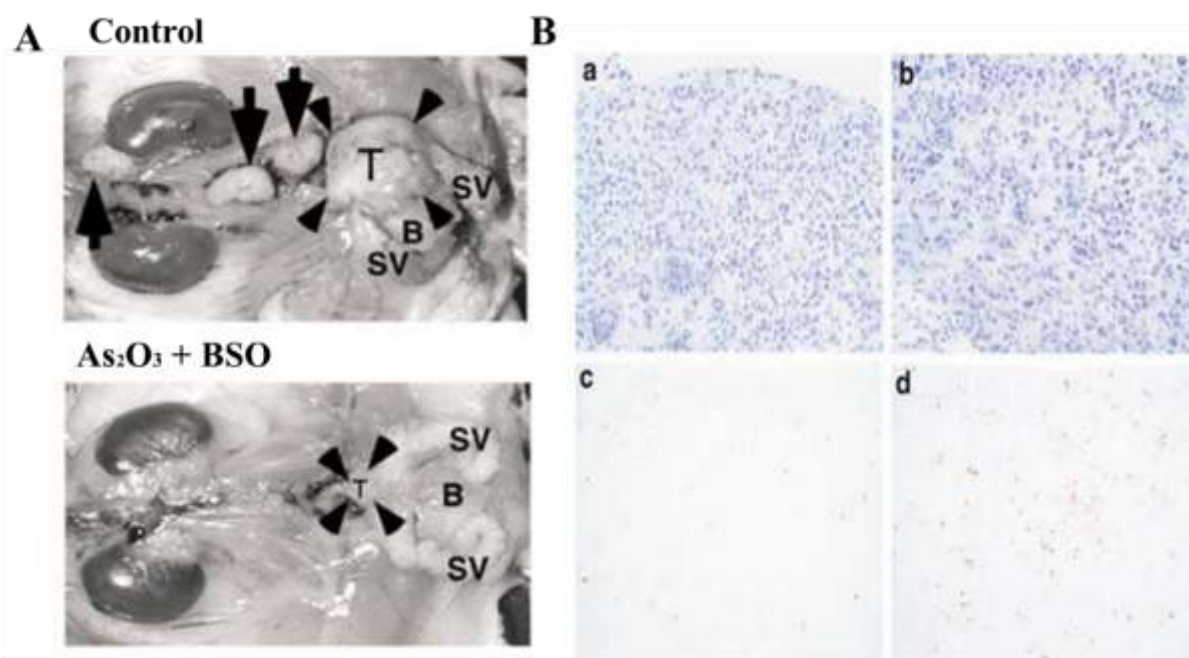


Fig 1: In vivo tumor growth inhibition and survival rates in the orthotopic mouse model of androgen-independent prostate cancer treated with As_2O_3 (Arsenic trioxide) and BSO (Buthionine-sulfoximine). (A) Representative cases 7 weeks after orthotopic inoculation of PC-3 cells. Seminal vesicles (SV) and the bladder (B) (B) Representative histology of an orthotopic tumor formed by PC-3 cells after treatment. Hematoxylin and eosin staining (a, b) and in situ TUNEL (TdT-mediated dUTP Nick End Labeling) assay for detection of apoptosis (c, d) were performed in the saline-treated group (a, c)

and the group treated with 2 mg/kg As_2O_3 plus BSO (b, d). Growth inhibition is clear both in the orthotopic tumor (black arrowheads) and retroperitoneal lymph node metastases (black arrows) in the mouse treated with 2 mg/kg As_2O_3 plus BSO. (Modified from Maeda H, Hori S, Ohizumi H, Segawa T, Kakehi Y, Ogawa O, et al. Effective treatment of advanced solid tumors by the combination of arsenic trioxide and L-buthionine-sulfoximine. *Cell Death Differ* 2004; 11:737–46. <https://doi.org/10.1038/sj.cdd.4401389>) (Maeda et al., 2004).

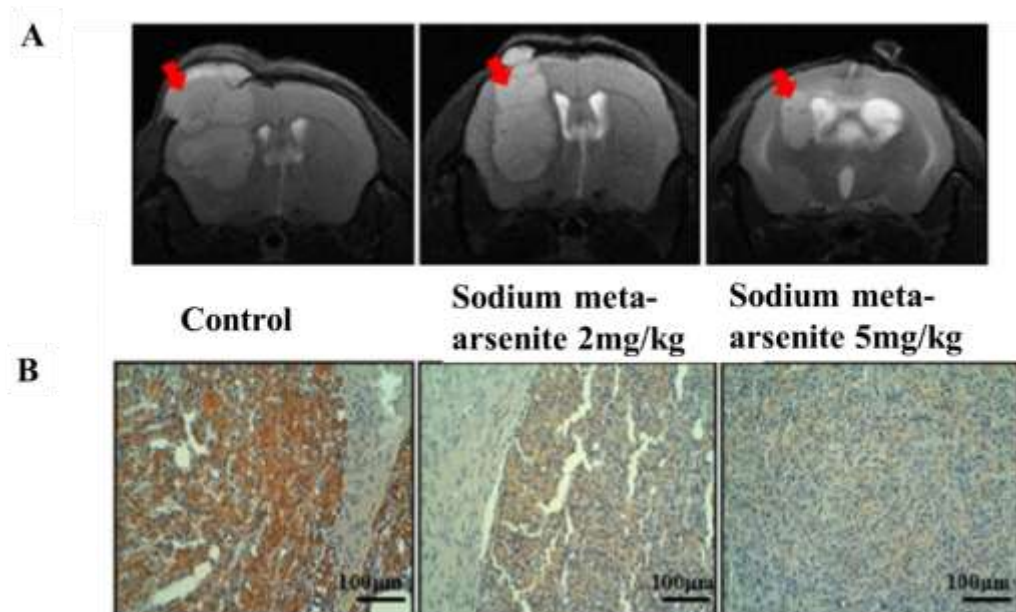


Fig 2: Anti-tumor effects of sodium meta-arsenite in orthotopic xenograft models. (A) Brain MRI images on day 21 of mice (control, and subjected to 2 mg/kg and 5 mg/kg of sodium meta-arsenite). Representative pictures indicate mouse brain regions of the corpus callosum and its surrounding structures. (B) Tumor tissues were fixed and stained with anti-pAkt antibodies. Scale bars = 100 μm . White arrows indicate tumors. (Adapted from Lee EJ, Sung JY, Koo KH, Park JB, Kim DH, Shim J, et al. Anti-tumor effects of sodium meta-arsenite in glioblastoma cells with higher akt activities. *Int J Mol Sci* 2020;21:1–19.

<https://doi.org/10.3390/ijms21238982>.)

(Lee et al., 2020)

Several radioisotopes of arsenic and their traces are used in medical, biomedical and environmental applications. Arsenic is versatile and is incorporated into several chemical structures that permit the synthesis of radiopharmaceuticals through possible usefulness in treatment and identification of numerous illness (Emran and Phillips, 1991).

A photosensitizer is used in combination with chemotherapy toward kill cells by ROS generation in the occurrence of oxygen and light radiation (Agostinis et al., 2011; Cheng et al., 2019; Dhas et al.,

2021a; Zhang et al., 2018; Zhou et al., 2016). The arsenical-based chemotherapy might not only encourage cell apoptosis, but also control cancer microenvironments to progress the photodynamic therapy against hypoxic tumors. A photodynamic therapy based on chemotherapy sensitization in contradiction of hypoxic tumors and demonstrated the effective drug filling of phenyl arsine oxide in the porphyrinic metal-organic structure along with surface modification of hyaluronic acid. The results demonstrated an improvement in biocompatibility and improved the special tumor buildup and exact cellular uptake of the surface-modified formulation (Yuan et al., 2020). Potassium arsenite (Fowler's solution) has been used for the management of malaria and syphilis in the late 1700s (Bjorklund et al., 2020; Drobna et al., 2009b). Numerous clinical applications for Fowler's solutions have been studied and applied over the years, but toxicities have limited their usefulness (Ho and Lowenstein, n.d.). The therapeutic effects of As included limitations of serious adverse reactions, unsatisfactory therapeutic effect and toxicity at a high level of dose (Ettlinger et al., 2019; Zhang et al., 2016).

4. REFERENCES

- Addo Ntim, S., Mitra, S., 2011. Removal of Trace Arsenic To Meet Drinking Water Standards Using Iron Oxide Coated Multiwall Carbon Nanotubes. *J. Chem. Eng. Data* 56, 2077–2083. <https://doi.org/10.1021/je1010664>
- Agostinis, P., Berg, K., Cengel, K.A., Foster, T.H., Girotti, A.W., Gollnick, S.O., Hahn, S.M., Hamblin, M.R., Juzeniene, A., Kessel, D., Korbelik, M., Moan, J., Mroz, P., Nowis, D., Piette, J., Wilson, B.C., Golab, J., 2011. Photodynamic therapy of cancer: An update. *CA. Cancer J. Clin.* 61, 250–281. <https://doi.org/10.3322/caac.20114>
- Al-jubouri, S.M., Holmes, S.M., 2020. Journal of Water Process Engineering Immobilization of cobalt ions using hierarchically porous 4A zeolite-based carbon composites : Ion-exchange and solidification. *J. Water Process Eng.* 33, 101059. <https://doi.org/10.1016/j.jwpe.2019.101059>
- Ali, H., Khan, E., 2018. What are heavy metals? Long-standing controversy over the scientific use of the term ‘heavy metals’—proposal of a comprehensive definition. *Toxicol. Environ. Chem.* 100, 6–19. <https://doi.org/10.1080/02772248.2017.1413652>
- Ali, H., Khan, E., Ilahi, I., 2019. Environmental chemistry and ecotoxicology of hazardous heavy metals: Environmental persistence, toxicity, and bioaccumulation. *J. Chem.* <https://doi.org/10.1155/2019/6730305>
- Alka, S., Shahir, S., Ibrahim, N., Ndejiko, M.J., Vo, D.V.N., Manan, F.A., 2021. Arsenic removal technologies and future trends: A mini review. *J. Clean. Prod.* 278. <https://doi.org/10.1016/j.jclepro.2020.123805>
- Allan, A.M., Hafez, A.K., Labrecque, M.T., Solomon, E.R., Shaikh, M.N., Zheng, X., Ali, A., 2015. Sex-dependent effects of developmental arsenic exposure on methylation capacity and methylation regulation of the glucocorticoid receptor system in the embryonic mouse brain. *Toxicol. Reports* 2, 1376–1390. <https://doi.org/10.1016/j.toxrep.2015.10.003>
- Andjelkovic, I., Tran, D.N.H., Kabiri, S., Azari, S., Markovic, M., Losic, D., 2015. Graphene aerogels decorated with α -FeOOH nanoparticles for efficient adsorption of arsenic from contaminated waters. *ACS Appl. Mater. Interfaces.* <https://doi.org/10.1021/acsami.5b01624>
- Andrewes, P., Kitchin, K.T., Wallace, K., 2003. Dimethylarsine and trimethylarsine are potent genotoxins in vitro. *Chem. Res. Toxicol.* 16, 994–1003. <https://doi.org/10.1021/tx034063h>
- Antman, K.H., 2001. Introduction: The History of Arsenic Trioxide in Cancer Therapy. *Oncologist* 6, 1–2. https://doi.org/10.1634/theoncologist.6-suppl_2-1
- Aposhian, H.V., 1997. Enzymatic methylation of arsenic species and other new approaches to arsenic toxicity. *Annu. Rev. Pharmacol. Toxicol.* <https://doi.org/10.1146/annurev.pharmtox.37.1.397>
- Arif, N., Yadav, V., Singh, Shweta, Singh, Swati, Ahmad, P., Mishra, R.K., Sharma, S., Tripathi, D.K., Dubey, N.K., Chauhan, D.K., 2016. Influence of high and low levels of plant-beneficial heavy metal ions on plant growth and development. *Front. Environ. Sci.* 4. <https://doi.org/10.3389/fenvs.2016.00069>
- Arora, N.K., Chauhan, R., 2021. Heavy metal toxicity and sustainable interventions for their decontamination. *Environ. Sustain.* 4,

- 1–3. <https://doi.org/10.1007/s42398-021-00164-y>
- Aung, K.H., Kurihara, R., Nakashima, S., Maekawa, F., Nohara, K., Kobayashi, T., Tsukahara, S., 2013. Inhibition of neurite outgrowth and alteration of cytoskeletal gene expression by sodium arsenite. *Neurotoxicology* 34, 226–235. <https://doi.org/10.1016/j.neuro.2012.09.008>
- Aung, K.H., Kyi-Tha-Thu, C., Sano, K., Nakamura, K., Tanoue, A., Nohara, K., Kakeyama, M., Tohyama, C., Tsukahara, S., Maekawa, F., 2016. Prenatal exposure to arsenic impairs behavioral flexibility and cortical structure in mice. *Front. Neurosci.* 10, 1–12. <https://doi.org/10.3389/fnins.2016.00137>
- Babar, N.-U.-A., Joya, K.S., Tayyab, M.A., Ashiq, M.N., Sohail, M., 2019. Highly Sensitive and Selective Detection of Arsenic Using Electrogenerated Nanotextured Gold Assemblage. *ACS Omega* 4, 13645–13657. <https://doi.org/10.1021/acsomega.9b00807>
- Balali-Mood, M., Naseri, K., Tahergorabi, Z., Khazdair, M.R., Sadeghi, M., 2021. Toxic Mechanisms of Five Heavy Metals: Mercury, Lead, Chromium, Cadmium, and Arsenic. *Front. Pharmacol.* 12, 1–19. <https://doi.org/10.3389/fphar.2021.643972>
- Bashir, S., Sharma, Y., Irshad, M., Gupta, S.D., Dogra, T.D., 2006. Arsenic-induced cell death in liver and brain of experimental rats. *Basic Clin. Pharmacol. Toxicol.* 98, 38–43. https://doi.org/10.1111/j.1742-7843.2006.pto_170.x
- Bhadauria, S., Flora, S.J.S., 2007. Response of arsenic-induced oxidative stress, DNA damage, and metal imbalance to combined administration of DMSA and monoisoamyl-DMSA during chronic arsenic poisoning in rats. *Cell Biol. Toxicol.* 23, 91–104. <https://doi.org/10.1007/s10565-006-0135-8>
- Biswas, S., Banna, H.U., Jahan, M., Anjum, A., Siddique, A.E., Roy, A., Nikkon, F., Salam, K.A., Haque, A., Himeno, S., Hossain, K., Saud, Z.A., 2020. In vivo evaluation of arsenic-associated behavioral and biochemical alterations in F0 and F1 mice. *Chemosphere* 245, 125619. <https://doi.org/10.1016/j.chemosphere.2019.125619>
- Bjørklund, G., Oliinyk, P., Lysiuk, R., Rahaman, M.S., Antonyak, H., Lozynska, I., Lenchyk, L., Peana, M., 2020. Arsenic intoxication: general aspects and chelating agents. *Arch. Toxicol.* <https://doi.org/10.1007/s00204-020-02739-w>
- Bock, J., Wainstock, T., Braun, K., Segal, M., 2015. Stress In Utero: Prenatal Programming of Brain Plasticity and Cognition. *Biol. Psychiatry* 78, 315–326. <https://doi.org/10.1016/j.biopsych.2015.02.036>
- Bodaghi-Namileh, V., Sepand, M.R., Omidi, A., Aghsami, M., Seyednejad, S.A., Kasirzadeh, S., Sabzevari, O., 2018. Acetyl-L-carnitine attenuates arsenic-induced liver injury by abrogation of mitochondrial dysfunction, inflammation, and apoptosis in rats. *Environ. Toxicol. Pharmacol.* 58, 11–20. <https://doi.org/10.1016/j.etap.2017.12.005>
- Calì, T., Ottolini, D., Brini, M., 2011. Mitochondria, calcium, and endoplasmic reticulum stress in Parkinson's disease. *BioFactors* 37, 228–240. <https://doi.org/10.1002/biof.159>
- Cavicchioli, A., La-Scalea, M.A., Gutz, I.G.R., 2004. Analysis and Speciation of Traces of Arsenic in Environmental, Food and Industrial Samples by Voltammetry: a Review.

- Electroanalysis 16, 697–711.
<https://doi.org/10.1002/elan.200302936>
- Chandravanshi, L.P., Shukla, R.K., Sultana, S., Pant, A.B., Khanna, V.K., 2014. Early life arsenic exposure and brain dopaminergic alterations in rats. *Int. J. Dev. Neurosci.* 38, 91–104. <https://doi.org/10.1016/j.ijdevneu.2014.08.009>
- Chang, F., Lee, J.T., Navolanic, P.M., Steelman, L.S., Shelton, J.G., Blalock, W.L., Franklin, R.A., McCubrey, J.A., 2003. Involvement of PI3K/Akt pathway in cell cycle progression, apoptosis, and neoplastic transformation: A target for cancer chemotherapy. *Leukemia* 17, 590–603. <https://doi.org/10.1038/sj.leu.2402824>
- Cheng, H., Fan, G.-L., Fan, J.-H., Yuan, P., Deng, F.-A., Qiu, X.-Z., Yu, X.-Y., Li, S.-Y., 2019. Epigenetics-inspired photosensitizer modification for plasma membrane-targeted photodynamic tumor therapy. *Biomaterials* 224, 119497. <https://doi.org/10.1016/j.biomaterials.2019.119497>
- Chin-Chan, M., Navarro-Yepes, J., Quintanilla-Vega, B., 2015. Environmental pollutants as risk factors for neurodegenerative disorders: Alzheimer and Parkinson diseases. *Front. Cell. Neurosci.* 9, 1–22. <https://doi.org/10.3389/fncel.2015.00124>
- Chivate, A., Garkal, A., Hariharan, K., Mehta, T., 2021. Exploring novel carrier for improving bioavailability of Itraconazole: Solid dispersion through hot-melt extrusion. *J. Drug Deliv. Sci. Technol.* 63, 102541. <https://doi.org/10.1016/j.jddst.2021.102541>
- Cholanians, A.B., Phan, A. V., Ditzel, E.J., Camenisch, T.D., Lau, S.S., Monks, T.J., 2016. Arsenic induces accumulation of α -synuclein: Implications for synucleinopathies and neurodegeneration. *Toxicol. Sci.* 153, 271–281. <https://doi.org/10.1093/toxsci/kfw117>
- Cui, X., Wakai, T., Shirai, Y., Hatakeyama, K., Hirano, S., 2006. Chronic oral exposure to inorganic arsenate interferes with methylation status of p16INK4a and RASSF1A and induces lung cancer in A/J mice. *Toxicol. Sci.* 91, 372–381. <https://doi.org/10.1093/toxsci/kfj159>
- Dai, X., Compton, R.G., 2006. Detection of As(III) via oxidation to As(V) using platinum nanoparticle modified glassy carbon electrodes: arsenic detection without interference from copper. *Analyst* 131, 516. <https://doi.org/10.1039/b513686e>
- Dai, X., Nekrassova, O., Hyde, M.E., Compton, R.G., 2004. Anodic Stripping Voltammetry of Arsenic(III) Using Gold Nanoparticle-Modified Electrodes. *Anal. Chem.* 76, 5924–5929. <https://doi.org/10.1021/ac049232x>
- Das, J., Ghosh, J., Manna, P., Sil, P.C., 2010. Protective role of taurine against arsenic-induced mitochondria-dependent hepatic apoptosis via the inhibition of PKC δ -JNK pathway. *PLoS One* 5, 1–19. <https://doi.org/10.1371/journal.pone.0012602>
- Dehghani, M.H., Taher, M.M., Bajpai, A.K., Heibati, B., Tyagi, I., Asif, M., Agarwal, S., Gupta, V.K., 2015. Removal of noxious Cr (VI) ions using single-walled carbon nanotubes and multi-walled carbon nanotubes. *Chem. Eng. J.* 279, 344–352. <https://doi.org/10.1016/j.cej.2015.04.151>
- Dhas, N., Kudarha, R., Garkal, A., Ghate, V., Sharma, S., Panzade, P., Khot, S., Chaudhari, P., Singh, A., Paryani, M., Lewis, S., Garg, N., Singh, N., Bangar, P., Mehta, T., 2021a. Molybdenum-based hetero-nanocomposites for

- cancer therapy, diagnosis and biosensing application: Current advancement and future breakthroughs. *J. Control. Release.* <https://doi.org/10.1016/j.jconrel.2020.12.015>
- Dhas, N., Mehta, T., Sharma, S., Garkal, A., Yadav, D., Hariharan, K., Shamjetshabam, B., Khot, S., Kudarha, R., Bangar, P., Arbade, G., Kalyankar, P., 2021b. Intranasal gene therapy for the treatment of neurological disorders, in: *Direct Nose-to-Brain Drug Delivery.* Elsevier, pp. 351–387. <https://doi.org/10.1016/B978-0-12-822522-6.00017-5>
- Dhas, N., Yadav, D., Singh, A., Garkal, A., Kudarha, R., Bangar, P., Savjani, J., Pardeshi, C. V., Garg, N., Mehta, T., 2021c. Direct transport theory: From the nose to the brain, in: *Direct Nose-to-Brain Drug Delivery.* Elsevier, pp. 15–37. <https://doi.org/10.1016/B978-0-12-822522-6.00001-1>
- Di Iorio, E., Colombo, C., Cheng, Z., Capitani, G., Mele, D., Ventrucci, G., Angelico, R., 2019. Characterization of magnetite nanoparticles synthesized from Fe(II)/nitrate solutions for arsenic removal from water. *J. Environ. Chem. Eng.* 7, 102986. <https://doi.org/10.1016/j.jece.2019.10.2986>
- Dilda, P.J., Hogg, P.J., 2007. Arsenical-based cancer drugs. *Cancer Treat. Rev.* 33, 542–564. <https://doi.org/10.1016/j.ctrv.2007.05.001>
- Dixit, S., Mehra, R.D., Dhar, P., 2020. Effect of α -lipoic acid on spatial memory and structural integrity of developing hippocampal neurons in rats subjected to sodium arsenite exposure. *Environ. Toxicol. Pharmacol.* 75, 103323. <https://doi.org/10.1016/j.etap.2020.10.3323>
- Drobna, Z., Styblo, M., Thomas, D.J., 2009a. An Overview of Arsenic Metabolism and Toxicity, in: *Current Protocols in Toxicology.* John Wiley & Sons, Inc., p. 4.31.1. <https://doi.org/10.1002/0471140856.tx0431s42>
- Drobna, Z., Styblo, M., Thomas, D.J., 2009b. An Overview of Arsenic Metabolism and Toxicity, in: *Current Protocols in Toxicology.* <https://doi.org/10.1002/0471140856.tx0431s42>
- Du, X., Tian, M., Wang, X., Zhang, J., Huang, Q., Liu, L., Shen, H., 2018. Cortex and hippocampus DNA epigenetic response to a long-term arsenic exposure via drinking water. *Environ. Pollut.* 234, 590–600. <https://doi.org/10.1016/j.envpol.2017.11.083>
- Emran, A.M., Phillips, D.R., 1991. Biomedical Use of Arsenic Radioisotopes. *New Trends Radiopharm. Synth. Qual. Assur. Regul. Control* 153–168. https://doi.org/10.1007/978-1-4899-0626-7_16
- Engwa, G.A., Ferdinand, P.U., Nwalo, F.N., Unachukwu, M.N., 2019. Mechanism and Health Effects of Heavy Metal Toxicity in Humans. *Mech. Heal. Eff. Heavy Met. Toxic. Humans* 23.
- Escudero-Lourdes, C., 2016. Toxicity mechanisms of arsenic that are shared with neurodegenerative diseases and cognitive impairment: Role of oxidative stress and inflammatory responses. *Neurotoxicology* 53, 223–235. <https://doi.org/10.1016/j.neuro.2016.02.002>
- Ettlinger, R., Moreno, N., Volkmer, D., Kerl, K., Bunzen, H., 2019. Zeolitic Imidazolate Framework-8 as pH-Sensitive Nanocarrier for “Arsenic Trioxide” Drug Delivery. *Chem. - A Eur. J.* 25, 13189–13196. <https://doi.org/10.1002/chem.201902599>
- Fatema, K., Shoily, S.S., Ahsan, T., Haidar,

- Z., Sumit, A.F., Sajib, A.A., 2021. Effects of arsenic and heavy metals on metabolic pathways in cells of human origin: Similarities and differences. *Toxicol. Reports* 8, 1109–1120. <https://doi.org/10.1016/j.toxrep.2021.05.015>
- Feeney, Kounaves, 2000. On-site analysis of arsenic in groundwater using a microfabricated gold ultramicroelectrode array. *Anal. Chem.* 72, 2222–8. <https://doi.org/10.1021/ac991185z>
- Felix, K., Manna, S.K., Wise, K., Barr, J., Ramesh, G.T., 2005. Low levels of arsenite activates nuclear factor- κ B and activator protein-1 in immortalized mesencephalic cells. *J. Biochem. Mol. Toxicol.* 19, 67–77. <https://doi.org/10.1002/jbt.20062>
- Firkin, F., Iland, H., 2013. protective effect of quercetin 1–16.
- Forsberg, G., O’Laughlin, J.W., Megargle, R.G., Koirtiyham, S.R., 1975. Determination of arsenic by anodic stripping voltammetry and differential pulse anodic stripping voltammetry. *Anal. Chem.* 47, 1586–1592. <https://doi.org/10.1021/ac60359a057>
- Frankel, S., Concannon, J., Brusky, K., Pietrowicz, E., Giorgianni, S., Thompson, W.D., Currie, D.A., 2009. Arsenic exposure disrupts neurite growth and complexity in vitro. *Neurotoxicology* 30, 529–537. <https://doi.org/10.1016/j.neuro.2009.02.015>
- Fu, Z., Xi, S., 2020. The effects of heavy metals on human metabolism. *Toxicol. Mech. Methods* 30, 167–176. <https://doi.org/10.1080/15376516.2019.1701594>
- Fukushima, M., Yanagi, H., Hayashi, S., Sugauma, N., Taniguchi, Y., 2003. Fabrication of gold nanoparticles and their influence on optical properties of dye-doped sol-gel films. *Thin Solid Films* 438–439, 39–43. [https://doi.org/10.1016/S0040-6090\(03\)00750-8](https://doi.org/10.1016/S0040-6090(03)00750-8)
- Garkal, A., Avachat, A., 2022. Development and in-vitro in-vivo characterization of in-situ gelling sustained-release nevirapine suspension. *J. Drug Deliv. Sci. Technol.* 67, 102938. <https://doi.org/10.1016/j.jddst.2021.102938>
- Gonzalez, B., Heijman, S.G.J., Rietveld, L.C., van Halem, D., 2019. Arsenic removal from geothermal influenced groundwater with low pressure NF pilot plant for drinking water production in Nicaraguan rural communities. *Sci. Total Environ.* 667, 297–305. <https://doi.org/10.1016/j.scitotenv.2019.02.222>
- Guha Mazumder, D., Dasgupta, U.B., 2011. Chronic arsenic toxicity: Studies in West Bengal, India. *Kaohsiung J. Med. Sci.* 27, 360–370. <https://doi.org/10.1016/j.kjms.2011.05.003>
- He, J., Wang, M., Jiang, Y., Chen, Q., Xu, S., Xu, Q., Jiang, B.H., Liu, L.Z., 2014. Chronic arsenic exposure and angiogenesis in human bronchial epithelial Cells via the ROS/miR-199a-5p/HIF-1 α /COX-2 pathway. *Environ. Health Perspect.* 122, 255–261. <https://doi.org/10.1289/ehp.1307545>
- He, Z.L., Yang, X.E., Stoffella, P.J., 2005. Trace elements in agroecosystems and impacts on the environment. *J. Trace Elem. Med. Biol.* 19, 125–140. <https://doi.org/10.1016/j.jtemb.2005.02.010>
- Healy, S.M., Casarez, E.A., Ayala-Fierro, F., Aposhian, H.V., 1998. Enzymatic methylation of arsenic compounds. V. Arsenite methyltransferase activity in tissues of mice. *Toxicol. Appl. Pharmacol.* 148, 65–70. <https://doi.org/10.1006/taap.1997.8306>
- Ho, D., Lowenstein, E.J., n.d. Fowler’s Solution and the Evolution of the Use of Arsenic in Modern Medicine.

- Skinmed 14, 287–289.
- Htway, S.M., Sein, M.T., Nohara, K., Win-Shwe, T.T., 2019. Effects of developmental arsenic exposure on the social behavior and related gene expression in C3H adult male mice. *Int. J. Environ. Res. Public Health* 16. <https://doi.org/10.3390/ijerph16020174>
- Hu, S., Lu, J., Jing, C., 2012. A novel colorimetric method for field arsenic speciation analysis. *J. Environ. Sci.* 24, 1341–1346. [https://doi.org/10.1016/S1001-0742\(11\)60922-4](https://doi.org/10.1016/S1001-0742(11)60922-4)
- Hu, Y., Yu, C., Yao, M., Wang, L., Liang, B., Zhang, B., Huang, X., Zhang, A., 2018. The PKC δ -Nrf2-ARE signalling pathway may be involved in oxidative stress in arsenic-induced liver damage in rats. *Environ. Toxicol. Pharmacol.* 62, 79–87. <https://doi.org/10.1016/j.etap.2018.05.012>
- Hughes, M.F., Devesa, V., Adair, B.M., Conklin, S.D., Creed, J.T., Styblo, M., Kenyon, E.M., Thomas, D.J., 2008. Tissue dosimetry, metabolism and excretion of pentavalent and trivalent dimethylated arsenic in mice after oral administration. *Toxicol. Appl. Pharmacol.* 227, 26–35. <https://doi.org/10.1016/j.taap.2007.10.011>
- Inesta-Vaquera, F., Navasumrit, P., Henderson, C.J., Frangova, T.G., Honda, T., Dinkova-Kostova, A.T., Ruchirawat, M., Wolf, C.R., 2021. Application of the in vivo oxidative stress reporter Hmox1 as mechanistic biomarker of arsenic toxicity. *Environ. Pollut.* 270, 116053. <https://doi.org/10.1016/j.envpol.2020.116053>
- Ivandini, T.A., Sato, R., Makide, Y., Fujishima, A., Einaga, Y., 2006. Electrochemical Detection of Arsenic(III) Using Iridium-Implanted Boron-Doped Diamond Electrodes. *Anal. Chem.* 78, 6291–6298. <https://doi.org/10.1021/ac0519514>
- Jadhav, S. V., Bringas, E., Yadav, G.D., Rathod, V.K., Ortiz, I., Marathe, K. V., 2015. Arsenic and fluoride contaminated groundwaters: A review of current technologies for contaminants removal. *J. Environ. Manage.* 162, 306–325. <https://doi.org/10.1016/j.jenvman.2015.07.020>
- Jomova, K., Jenisova, Z., Feszterova, M., Baros, S., Liska, J., Hudecova, D., Rhodes, C.J., Valko, M., 2011. Arsenic: Toxicity, oxidative stress and human disease. *J. Appl. Toxicol.* 31, 95–107. <https://doi.org/10.1002/jat.1649>
- Jou, Y., Wang, S., Dai, Y., Chen, S., Shen, C., Lee, Y., Chen, L., Liu, Y., 2019. Gene expression and DNA methylation regulation of arsenic in mouse bladder tissues and in human urothelial cells. *Oncol. Rep.* <https://doi.org/10.3892/or.2019.7235>
- Joya, K.S., de Groot, H.J.M., 2016. Controlled Surface-Assembly of Nanoscale Leaf-Type Cu-Oxide Electrocatalyst for High Activity Water Oxidation. *ACS Catal.* 6, 1768–1771. <https://doi.org/10.1021/acscatal.5b02950>
- Karakurt, Sevtap, Pehlivan, E., Karakurt, Serdar, 2019. Removal of Carcinogenic Arsenic from Drinking Water By the Application of Ion Exchange Resins. *Oncogen* 2, 1–8. <https://doi.org/10.35702/onc.10005>
- Katsoyiannis, I.A., Zouboulis, A.I., 2006. Comparative Evaluation of Conventional and Alternative Methods for the Removal of Arsenic from Contaminated Groundwaters. *Rev. Environ. Health* 21. <https://doi.org/10.1515/REVEH.2006.21.1.25>
- Kim, S.J., Kim, E.S., Kim, S., Uhm, J., Won, Y.W., Park, B.B., Choi, J.H., Lee, Y.Y., 2017. Antitumoral effect of arsenic compound, sodium

- metaarsenite (KML001), on multiple myeloma cells. *Int. J. Oncol.* 51, 1739–1746.
<https://doi.org/10.3892/ijo.2017.4161>
- Kumar, M., Rao T., S., Isloor, A.M., Ibrahim, G.P.S., Inamuddin, Ismail, N., Ismail, A.F., Asiri, A.M., 2019. Use of cellulose acetate/polyphenylsulfone derivatives to fabricate ultrafiltration hollow fiber membranes for the removal of arsenic from drinking water. *Int. J. Biol. Macromol.* 129, 715–727.
<https://doi.org/10.1016/j.ijbiomac.2019.02.017>
- Kumar, R., Patel, M., Singh, P., Bundschuh, J., Pittman, C.U., Trakal, L., Mohan, D., 2019. Emerging technologies for arsenic removal from drinking water in rural and peri-urban areas: Methods, experience from, and options for Latin America. *Sci. Total Environ.* 694, 133427.
<https://doi.org/10.1016/j.scitotenv.2019.07.233>
- Kwang-Seok Yun, Hong-Jeong Kim, Segyeong Joo, Juhyoung Kwak, Euisik Yoon, n.d. Analysis of heavy-metal-ions using mercury microelectrodes and a solid-state reference electrode fabricated on a Si wafer, in: *Digest of Papers Microprocesses and Nanotechnology 2000*. 2000 International Microprocesses and Nanotechnology Conference (IEEE Cat. No.00EX387). Japan Soc. Appl. Phys, pp. 72–73.
<https://doi.org/10.1109/IMNC.2000.872628>
- Le, X.C., Yalcin, S., Ma, M., 2000. Speciation of Submicrogram per Liter Levels of Arsenic in Water: On-Site Species Separation Integrated with Sample Collection. *Environ. Sci. Technol.* 34, 2342–2347.
<https://doi.org/10.1021/es991203u>
- Lee, E.J., Sung, J.Y., Koo, K.H., Park, J.B., Kim, D.H., Shim, J., Lee, C.H., Park, J., Kim, Y.N., 2020. Anti-tumor effects of sodium meta-arsenite in glioblastoma cells with higher akt activities. *Int. J. Mol. Sci.* 21, 1–19.
<https://doi.org/10.3390/ijms21238982>
- Leslie, E.M., Haimeur, A., Waalkes, M.P., 2004. Arsenic Transport by the Human Multidrug Resistance Protein 1 (MRP1/ABCC1). *J. Biol. Chem.* 279, 32700–32708.
<https://doi.org/10.1074/jbc.M404912200>
- Leu, L., Mohassel, L., 2009. Arsenic trioxide as first-line treatment for acute promyelocytic leukemia. *Am. J. Heal. Pharm.* 66, 1913–1918.
<https://doi.org/10.2146/ajhp080342>
- Li, D., Wei, Y., Xu, S., Niu, Q., Zhang, M., Li, S., Jing, M., 2018. A systematic review and meta-analysis of bidirectional effect of arsenic on ERK signaling pathway. *Mol. Med. Rep.*
<https://doi.org/10.3892/mmr.2018.8383>
- Li, Y., Wang, M., Piao, F., Wang, X., 2012. Subchronic exposure to arsenic inhibits spermatogenesis and downregulates the expression of Ddx3y in testis and epididymis of mice. *Toxicol. Sci.* 128, 482–489.
<https://doi.org/10.1093/toxsci/kfs169>
- Liu, C.H., Chuang, Y.H., Chen, T.Y., Tian, Y., Li, H., Wang, M.K., Zhang, W., 2015. Mechanism of Arsenic Adsorption on Magnetite Nanoparticles from Water: Thermodynamic and Spectroscopic Studies. *Environ. Sci. Technol.* 49, 7726–7734.
<https://doi.org/10.1021/acs.est.5b00381>
- Liu, L., Luo, X.B., Ding, L., Luo, S.L., 2018. Application of Nanotechnology in the Removal of Heavy Metal From Water, Nanomaterials for the Removal of Pollutants and Resource Reutilization. Elsevier Inc.
<https://doi.org/10.1016/B978-0-12-814837-2.00004-4>
- Liu, Z.-G., Chen, X., Jia, Y., Liu, J.-H., Huang, X.-J., 2014. Role of Fe(III) in preventing humic interference during

- As(III) detection on gold electrode: Spectroscopic and voltammetric evidence. *J. Hazard. Mater.* 267, 153–160.
<https://doi.org/10.1016/j.jhazmat.2013.12.054>
- Long, H., Zheng, Y.J., Peng, Y.L., Jin, G.Z., Deng, W.H., Zhang, S.C., 2019. Study on arsenic removal in aqueous chloride solution with lead oxide. *Int. J. Environ. Sci. Technol.* 16, 6999–7010. <https://doi.org/10.1007/s13762-018-2158-0>
- Ma, M.D., Wu, H., Deng, Z.Y., Zhao, X., 2018. Arsenic removal from water by nanometer iron oxide coated single-wall carbon nanotubes. *J. Mol. Liq.* 259, 369–375.
<https://doi.org/10.1016/j.molliq.2018.03.052>
- Maeda, H., Hori, S., Ohizumi, H., Segawa, T., Kakehi, Y., Ogawa, O., Kakizuka, A., 2004. Effective treatment of advanced solid tumors by the combination of arsenic trioxide and L-buthionine-sulfoximine. *Cell Death Differ.* 11, 737–746.
<https://doi.org/10.1038/sj.cdd.4401389>
- Maekawa, F., Tsuboi, T., Oya, M., Aung, K.H., Tsukahara, S., Pellerin, L., Nohara, K., 2013. Effects of sodium arsenite on neurite outgrowth and glutamate AMPA receptor expression in mouse cortical neurons. *Neurotoxicology* 37, 197–206.
<https://doi.org/10.1016/j.neuro.2013.05.006>
- Mann, K.K., Colombo, M., Miller, W.H., 2008. Arsenic trioxide decreases AKT protein in a caspase-dependent manner. *Mol. Cancer Ther.* 7, 1680–1687. <https://doi.org/10.1158/1535-7163.MCT-07-2164>
- Manoj, S.R., Karthik, C., Kadirvelu, K., Arulselvi, P.I., Shanmugasundaram, T., Bruno, B., Rajkumar, M., 2020. Understanding the molecular mechanisms for the enhanced phytoremediation of heavy metals through plant growth promoting rhizobacteria: A review. *J. Environ. Manage.* 254, 109779.
<https://doi.org/10.1016/j.jenvman.2019.109779>
- Marafante, E., Vahter, M., Envall, J., 1985. The role of the methylation in the detoxication of arsenate in the rabbit. *Chem. Biol. Interact.* 56, 225–238.
[https://doi.org/10.1016/0009-2797\(85\)90008-0](https://doi.org/10.1016/0009-2797(85)90008-0)
- Marzo, T., La Mendola, D., 2021. Strike a balance: between metals and non-metals, metalloids as a source of anti-infective agents. *Inorganics* 9. <https://doi.org/10.3390/inorganics9060046>
- Masindi, V., Muedi, K.L., 2018. Environmental Contamination by Heavy Metals. *Heavy Met.* <https://doi.org/10.5772/intechopen.76082>
- Mathews, V., Chandy, M., Srivastava, A., 2001. leukaemia 14.
- Mochizuki, H., 2019. Arsenic neurotoxicity in humans. *Int. J. Mol. Sci.* 20. <https://doi.org/10.3390/ijms20143418>
- Mohammed Abdul, K.S., Jayasinghe, S.S., Chandana, E.P.S., Jayasumana, C., De Silva, P.M.C.S., 2015. Arsenic and human health effects: A review. *Environ. Toxicol. Pharmacol.* 40, 828–846.
<https://doi.org/10.1016/j.etap.2015.09.016>
- Mukherjee, S., Mukhopadhyay, P.K., 2009. Studies on Arsenic Toxicity in Male Rat Gonads and its Protection by High Dietary Protein Supplementation. *Al Ameen J. Med. Sci.* 2, 73–77.
- Muthumani, M., Prabu, S.M., 2012. Silibinin potentially protects arsenic-induced oxidative hepatic dysfunction in rats. *Toxicol. Mech. Methods* 22, 277–288.
<https://doi.org/10.3109/15376516.2011.647113>
- NAISU, 2003. Arsenic 2002: An overview of Arsenic Issues and Mitigation Initiatives in Bangladesh. NGOs

- Arsen. Inf. Support Unit NGO Forum Drink. Water Supply Sanit. 1–125.
- Ng, J.C., 2005. Environmental contamination of arsenic and its toxicological impact on humans. *Environ. Chem.* 2, 146–160. <https://doi.org/10.1071/EN05062>
- Niño, S.A., Morales-Martínez, A., Chi-Ahumada, E., Carrizales, L., Salgado-Delgado, R., Pérez-Severiano, F., Díaz-Cintra, S., Jiménez-Capdeville, M.E., Zarazúa, S., 2019. Arsenic Exposure Contributes to the Bioenergetic Damage in an Alzheimer's Disease Model. *ACS Chem. Neurosci.* 10, 323–336. <https://doi.org/10.1021/acchemneuro.8b00278>
- Nriagu, J.O., 1989. A global assessment of natural sources of atmospheric trace metals. *Nature.* <https://doi.org/10.1038/338047a0>
- Nutrition, Arsenic and Cognitive Function in Children - Tabular View - ClinicalTrials.gov [WWW Document], n.d.
- Patlolla, A.K., Tchounwou, P.B., 2005. Serum acetyl cholinesterase as a biomarker of arsenic induced neurotoxicity in Sprague-Dawley rats. *Int. J. Environ. Res. Public Health* 2, 80–83. <https://doi.org/10.3390/ijerph2005010080>
- Patti, F., Fiore, M., Chisari, C.G., D'Amico, E., Lo Fermo, S., Toscano, S., Copat, C., Ferrante, M., Zappia, M., 2020. CSF neurotoxic metals/metalloids levels in amyotrophic lateral sclerosis patients: comparison between bulbar and spinal onset. *Environ. Res.* 188, 109820. <https://doi.org/10.1016/j.envres.2020.109820>
- Piyajit Watcharasit, 1, Satayavivad, * Apinya Thiantanawat1 and Jutamaad, 2007. GSK3 promotes arsenite-induced apoptosis via facilitation of mitochondria disruption. *J. Appl. Toxicol.* 27, 511–518. <https://doi.org/10.1002/jat>
- Prakash, C., Chhikara, S., Kumar, V., 2022. Mitochondrial Dysfunction in Arsenic-Induced Hepatotoxicity: Pathogenic and Therapeutic Implications. *Biol. Trace Elem. Res.* 200, 261–270. <https://doi.org/10.1007/s12011-021-02624-2>
- Prakash, C., Kumar, V., 2016. Chronic Arsenic Exposure-Induced Oxidative Stress is Mediated by Decreased Mitochondrial Biogenesis in Rat Liver. *Biol. Trace Elem. Res.* 173, 87–95. <https://doi.org/10.1007/s12011-016-0622-6>
- Prakash, C., Soni, M., Kumar, V., 2016. Mitochondrial oxidative stress and dysfunction in arsenic neurotoxicity: A review. *J. Appl. Toxicol.* 36, 179–188. <https://doi.org/10.1002/jat.3256>
- Pramod, L., Gandhimathi, R., Lavanya, A., Ramesh, S.T., Nidheesh, P. V., 2020. Heterogeneous Fenton process coupled with microfiltration for the treatment of water with higher arsenic content. *Chem. Eng. Commun.* 207, 1646–1657. <https://doi.org/10.1080/00986445.2019.1674814>
- Rachman, T., 2018. Mechanisms of Arsenic Toxicity in Humans. *Angew. Chemie Int. Ed.* 6(11), 951–952. 10–27.
- Rahaman, M.S., Rahman, M.M., Mise, N., Sikder, M.T., Ichihara, G., Uddin, M.K., Kurasaki, M., Ichihara, S., 2021. Environmental arsenic exposure and its contribution to human diseases, toxicity mechanism and management. *Environ. Pollut.* 289, 117940. <https://doi.org/10.1016/j.envpol.2021.117940>
- Rahman, M.R., Okajima, T., Ohsaka, T., 2010. Selective Detection of As(III) at the Au(111)-like Polycrystalline Gold Electrode. *Anal. Chem.* 82, 9169–9176. <https://doi.org/10.1021/ac101206j>
- Ratna Kumar, P., Chaudhari, S., Khilar,

- K.C., Mahajan, S., 2004. Removal of arsenic from water by electrocoagulation. *Chemosphere* 55, 1245–1252.
<https://doi.org/10.1016/j.chemosphere.2003.12.025>
- Ratnaike, R.N., 2003. Acute and chronic arsenic toxicity. *Postgr. Med J* 391–397.
- Rehman, K., Fatima, F., Akash, M.S.H., 2019. Biochemical investigation of association of arsenic exposure with risk factors of diabetes mellitus in Pakistani population and its validation in animal model. *Environ. Monit. Assess.* 191, 511.
<https://doi.org/10.1007/s10661-019-7670-2>
- Rehman, K., Fatima, F., Waheed, I., Akash, M.S.H., 2018. Prevalence of exposure of heavy metals and their impact on health consequences. *J. Cell. Biochem.* 119, 157–184.
<https://doi.org/10.1002/jcb.26234>
- Renu, K., Saravanan, A., Elangovan, A., Ramesh, S., Annamalai, S., Namachivayam, A., Abel, P., Madhyastha, H., Madhyastha, R., Maruyama, M., Balachandar, V., Valsala Gopalakrishnan, A., 2020. An appraisal on molecular and biochemical signalling cascades during arsenic-induced hepatotoxicity. *Life Sci.* 260, 118438.
<https://doi.org/10.1016/j.lfs.2020.118438>
- Rosenblatt, A.E., Burnstein, K.L., 2009. Inhibition of androgen receptor transcriptional activity as a novel mechanism of action of arsenic. *Mol. Endocrinol.* 23, 412–421.
<https://doi.org/10.1210/me.2008-0235>
- Saint-Jacques, N., Parker, L., Brown, P., Dummer, T.J., 2014. Arsenic in drinking water and urinary tract cancers: A systematic review of 30 years of epidemiological evidence. *Environ. Heal. A Glob. Access Sci. Source* 13, 44.
<https://doi.org/10.1186/1476-069X-13-44>
- Salimi, A., Hyde, M.E., Banks, C.E., Compton, R.G., 2004. Boron doped diamond electrode modified with iridium oxide for amperometric detection of ultra trace amounts of arsenic(III). *Analyst* 129, 9.
<https://doi.org/10.1039/b312285a>
- Sánchez-Peña, L.C., Petrosyan, P., Morales, M., González, N.B., Gutiérrez-Ospina, G., Del Razo, L.M., Gonshebat, M.E., 2010. Arsenic species, AS3MT amount, and AS3MT gene expression in different brain regions of mouse exposed to arsenite. *Environ. Res.* 110, 428–434.
<https://doi.org/10.1016/j.envres.2010.01.007>
- Sarankumar, R.K., Selvi, A., Murugan, K., Rajasekar, A., 2020. Electrokinetic (EK) and Bio-electrokinetic (BEK) Remediation of Hexavalent Chromium in Contaminated Soil Using Alkaliphilic Bio-anolyte. *Indian Geotech. J.* 50, 330–338.
<https://doi.org/10.1007/s40098-019-00366-6>
- Sarkar, A., Paul, B., 2016. The global menace of arsenic and its conventional remediation - A critical review. *Chemosphere* 158, 37–49.
<https://doi.org/10.1016/j.chemosphere.2016.05.043>
- Senuma, M., Mori, C., Ogawa, T., Kuwagata, M., 2014. Prenatal sodium arsenite affects early development of serotonergic neurons in the fetal rat brain. *Int. J. Dev. Neurosci.* 38, 204–212.
<https://doi.org/10.1016/j.ijdevneu.2014.09.005>
- Shafiuddin Ahmed, A.S., Sultana, S., Habib, A., Ullah, H., Musa, N., Mahfujur Rahman, M., Shafiqul Islam Sarker, M., 2019. Bioaccumulation of heavy metals in commercially important fish species from the tropical river estuary suggests higher potential child health risk than adults. *bioRxiv* 1–21.

- <https://doi.org/10.1101/681478>
Shankar, S., Shanker, U., Shikha, 2014. Arsenic contamination of groundwater: a review of sources, prevalence, health risks, and strategies for mitigation. *ScientificWorldJournal*. 2014, 304524.
<https://doi.org/10.1155/2014/304524>
- Shao, Y., Zhao, H., Wang, Y., Liu, J., Li, J., Chai, H., Xing, M., 2018. Arsenic and/or copper caused inflammatory response via activation of inducible nitric oxide synthase pathway and triggered heat shock protein responses in testis tissues of chicken. *Environ. Sci. Pollut. Res.* 25, 7719–7729.
<https://doi.org/10.1007/s11356-017-1042-7>
- Sharma, A., Kumar, S., 2019. Arsenic exposure with reference to neurological impairment: An overview. *Rev. Environ. Health*. <https://doi.org/10.1515/reveh-2019-0052>
- Shen, L.-L., Zhang, G.-R., Li, W., Biesalski, M., Etzold, B.J.M., 2017. Modifier-Free Microfluidic Electrochemical Sensor for Heavy-Metal Detection. *ACS Omega* 2, 4593–4603.
<https://doi.org/10.1021/acsomega.7b00611>
- Shi, J., Cai, Y., 2020. Environmental chemistry and toxicology of heavy metals. *Ecotoxicol. Environ. Saf.* 202, 110926.
<https://doi.org/10.1016/j.ecoenv.2020.110926>
- Silman, I., Sussman, J.L., 2005. Acetylcholinesterase: “Classical” and “non-classical” functions and pharmacology. *Curr. Opin. Pharmacol.* 5, 293–302.
<https://doi.org/10.1016/j.coph.2005.01.014>
- Simm, A., Banks, C., Compton, R., 2005. Sonoelectroanalytical Detection of Ultra-Trace Arsenic. *Electroanalysis* 17, 335–342.
<https://doi.org/10.1002/elan.200403110>
- Simm, A.O., Banks, C.E., Compton, R.G., 2005. The Electrochemical Detection of Arsenic(III) at a Silver Electrode. *Electroanalysis* 17, 1727–1733.
<https://doi.org/10.1002/elan.200503299>
- Simm, A.O., Banks, C.E., Compton, R.G., 2004. Sonically Assisted Electroanalytical Detection of Ultratrace Arsenic. *Anal. Chem.* 76, 5051–5055.
<https://doi.org/10.1021/ac049331a>
- Singh, R., Gautam, N., Mishra, A., Gupta, R., 2011. Heavy metals and living systems: An overview. *Indian J. Pharmacol.* 43, 246–253.
<https://doi.org/10.4103/0253-7613.81505>
- Srivastava, P., Yadav, R.S., Chandravanshi, L.P., Shukla, R.K., Dhuriya, Y.K., Chauhan, L.K.S., Dwivedi, H.N., Pant, A.B., Khanna, V.K., 2014. Unraveling the mechanism of neuroprotection of curcumin in arsenic induced cholinergic dysfunctions in rats. *Toxicol. Appl. Pharmacol.* 279, 428–440.
<https://doi.org/10.1016/j.taap.2014.06.006>
- Sun, X., Li, S., He, Y., Zhao, H., Wang, Y., Zeng, X., Xing, M., 2017. Arsenic-induced testicular toxicity in *Gallus gallus*: Expressions of inflammatory cytokines and heat shock proteins. *Poult. Sci.* 96, 3399–3406.
<https://doi.org/10.3382/ps/pex073>
- Sung, K., Kim, M., Kim, H., Hwang, G.W., Kim, K., 2019. Perinatal Exposure to Arsenic in Drinking Water Alters Glutamatergic Neurotransmission in the Striatum of C57BL/6 Mice. *Biol. Trace Elem. Res.* 187, 224–229.
<https://doi.org/10.1007/s12011-018-1374-2>
- Susan, A., Rajendran, K., Sathyasivam, K., Krishnan, U.M., 2019. An overview of plant-based interventions to ameliorate arsenic toxicity. *Biomed.*

- Pharmacother. 109, 838–852.
<https://doi.org/10.1016/j.biopha.2018.10.099>
- Tan, Y., Li, Y., Zhu, D., 2002. Fabrication of Gold Nanoparticles Using a Trithiol (Thiocyanuric Acid) as the Capping Agent. *Langmuir* 18, 3392–3395.
<https://doi.org/10.1021/la011612f>
- Tchounwou, P.B., Yedjou, C.G., Patlolla, A.K., Sutton, D.J., 2012. Heavy Metals Toxicity and the Environment, Molecular, Clinical and Environmental Toxicology. <https://doi.org/10.1007/978-3-7643-8340-4>
- Tian, W., Wang, Z., Tang, N.N., Li, J.T., Liu, Y., Chu, W.F., Yang, B.F., 2020. Ascorbic Acid Sensitizes Colorectal Carcinoma to the Cytotoxicity of Arsenic Trioxide via Promoting Reactive Oxygen Species-Dependent Apoptosis and Pyroptosis. *Front. Pharmacol.* 11, 1–13.
<https://doi.org/10.3389/fphar.2020.00123>
- Tolins, M., Ruchirawat, M., Landrigan, P., 2014. The developmental neurotoxicity of arsenic: Cognitive and behavioral consequences of early life exposure. *Ann. Glob. Heal.* 80, 303–314.
<https://doi.org/10.1016/j.aogh.2014.09.005>
- Tyler, C.R., Labrecque, M.T., Solomon, E.R., Guo, X., Allan, A.M., 2017. Prenatal arsenic exposure alters REST/NRSF and microRNA regulators of embryonic neural stem cell fate in a sex-dependent manner. *Neurotoxicol. Teratol.* 59, 1–15.
<https://doi.org/10.1016/j.ntt.2016.10.004>
- Ungureanu, G., Santos, S., Boaventura, R., Botelho, C., 2015. Arsenic and antimony in water and wastewater: Overview of removal techniques with special reference to latest advances in adsorption. *J. Environ. Manage.* 151, 326–342.
<https://doi.org/10.1016/j.jenvman.2014.12.051>
- Vadahanambi, S., Lee, S.H., Kim, W.J., Oh, I.K., 2013. Arsenic removal from contaminated water using three-dimensional graphene-carbon nanotube-iron oxide nanostructures. *Environ. Sci. Technol.* 47, 10510–10517.
<https://doi.org/10.1021/es401389g>
- Vahidnia, A., Romijn, F., Tiller, M., Van Der Voet, G.B., De Wolff, F.A., 2006. Arsenic-induced toxicity: Effect on protein composition in sciatic nerve. *Hum. Exp. Toxicol.* 25, 667–674.
<https://doi.org/10.1177/0960327106070671>
- Vahidnia, A., Romijn, F., van der Voet, G.B., de Wolff, F.A., 2008a. Arsenic-induced neurotoxicity in relation to toxicokinetics: Effects on sciatic nerve proteins. *Chem. Biol. Interact.* 176, 188–195.
<https://doi.org/10.1016/j.cbi.2008.07.001>
- Vahidnia, A., van der Straaten, R.J.H.M., Romijn, F., van Pelt, J., van der Voet, G.B., de Wolff, F.A., 2008b. Mechanism of arsenic-induced neurotoxicity may be explained through cleavage of p35 to p25 by calpain. *Toxicol. Vitro.* 22, 682–687.
<https://doi.org/10.1016/j.tiv.2007.12.010>
- Vahidnia, A., Van Der Voet, G.B., De Wolff, F.A., 2007. Arsenic neurotoxicity - A review. *Hum. Exp. Toxicol.* 26, 823–832.
<https://doi.org/10.1177/0960327107084539>
- Valles, S., Hernández-Sánchez, J., Dipp, V.R., Huerta-González, D., Olivares-Bañuelos, T.N., González-Fraga, J., Bardullas, U., 2020. Exposure to low doses of inorganic arsenic induces transgenerational changes on behavioral and epigenetic markers in zebrafish (*Danio rerio*). *Toxicol. Appl. Pharmacol.* 396, 115002.
<https://doi.org/10.1016/j.taap.2020.115002>

- Wang, C.H., Jeng, J.S., Yip, P.K., Chen, C.L., Hsu, L.I., Hsueh, Y.M., Chiou, H.Y., Wu, M.M., Chen, C.J., 2002. Biological gradient between long-term arsenic exposure and carotid atherosclerosis. *Circulation* 105, 1804–1809. <https://doi.org/10.1161/01.CIR.000015862.64816.B2>
- Wang, D., Zhao, Y., Jin, H., Zhuang, J., Zhang, W., Wang, S., Wang, J., 2013. Synthesis of Au-Decorated Tripod-Shaped Te Hybrids for Applications in the Ultrasensitive Detection of Arsenic. *ACS Appl. Mater. Interfaces* 5, 5733–5740. <https://doi.org/10.1021/am401205w>
- Wang, X., Meng, D., Chang, Q., Pan, J., Zhang, Z., Chen, G., Ke, Z., Luo, J., Shi, X., 2010. Arsenic inhibits neurite outgrowth by inhibiting the LKB1-AMPK signaling pathway. *Environ. Health Perspect.* 118, 627–634. <https://doi.org/10.1289/ehp.0901510>
- Wang, Y., Yang, X., Yu, H., Wang, H., Qi, Y., Geng, M., 2020. Effects of arsenic exposure on d-serine metabolism in the hippocampus of offspring mice at different developmental stages. *Arch. Toxicol.* 94, 77–87. <https://doi.org/10.1007/s00204-019-02616-1>
- Watanabe, T., Hirano, S., 2013. Metabolism of arsenic and its toxicological relevance. *Arch. Toxicol.* 87, 969–979. <https://doi.org/10.1007/s00204-012-0904-5>
- Welch, C.M., Nekrassova, O., Dai, X., Hyde, M.E., Compton, R.G., 2004. Fabrication, characterisation and voltammetric studies of gold amalgam nanoparticle modified electrodes. *Chemphyschem* 5, 1405–10. <https://doi.org/10.1002/cphc.200400263>
- WHO, 1996. Trace elements in human nutrition and health World Health Organization. World Heal. Organ.
- Wise, J.P., Young, J.L., Cai, J., Cai, L., 2022. Current understanding of hexavalent chromium [Cr(VI)] neurotoxicity and new perspectives. *Environ. Int.* 158, 1–43. <https://doi.org/10.1016/j.envint.2021.106877>
- Yang, M., Chen, X., Jiang, T.-J., Guo, Z., Liu, J.-H., Huang, X.-J., 2016. Electrochemical Detection of Trace Arsenic(III) by Nanocomposite of Nanorod-Like α -MnO₂ Decorated with ~5 nm Au Nanoparticles: Considering the Change of Arsenic Speciation. *Anal. Chem.* 88, 9720–9728. <https://doi.org/10.1021/acs.analchem.6b02629>
- Yoon, J.S., Hwang, D.W., Kim, E.S., Kim, J.S., Kim, S., Chung, H.J., Lee, S.K., Yi, J.H., Uhm, J., Won, Y.W., Park, B.B., Choi, J.H., Lee, Y.Y., 2016. Anti-tumoral effect of arsenic compound, sodium metaarsenite (KML001), in non-Hodgkin's lymphoma: an in vitro and in vivo study. *Invest. New Drugs* 34, 1–14. <https://doi.org/10.1007/s10637-015-0301-z>
- Yu, B., Yuan, B., Li, J.Z., Kiyomi, A., Kikuchi, H., Hayashi, H., Hu, X., Okazaki, M., Sugiura, M., Hirano, T., Fan, Y., Pei, X., Takagi, N., 2020. JNK and Autophagy Independently Contributed to Cytotoxicity of Arsenite combined With Tetrandrine via Modulating Cell Cycle Progression in Human Breast Cancer Cells. *Front. Pharmacol.* 11, 1–15. <https://doi.org/10.3389/fphar.2020.01087>
- Yuan, P., Fan, G.L., Zhao, L.P., Liu, L.S., Deng, F.A., Jiang, X.Y., Hu, A.H., Yu, X.Y., Chen, A.L., Cheng, H., Li, S.Y., 2020. Tumor targeted self-synergistic nanoplatforams for arsenic-sensitized photodynamic therapy. *Acta Biomater.* 117, 349–360. <https://doi.org/10.1016/j.actbio.2020.09.047>
- Zhang, J., Jiang, C., Figueiró Longo, J.P.,

- Azevedo, R.B., Zhang, H., Muehlmann, L.A., 2018. An updated overview on the development of new photosensitizers for anticancer photodynamic therapy. *Acta Pharm. Sin. B* 8, 137–146. <https://doi.org/10.1016/j.apsb.2017.09.003>
- Zhang, L., Zhou, Y., Kong, J., Zhang, Li, Yuan, M., Xian, S., Wang, Y., Cheng, Y., Yang, X., 2020. Effect of arsenic trioxide on cervical cancer and its mechanisms. *Exp. Ther. Med.* 20, 1–1. <https://doi.org/10.3892/etm.2020.9299>
- Zhang, R.Y., Tu, J.B., Ran, R.T., Zhang, W.X., Tan, Q., Tang, P., Kuang, T., Cheng, S.Q., Chen, C.Z., Jiang, X.J., Chen, C., Han, T.L., Zhang, T., Cao, X.Q., Peng, B., Zhang, H., Xia, Y.Y., 2020. Using the Metabolome to Understand the Mechanisms Linking Chronic Arsenic Exposure to Microglia Activation, and Learning and Memory Impairment. *Neurotox. Res.* <https://doi.org/10.1007/s12640-020-00286-x>
- Zhang, Z., Liu, H., Zhou, H., Zhu, X., Zhao, Z., Chi, X., Shan, H., Gao, J., 2016. A facile route to core-shell nanoparticulate formation of arsenic trioxide for effective solid tumor treatment. *Nanoscale* 8, 4373–4380. <https://doi.org/10.1039/c5nr07860a>
- Zhao, P., Guo, Y., Zhang, W., Chai, H., Xing, H., Xing, M., 2017. Neurotoxicity induced by arsenic in Gallus Gallus: Regulation of oxidative stress and heat shock protein response. *Chemosphere* 166, 238–245. <https://doi.org/10.1016/j.chemosphere.2016.09.060>
- Zhao, Y., Zang, G., Yin, T., Ma, X., Zhou, L., Wu, L., Daniel, R., Wang, Y., Qiu, J., Wang, G., 2021. A novel mechanism of inhibiting in-stent restenosis with arsenic trioxide drug-eluting stent: Enhancing contractile phenotype of vascular smooth muscle cells via YAP pathway. *Bioact. Mater.* 6, 375–385. <https://doi.org/10.1016/j.bioactmat.2020.08.018>
- Zhou, Z., Song, J., Nie, L., Chen, X., 2016. Reactive oxygen species generating systems meeting challenges of photodynamic cancer therapy. *Chem. Soc. Rev.* 45, 6597–6626. <https://doi.org/10.1039/C6CS00271D>
- Zhu, J., Chen, Z., Lallemand-Breitenbach, V., De Thé, H., 2002. How acute promyelocytic leukaemia revived arsenic. *Nat. Rev. Cancer* 2, 705–713. <https://doi.org/10.1038/nrc887>