

EVALUATION OF THE CHEMICAL AND MICROBIOLOGICAL CONTAMINATION OF THE RIVER SITNICA WATERS

(KOSOVO): A STATISTICAL APPROACH

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Chemical and microbiological contamination of ground water has several implications for human health. The study of chemical and microbiological contamination of water is carried out in Sitnica river (Kosova). Water samples are collected at 10 sampling points and fourteen different parameters are investigated. The program, Statistica 6.0, is used for calculations of basic statistical parameters and anomalies (extremes and outliers). The chemometric approach and the results yielded useful information about water quality and can lead to better water resource management. Dendograms and box-whisker plots are drawn to evaluate the chemical variation. The level of some microbiological and phsico-chemical parameters and eco-toxic ions from surface water are compared with the results from the river source where anthropogenic effects are negative. Our results show that chemical and microbiological pollutions are the results of anthropogenic factors coming from settlements, peripheral rivers and agricultural in the region surround.

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Introduction

The sources of microbial and chemical contamination are numerous and include the land disposal of sewage effluents, sludge and solid waste, septic tank effluent, urban runoff, and agricultural, mining and industrial practices.^{1,2} Chemical contamination of drinking water is often considered a lower priority than microbial contamination by regulators, because adverse health effects from chemical contaminations are generally associated with long-term exposures, whereas the effects from microbial contamination are usually immediate.3 The quality of drinking water is an issue of primary interest for the residents of the European Union. 4 In peat bogs, water flows freely in the active layer of water or acrotelm. Water storage is critical to the balance of water in peat swamps and at surrounding areas. Logging activity, agriculture, peat extraction and destruction of peat swamp drainage activity also give a negative effect and has a bad implication on the hydrology.⁵

Decomposition of organic matter and pollution due to anthropogenic activity are the main sources of pollution of water.6 Therefore, multidisciplinary collaborative research is essential for understanding the pollution processes. As reported by Brils,7 adequate water quality in Europe is one of the most eminent concerns for the future. Good management of natural and environmental waters will give results if leading institutions constantly monitor information about environmental situation. Therefore, seeing it as a challenge for environmental chemists, our goal is to determine the amount and nature of pollutants in the environment.

This work is a continuation of our earlier studies of surface waters in Kosovo.8-11 One could claim that the most polluted areas in the world are those with the dense population. It should therefore be the foremost goal of environmentalists to prevent such pollution, and to educate the population towards proper management of ecosystems.12

The aim of the current work is to perform, a systematic research on the water of Sitnica river. The research is about chemical and microbiological aspects as they affect directly and indirectly the water quality, also impact the aquatic life. A major impact will be on biota where a large amount of this river flow is used for irrigation of agricultural lands and thus it may have an indirect impact on people's lives as they consume agricultural products that can deposit pollutants in the body of people through the food chain.¹³

Experimental

Study area

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The objective of this study is microbial and chemical examination of water quality of Sitnica (length = 90 km). Sampling strategy was planned at 10 monitoring stations spread over the source in the mountain and downstream to the end of the river within Kosovo's territory. Although there are more than 50 water quality parameters available, only 14 parameters are selected for our investigation. These parameters are: water temperature, conductivity, pH, turbidity, consumption of KMnO₄, total oxygen, nitrate nitrogen, nitrite nitrogen, ammonia nitrogen, total coliform, coliform bacteria of faecal origin, aerobic bacteria, sulfide-reducing anaerobic sporogens bacteria and streptococcus of faecal origin. The sampling sites are geographically positioned using geographic information system (GIS). The results are interpreted using modern statistical methods that can be used to locate pollution sources. Surface water sampling of champions and their elaboration at the depth ≥ 20 cm are done with Pyrex non-contaminating bottles according to standards methods for surface water. Some of the natural water samples are filtered with Whatman paper (0.45 μ m) made from cellulose nitrate in the bottle of Teflon under pressure of nitrogen (purity 99.99 %).

Sampling and sample preparation

For chemical analysis water samples are collected on 3 March 2010 in plastic bottles, previously rinsed three times with sampled water, and labeled with the date and the name of the sample. These samples are transferred to refrigerator (at 4 °C) for analysis in the laboratories.

For bacteriological evaluation, samples, taken on 7 March 2010, in plastic sterile bottles are refrigerated (at 4°C) and transported immediately to laboratory and analyzed on the same day.

All tests are performed at least thrice to calculate the average value. The sampling locations are chosen at points where pollution is expected due to closeness of factories, traffic, settlements or combinations of these factors. Sampling, preservation and experimental procedure for the water samples are carried out according to the standard methods for examination of water. ¹⁵⁻¹⁸ Samples are preserved in refrigerator after treatment. The study area and the sampling locations are shown in Figure 1. The details about sampling sites are presented in Table 1



Figure 1. Study area with sampling stations.

Chemical and microbiological characterization

Double distilled water was used in all experiments. All instruments are calibrated according to manufacturer's recommendations. Temperature of water was measured immediately after sampling, using digital thermometer, model Quick 63142. pH measurements were performed

using pH/ion-meter of Hanna Instruments. Electric conductivity is measured by InoLab WTW conductometer, turbidity with turbidimetric method with formazine standard, chemical expense with KMnO₄ using Thiemann Küebel volumetric method (boiling in acidic environment). Some of physico-chemical parameters (total oxygen, NO₂-, NO₃- and NH₄+) are determined using UV-VIS spectrometry method.

Table 1. Detailed description of sampling stations.

Sample	Locality	Coordinates	Possible pollution sources			
S_1	Jezerc	34T 0501340	Settlement			
		UTH4689442				
S_2	Rubofc	34T 0511168	Waste water, agriculture			
		UTH0701529				
S_3	Lypjan	34T 0509360	Road, waste water from			
		UTH4706698	Lypjan			
S_4	Lismir	34T 0505443				
		UTH4721072	flotation waste, waste water			
			from Prishtina			
S_5	Palaj	34T 0504992	Agriculture, waste water			
		UTH4725302	from Fushë kosova			
S_6	Plemetin	34T 0503152	Thermal power plant, waste			
		UTH4728263	water from Kastriot			
S 7	Pestova	34T 0499284	Agriculture			
		UTH4736665				
S_8	Vushtrri	34T 0496273	Agriculture, tin factory,			
	(exit)	UTH4741288	waste water from Vushtrria			
S 9	Mitrovica	34T 0490408	Phosphate and accu factory,			
	(exit)	UTH4749416	settlement, road, waste			
			water from Mitrovica			
S ₁₀	Zveqan	34T 0487012	Road, waste water from			
		UTH4754445	Mitrovica			

The total coliform bacteria are evaluated at 30 °C (agar plate) according to probable number technique (MPN) per 100 mL. The coliform bacteria of fecal origin were evaluated at 44 °C (endo agar plate). The total aeorobic bacteria in 1 mL are evaluated at 37 °C (plate count agar) according to "colony star" (Frank Gerber), 1MPN technique. Sulphide reducing anaerobic sporogens bacteria are evaluated at 37 °C (sulphite agar plate). The streptococcus bacteria of fecal origin are evaluated in 100 mL of water.

Statistical Methods

Program Statistica 6.0^{19} is used for all statistical calculations such as determination of basic statistical parameters (mean, geometric mean, median, maximum minimum variance and standard deviations) and determination of anomalies (extremes and outliers) for solution data. Outlier values are between 1.5 and 3, and extreme values are above 3 standard deviations.

Results

The physico-chemical parameters i.e. water temperature, EC, pH, turbidity, consumption of KMnO₄, total oxygeny and concentrations of NO₂⁻, NO₃⁻ and NH₄⁺ are presented in Table 2.

Table 2. Some physico-chemical parameters of the river water

Sample	Water temp. /° C	EC /μS cm ⁻¹	pH /1	Turbidi-ty, / NTU	Cons. of MnO ₄ -/mg L ⁻¹	TO /mg L ⁻¹	NO ₂ /mg L ⁻¹	NO ₃ /mg L ⁻¹	NH ₄ ⁺ /mg L ⁻¹
S_1	11.90	405	7.18	0.13	11.06	9.4	0.005	0.05	0.06
S_2	11.90	297	7.34	94	37.83	8.3	0.06	0.50	0.09
S ₃	11.80	309	7.39	79	28.44	7.8	0.05	0.50	0.08
S 4	11.90	594	7.09	51	37.6	7.1	0.11	0.50	0.17
S ₅	11.80	423	7.93	91	26.86	7.7	0.05	0.50	0.38
S_6	11.90	446	7.84	92	23.70	7.8	0.04	1.20	0.50
S_7	11.90	435	7.51	61	24.65	7.8	0.04	1.40	0.27
S_8	11.90	455	7.73	69	24.33	8.0	0.04	0.50	0.22
S 9	12.00	462	7.68	59	30.02	7.3	0.04	0.50	0.28
S ₁₀	11.90	445	7.68	59	22.12	8.1	0.04	0.50	0.23

Table 3. Microbiological parameters of the river water

Sample	Coliform bacteria /100 mL	Coliform bacteria (faecal origin) /100 mL	Aerobic bacteria at 37 ° C / 1 mL	Anaerobic sporogens sulphide reducing bacteria /100 mL	Streptococcus (faecal origin) /100 mL
WHO/MPN	100	0	300	10	0
S_1	0	0	0	0	0
S_2	>300	>300	>300	5	>300
S_3	300	300	>300	20	>300
S ₄	>300	300	>300	10	300
S_5	>300	>300	>300	>20	>300
S ₆	>300	>300	>300	>25	>300
S 7	>300	>300	>300	>100	>300
S ₈	>300	>300	>300	>100	>300
S 9	>300	>300	>300	>50.0	>300
S ₁₀	>300	>300	>300	10	>300

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The bacteriological parameters viz. total coliform bacteria, coliform bacteria of fecal origin, number of aerobic bacteria, numbers of anaerobic sulphide reducing sporogens and streptococcus of fecal origin are presented in Table 3. The basic statistics of the measured data set and anomalous values (outliers and extremes) from 9 variables of water are summarized in Table 4. Using experimental data (Table 2) and box plot approach of Tukey²⁰, anomalous values (extremes and outliers) in waters were determined for the whole region (Figure 2).

Discussion

The temperature variation at different locations of the river, indicated by the in situ readings, is given Table 2. The pH of the investigated samples, ranging from 7.09 to 7.93, indicates that all water samples are slightly alkaline. Electrical conductivity, ranging from 297-594 μScm^{-1} , is within highest desirable limit (HDL). If we compare EC separately, the lowest value is observed at S_2 station and the highest value is observed at S_4 station. Higher EC values indicate the presence of higher content of dissolved salts in the water. The lowest value of turbidity is observed at point S_1 (near the river source) while at all points we observed higher values of turbidity coming

from settlements, gravel separations and anthropogenic waste. Consumption of KMnO₄ has maximum values at S_2 and S_4 while the lowest vale is at S_1 . The total oxygen (TO) is in the range of 7.1 mg L⁻¹ at S_4 and 9.4 mg L⁻¹ at S_1 (source of the river).

Presence of nutrient salt of nitrogen in water as ammonia, nitrites and nitrates causes the process of eutrophication. The concentration of ammonia presents the potentional risk for the aquatic organisms. Total ammonia values changed downstream of the river and the lowest value is found at point S_1 and highest value at points S_5 and S_6 . The highest value of NO_2^- is found at S_4 and that for NO_3^- it is found at points S_6 and S_7 . Concentration of NO_2^- , NO_3^- and NH_4^+ are within highest desirable limit (HDL) according to WHO standards for drinking water.

Statistical methods are applied to find anomalous values also. Outlier value for NH_4^+ (0.5 μg L^{-1}) is found in the S_5 sample, extreme for NO_2^- (0.11 μg L^{-1}) is in the S_2 sample and extreme for NO_3^- (1.2-1.4 μg L^{-1}) is found in S_6 and S_7 samples.

The National Primary Drinking Water Regulations (NPDWRs) have prescribed limits of the amount of total coliforms, faecal coliform, *E. coli, Salmonella spp.*

Table 4. Basic statistical parameters for 9 variables in 10 water samples.

Variable	Descriptive Statistics						
	Mean	Geo. mean	Median	Minimum	Maximum	Variance	SD
Water temp. /°C	11.9	11.9	11.9	11.8	12.0	0.003	0.05676
EC /μS cm ⁻¹	427.1	419.7	440.0	297.0	594.0	6885.656	82.97985
pH /1	7.54	7.53	7.56	7.09	7.93	0.079	0.28143
Turbidity/NTU	65.51	37.85	65.0	0.13	94.00	771.943	27.78386
Cons. of MnO ₄ -/mg L-1	26.66	25.44	25.76	11.06	37.83	60.128	7.75423
TO /mg L ⁻¹	7.93	7.91	7.80	7.10	9.40	0.391	0.62548
NO ₂ -/mg L-1	0.0475	0.0391	0.0400	0.0050	0.1100	0.001	0.02617
NO ₃ -/mg L-1	0.6150	0.4805	0.5000	0.0500	1.4000	0.152	0.39019
NH ₄ ⁺ /mg L ⁻¹	0.2280	0.1874	0.2250	0.0600	0.5000	0.019	0.13911

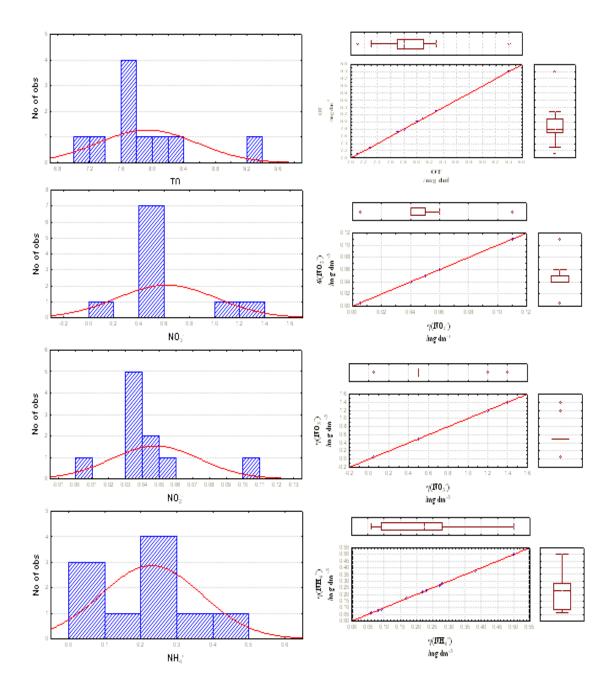


Figure 2. Frequency histograms and scatter box plot diagrams of 4 measured variables.

Shigella spp. and Vibrio spp. as primary standards to protect the health of the public by limiting the levels of contaminants in drinking water.²² In the present study (Table 3) the number of total coliform bacteria in 100 mL river water is found to be greater than 300 in samples S₂-S₁₀ and are well above recommended number by WHO standards (MPN = 100). The number of total coliform bacteria of faecal origin (in100 mL river water) is greater than 300 in samples S₂-S₁₀ and are above recommended number by WHO standards (MPN = 0). Osuinde and Enuezie reported that high coliform count obtained in water samples may be attributed to faecal contamination of the water sources.²³ Hence none of the water samples analyzed complies with EPA standard for coliform in water. According to EPA standard, every water sample that has coliform must be analyzed for either faecal coliforms or E. coli to ascertain whether the source of contamination is from human or animal waste, and possibly pathogenic bacteria or organism, such as Gardia and Cryptosporidium, may be present. The number of aerobic bacteria in 1 mL at 37 °C are found to be greater than 300 in samples S_2 - S_{10} (MPN = 300). The number of sulfide reducing anaerobic sporogens bacteria in 100 mL is above the recommended number by WHO standards (MPN = 10) in S_3 and in S_5 - S_9 . The number of streptococci bacteria of faecal origin in 100 mL is greater than 300 in samples S₂-S₁₀ and therefore much above the recommended number by WHO standards (MPN = 0).

Conclusions

The chemical and microbiological analysis shows that Sitnica river water is highly polluted. Therefore this water cannot be used without previous physical and chemical treatment. A long and uncontrolled discharge of municipal waste impacts the water quality. The large number of total coliforms and coliforms bacteria in faecal origin, sulfide reducing bacteria and streptococcus bacteria, and high concentration of nutrient salt of nitrogen as ammonia, nitrites and nitrates showed the faecal contamination of the river Sitnica water in some samples is beginning to reach the aquifer. For this reason, we recommend avoiding the discharge of waste water without treatment, mainly from septic tanks, which are extensively used in the area.

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