

PHYTOCHEMICAL ANALYSIS OF SALVINIA MOLESTA D.S. MITCHELL. BY GC-MS ANALYSIS

Ambili SN^{1*}, Medo Merina R²

ABSTRACT

The present investigation was undertaken to explore the potential bioactive components present in the ethanol, acetic acid and ethyl acetate extract of S. molesta by Gas Chromatography-Mass Spectrometry (GC-MS) analysis. The GC-MS analysis of ethanol extract showed 10 bioactive compounds. The major compounds identified were 3-Pyrrolidinol, 4-Imidazolidinone,5,5-diphenyl-2-thioxo-, 1,2-Dimethyl-3-nitro-4-nitrosobenzene, Harmaline, Tridecanoic acid, Phenytoin, Benzofuran-2-one, 2,3-dihydro-3, 3-dimethyl-4-nitro-, i-Propyl 9-tetradecenoate, 8-Isopropyl-5-methyl-5,6,7,8-tetrahydro-2, 4-quinazolinedione and Benzenemethanimine, .alpha.-phenyl-. Acetic acid extract shows the bioactive compounds were be 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4, 5, 6, 7-tetrahydro-, isopropyl ester, 1H-Indole, 5methyl-2-phenyl-, Indole-2-one, 2,3-dihydro-N-hydroxy-4-methoxy-3, 3-dimethyl-, Pyrido [2, 3-d] pyrimidine, 4-phenyl- and Ethyl 2-(2-chloroacetamido)-3,3,3-trifluoro-2-(2-fluoroanilino)propionate. The ethyl acetate shows the phytochemical compounds like Cholesta-6, 22, 24-triene, 4,4-dimethyl and 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4,5,6,7-tetrahydro-, isopropyl ester. The compounds identified were phenolic compound, alkaloid, saturated fatty acid, steroid and indole derivative.

Keywords: Analysis, compounds, extract, GC-MS, and retention time,

^{1*}Research Scholar, Department of Botany & Research Centre, Women's Christian College, Nagercoil. Email id: ambilisn0704@gmail.com

²Assistant Professor, Department of Botany & Research Centre, Women's Christian College, Nagercoil. Email id: medomerina2015@gmail.com, Affiliated to Manonmaniam Sundaranar University, Abishekapatti, Tirunelveli, Tamil Nadu, India.

*Corresponding author: Ambili SN

*Email id: ambilisn0704@gmail.com

DOI: 10.53555/ecb/2022.11.12.446

INTRODUCTION

Phytochemicals are natural bioactive compound found in plants which acts as a defense system against diseases to protect against diseases (James and Anderson, 1983).

GC-MS is normally used for direct analysis of components that in traditional medicines composed of two major building blocks; the gas chromatography and the mass spectrophotometer (Sridharan *et al.*, 2011). GC-MS is the best technique to identify the bioactive components of long chain hydrocarbons, alcohols and acids used in the analysis of the herbal medicines (Sridharan *et al.*, 2011). The two components GC and MS used together allows a much finer degree of substance identification than either unit used separately. The Mass spectrometry capture, ionize, accelerate, deflect, and detect the ionized molecules separately by detecting ionized fragments using their mass to charge ratio (Amirav *et al.*, 2008).

Globally Salvinia species are known as one of the most troublesome fresh water weeds because of their negative impacts on the environment. These ferns are spread all over the surface of the water levels by forming a mat-like mesh and affecting the ecosystem of fresh water resources like ponds, rivers, lakes, and rice paddies (Mujaju et al., 2021). Different species of Salvinia were reported for the presence of many pharmacological compounds and various activities such as antimicrobial, antioxidant and immunomodulatory activity (Purgato et al., 2021; Naheed et al., 2021; Nithya et al., 2019). So, the aim of the present study is to identify the active compounds present in S. molesta by GC-MS analysis and collected from Poovangaparambu, Kanniyakumari Dist.

MATERIALS AND METHODS

Collection, Identification and Preparation of extracts

The fresh samples of *S. molesta* were manually collected and were identified by the experts. It is initially washed with pond water, followed by running tap water and finally with sterile distilled water to remove the adhering microscopic and macroscopic debris. It was shade dried and powdered. The fine powdered samples were stored in an airtight container for further use.

The dried powdered samples (10gm) extracted using various solvent extracts (ethanol, acetic acid, and ethyl acetate). Then the extracts were stored in a refrigerator for further use (Rebecca *et al.*, 2012).

GC-MS (Gas Chromatography-Mass Spectrometry)

A high-resolution mass spectrum equipped with a combination with data system in Gas Chromatography was used for the chemical analysis of aquatic pteridophyte. GC-MS analysis of the extracts was carried out by the following method of Hema et al. (2010) using a GC-MS Clarus 500 Perkin Elmer system and gas chromatography interfaced to mass spectrometer (GC-MS). The detection of the compounds employed with the NIST (National Institute of Standards and Technology). The relative % amount was calculated by comparing its peak area to the total areas. Software adopted to handle mass spectra and chromatogram was Turbomass (Version 5.2).

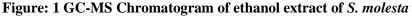
RESULTS AND DISCUSSION

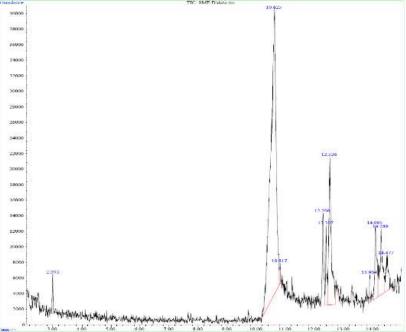
GC-MS analysis was carried out using of GC Clarus 500 Perkin Elmer System and Gas chromatography (GC-MS) instrument. The GC-MS identification of the chemical constituents was based on comparison of their mass spectra with NIST library. Structure was defined by percentage similarity values. They are confirmed by the study of base peaks, retention time (RT) and molecular weight (MW) of the compounds.

The active constituent of ethanol extract of S. molesta were analysed by GC-MS analysis, the detail tabulation is given in Table. 1. The ethanol extract showed the presence of ten phytochemical constituents. The highest peak was identified was 3-Pyrrolidinol with the retention time of 2.955. It showed the peak area of 1.38 %. The other peaks identified was 4-Imidazolidinone, 5,5-diphenyl-2thioxo- with the retention time of 10.625 and the peak area of 58.83 %. 1,2-Dimethyl-3-nitro-4nitroso-benzene had the retention time of 10.817 with a peak area of 0.69 %. Harmaline showed retention time of 12.290 with a peak area of 5.21%. Tridecanoic acid and Phenytoin with retention time of 12.397 have a peak area of 3.20 % and 12.526 with a peak area of 14.38 % respectively. The compound Benzofuran-2-one, 2,3-dihydro-3,3dimethyl-4-nitro- showed the sharp peak area of 0.86 % at a retention time (RT) of 13.904 minutes. i-Propyl 9-tetradecenoate is a compound with a peak area of 5.26 % with the retention time of (RT) 14.095 minutes. 8-Isopropyl-5-methyl-5,6,7,8tetrahydro-2, 4-quinazolinedione is a compound had a peak area of 6.80 % and showed its presence at a retention time of 14.289 minutes. The compound Benzenemethanimine, .alpha.-phenylhad a peak area of 3.39 % with the retention time of 14.477 minutes Table: 1; Fig: 1.

Table. I Compounds furthined as channel extract of 5. motesti								
Peak No.	Compound Name	Area of %	Retention Time (Min.)	Molecular Weight (G/mol.)	Molecular Formula			
1	3-Pyrrolidinol	1.38	2.995	87.12	C ₄ H ₉ NO			
2	4-Imidazolidinone, 5,5-dipenyl-	58.83	10.625	268.3	$C_{15}H_{12}N_2OS$			
	2-thioxo-							
3	1,2-Dimethyl-3-nitro-4-nitroso-	0.69	10.817	180	$C_8H_8N_2O_3$			
	benzene							
4	Harmaline	5.21	12.290	214.26	$C_{13}H_{14}N_2O$			
5	Tridecanoic acid	3.20	12.397	214.34	$C_{13}H_{26}O_2$			
6	Phenytoin	14.38	12.526	252.27	$C_{15}H_{12}N_2O_2$			
7	Benzofuran-2-one, 2,3-dihydro-	0.86	13.904	207.18	$C_{10}H_9NO_4$			
	3, 3-dimethyl-4-nitro-							
8	i-Propyl	5.26	14.095	268	$C_{17}H_{32}O_2$			
	9-tetradecenoate							
9	8-Isopropyl-5-methyl-5,6,7,8-	6.80	14.289	222.28	$C_{12}H_{18}N_2O_2$			
	tetrahydro-2, 4-							
	quinazolinedione							
10	Benzenemethanimine,.alpha	3.39	14.477	181.23	$C_{13}H_{11}N$			
	phenyl-							

Table: 1 Compounds identified as ethanol extract of S. molesta





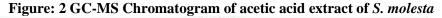
The compounds present in acetic acid extract of *S. molesta* is displayed in Table: 2; Fig: 2. Five bioactive compounds were detected. The bioactive compounds were found to be 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4, 5, 6, 7-tetrahydro-, isopropyl ester showed a highest peak area of 16.17 % and a retention time of (RT) 14.433 minutes. The compound 1H-Indole, 5-methyl-2-phenyl- had a peak area of 26.29 % with the retention time of (RT) 14.529 minutes.

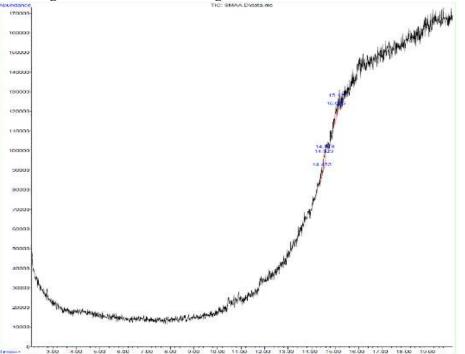
Indole-2-one, 2,3-dihydro-N-hydroxy-4-methoxy-3, 3-dimethyl- is a compound with a peak area of 15.35 % and a retention time (RT) of 14.578 minutes. Pyrido [2, 3-d] pyrimidine, 4-phenyl- had a peak area of 25.59 % with the retention time of 15.035. A chemical compound identified as Ethyl 2-(2-chloroacetamido)-3,3,3-trifluoro-2-(2-

fluoroanilino) propionate with the peak area of 16.60 % and the retention time was found to be 15.121 minutes.

Peak No.	Compound Name	Area of %	Retention Time (Min.)	Molecular Weight (G/mol.)	Molecular Formula
1	1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3- methyl-4-oxo-4,5,6,7- tetrahydro-, isopropyl ester	16.17	14.433	355.43	C ₂₁ H ₂₅ NO ₄
2	1H-Indole, 5-methyl-2- phenyl-	26.29	14.529	207.27	$C_{15}H_{13}N$
3	Indole-2-one, 2,3-dihydro-N- hydroxy-4-methoxy-3, 3- dimethyl-	15.35	14.578	207.23	$C_{11}H_{13}NO_3$
4	Pyrido [2, 3-d] pyrimidine, 4- phenyl-	25.59	15.035	207.23	$C_{13}H_9N_3$
5	Ethyl 2-(2-chloroacetamido)- 3,3,3-trifluoro-2-(2- fluoroanilino) propionate	16.60	15.121	356.70	$C_{13}H_{13}ClF_4N_2O_3$

 Table: 2 Compounds identified as acetic acid extract of S. molesta





The ethyl acetate extract of *S. molesta* showed the presence of two phytochemical constituents. The phytochemicals were found to be Cholesta-6, 22, 24-triene, 4, 4-dimethyl- and 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4, 5, 6, 7-tetrahydro-, isopropyl ester. The compound Cholesta-6, 22, 24-triene, 4, 4-dimethyl-showed the highest sharp peak area of 50.04 % at a retention time of 14.620 minutes and 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4, 5, 6, 7-tetrahydro-, isopropyl ester has a peak area of 49.96 % at a retention time (RT) of 16.348 minutes Table: 3; Fig: 3.

The compounds identified from three extracts were phenolic compound, alkaloid, saturated fatty acid, steroid and indole derivative.

Most of the identified compounds were phenols. Fatty acids were also seen. Natural phytochemical including phenolic compounds and fatty acids are major bioactive compounds known to be beneficial against many diseases and have been reported to possess a wide range of biological effects like anti-oxidant and antibacterial activities (Sofowora, 1996). Phenols have been reported to possess antioxidant (Heima *et al.*, 2002), antibacterial and antifungal activities (Alasalvar *et al.*, 2001; Acamovic and Brooker, 2005; Edreva *et al.*, 2008).

Alkaloids, steroids were also noticed in the extracts. Alkaloids are also essential for the treatment of cardiovascular and kidney disorders (Sweetman, 2005). It was also reported that alkaloids have a wide range of pharmacological activities including antimalarial, antibacterial (Cushnie *et al.*, 2014), anticancer (Kittakoop *et al.*, 2014) and antihyperglycemic activities (Qiu *et al.*, 2014).

Plant steroids possess many interesting medicinal, pharmaceutical and agrochemical activities like anti-tumor, immunosuppressive, hepatoprotective, antibacterial, plant growth hormone regulator, sex hormone, antihelminthic, cytotoxic and cardiotonic activity (Patel and Savjani, 2015).

The presence of Indole derivatives in the extracts also known to possess various biological activities, i.e., antiviral, anti-inflammatory, anticancer, antiHIV, antioxidant, antimicrobial, antitubercular, antidiabetic, antimalarial and anticholinesterase activities etc, which created interest among researchers to synthesize a variety of indole derivatives (Kumar and Ritika, 2020).

Ethanol extract revealed the presence of a greater number of compounds. In previous studies also ethanol extract revealed the presence of a greater number of compounds (Tyagi and Agarwal, 2017; Abdelhamid *et al.*, 2015; Sreenath *et al.*, 2016; Kumar *et al.*, 2018).

The ethyl acetate showed least number of compounds. this is because of geographical location, climatic conditions and edaphic factors amongst others are known to affect the chemical composition of plants and their medicinal properties (Gairola *et al.*, 2010).

Pea	Compound Name	Area	Retenti	Molecula	Molecula
k		of %	on Time	r Weight	r
No.			(Min.)	(G/mol.)	Formula
1	Cholesta-6, 22, 24-triene,	50.0	14.620	394.7	$C_{29}H_{46}$
	4,4-dimethyl-	4			
2	1H-Indole-2-carboxylic	49.9	16.348	355.43	$C_{21}H_{25}NO$
	acid, 6-(4-ethoxyphenyl)-	6			4
	3-methyl-4-oxo-4,5,6,7-				
	tetrahydro-, isopropyl				
	ester				

 Table: 3 Compounds identified as ethyl acetate extract of S. molesta

Figure: 3 GC-MS Chromatogram of ethyl acetate extract of S. molesta



CONCLUSION

GC-MS analysis have facilitated to identify bioactive compounds. In the present studies all the studied extracts showed the presence of phyto *Eur. Chem. Bull. 2022, 11(Regular Issue 12), 4710-4716*

chemical compounds and ethanol extract of the *S. molesta* has proved to contain more number of phytochemical compounds. Isolation of the individual compound and subjecting it to biological

activity will give fruitful results. Very few works done by GC-MS analysis in this plant so it is the first step towards understanding the nature of active principles in *S. molesta* extracts and this study paves way for further research.

ACKNOWLEDGEMENT

The authors are very thankful to the Manonmaniam Sundaranar University, Abishekapatti, Tirunelveli-627 012, Tamil Nadu, India.

REFERENCE

- Abdelhamid, MS, Kondratenko, EI & Lomteva, NA 2015, 'GC-MS analysis of phytocomponents in the ethanolic extract of *Nelumbo nucifera* seeds from Russia', Journal of Applied Pharmaceutical Science, vol. 5, no. 04, pp. 115-118.
- 2. Acamovic, T & Brooker, JD 2005, 'Biochemistry of plant secondary metabolites and their effects in animals', Proceedings of the Nutrition Society, vol. 64, pp. 403-412.
- Alasalvar, C, Grigor, JM, Zhang, DL, Quantick, PC & Shahidi, F 2001, 'Comparison of volatiles, phenolics, sugars, antioxidant vitamins and sensory quality of different colored carrot varieties', Journal of Agricultural and Food Chemistry, vol. 49, no. 3, pp. 1410-1416.
- Amirav, A, Gordin, A, Poliak, M, & Fialkov, AB 2008, 'Gas Chromatography Mass Spectrometry with supersonic molecular beams', Journal of Mass Spectrometry, vol. 43, no. 2, pp. 141-163.
- Cushnie, TPT, Cushnie, B & Lamb, AJ 2014, 'Alkaloids: an overview of their antibacterial, antibiotic-enhancing and antivirulence activities', International Journal of antimicrobial Agents, vol. 44, no. 5, pp. 377-386.
- Edreva, A, Velikova, V, Tsonev, T, Dagnon, S, Gurel, AL, Aktas, L & Gesheva, E 2008, 'Stress-protective role of secondary metabolites: Diversity of functions and mechanisms', General and Applied and Plant Physiology, vol. 34, no. 1-2, pp. 67-78.
- Gairola, S, Shariff, NM, Bhatt, A & Kala, CP 2010, 'Influence of climate change on production of secondary chemicals in high altitude medicinal plants: Issues needs immediate attention', Journal of Medicinal Plants Research, vol. 4, no. 18, pp. 1825-1829.
- Heima, KE, Tagliaferroa, AR & Bobilya, DJ 2002, 'Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships', Journal of Nutritional Biochemistry, vol. 13, no. 10, pp. 572-584.

- Hema, R, Kumaravel, S, Gomathi, S & Sivasubramaniam, C 2010, 'Gas chromatography- mass spectroscopic analysis of *Lawsonia inermis* leaves', New York Science Journal, vol. 3, no. 11, pp. 141-143.
- 10.James, W & Anderson, MD 1983, 'Plant fiber and blood pressure', Annals of International Medicine, vol. 98, no. 5, pp. 842-846.
- 11.Kittakoop, P, Mahidol, C & Ruchirawat, S 2014, 'Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis and smoking cessation', Current Topics in Medicinal Chemistry, vol. 14, no. 2, pp. 239-252.
- 12. Kumar, DG, Karthik, M & Rajakumar, R 2018, 'GC-MS analysis of bioactive compounds from ethanolic leaves extract of *Eichhornia crassipes* (Mart) Solms and their pharmacological activities', The Pharma Innovation Journal, vol. 7, no. 8, pp. 459-462.
- 13.Kumar, S & Ritika 2020, 'A brief review of the biological potential of indole derivatives', Future Journal of Pharmaceutical Sciences, vol. 6, no. 121
- 14.Mujaju, C, Mudada, N & Chikwenhere, GP 2021, 'Invasive alien species in Zimbabwe (Southern Africa)', In book: Invasive Alien Species, pp. 330-361.
- 15.Naheed, N, Maher, S, Saleem, F, Khan, A, Wadood, A, Rasheed, S, Choudhary, MI, Froyen, M, Abdullah, I, Mizra, MU & Trant, JF 2021, 'New isolate from *Salvinia molesta* with antioxidant and urease inhibitory activity', Drug Development Research, vol. 82, pp. 1169-1181.
- 16.Nithya, TG, Sumalatha, D, Ragunathan, MG & Jayanthi, J 2019, 'Immunomodulatory activity of *Salvinia molesta* DS Mitchell in fresh water crab Oziotelphusa senex senex bacterially challenged with *Pseudomonos aeruginosa*', Journal of King Saud University-Science, vol. 31, pp. 1471-1477.
- 17.Patel, SS & Savjani, JK 2015, 'Systematic review of plant steroids as potential antiinflammatory agents: Current status and future perspectives', The Journal of Phytopharmacology, vol. 4, no. 2, pp. 121-125.
- 18.Purgato, GA, Lima, S, Baeta, JVPB, Pizziolo, VR, De Souza, GN, Munoz, DG & Diaz, MAN 2021, 'Salvinia auriculata: chemical profile and biological activity against Staphylococcus aureus isolated from bovine mastitis', Brazilian Journal of Microbiology, vol. 52, pp. 2401-2411.
- 19. Qiu, S, Sun, H, Zhang, A, Xu, H, Yan, G, Han, Y & Wang, X 2014, 'Natural alkaloids: basic aspects, biological roles and future

perspectives', Chinese Journal of Natural Medicines, vol. 12, no. 6, pp. 401-406.

- 20. Rebecca, LJ, Dhanalakshmi, V & Shekhar, C 2012, 'Antibacterial activity of *Sargassum ilicifolium* and *Kappaphycus alvarezii*', Journal of Chemical and Pharmaceutical Research, vol. 4, no. 1, pp. 700-705.
- 21.Sofowara, A 1996, 'Research on medicinal plants and traditional medicine in Africa', Journal of Alternative and Complementary Medicine', vol. 2, pp. 365-72.
- 22.Sreenath, KB, Sundaram, S, Gopalakrishnan, VK & Poornima, K 2016, 'Quantitative phytochemical analysis, *in vitro* antioxidant potential and Gas Chromatography-Mass Spectrometry studies in ethanolic extract of *Azolla microphylla*', Asian Journal of Pharmaceutical and Clinical Research, vol. 9, no. 2, pp. 318-323.
- 23.Sridharan, O, Meena, S, Kavitha, V & Nayagam, AAJV 2011, 'GC-MS study and phytochemical profiling of *Mimosa pudica* Linn', Journal of Pharmacy Research, vol. 4, no. 3, pp. 741-742.
- 24. Sweetman, SC 2005, 'Martindale In the complete drug Reference Pharmaceutical Press, Williams clowes, Suffock, UK', pp. 907.
- 25. Tyagi, T & Agarwal, M 2017, 'Ethnomedicine analysis of bioactive constituents in ethanolic leaf extract of *Pistia stratiotes* L. and *Eichhornia crassipes* (Mart.) Solms by GC-MS', Advances in Bioresearch, vol. 8, no. 5, pp. 204-211.