



Assessment of antioxidant potential of β -carotene and some edible fruits

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Abstract

Gac fruit (*Momordica cochinchinensis*), watermelon (*Citrullus lanatus*), and tomato fruit (*Solanum lycopersicum*) are high in natural antioxidants and have anticancer properties. Gac is a rich source of lycopene. These are commonly used in cancer, hypertension, dyslipidemia, inflammation, wound healing, anti-wrinkle products, moisturizers, and ulceration. In the present study, the freeze-dried powder of juices from the fruits of *Momordica cochinchinensis* (MC), *Citrullus lanatus* (CL), and *Solanum lycopersicum* (SL), as well as β -carotene (BC), are evaluated, using ascorbic acid (AA) as a reference standard. Three herbal products wise Sample-A MC, Sample-B CL, Sample-C SL, and β -carotene as sample-D were evaluated for invitro antioxidant activity. Free radical scavenging properties assessed by five invitro methods are DPPH, hydrogen peroxide (H₂O₂), superoxide, ferrous reducing antioxidant capacity assay (FRAC), and nitric oxide radical-scavenging activity. All the products were found to have potent antioxidant activity, which was significant ($P < 0.05$) compared to ascorbic acid. All the assays were performed in triplicate, and the results are represented as mean SD. P values are calculated by ANOVA at a 95% confidence interval by GraphPad Prism 8.0.2.

Keywords: Antioxidant; fruits; *in vitro*; edible; DPPH

1. Introduction

The fruit of MC has the potential to prevent breast and melanoma cell cancer¹. MC is a plant of the Cucurbitaceae², the extract and various phytochemicals from its fruit has anti-gastritis, wound healing³, anti-inflammatory, anti-proliferative⁴, Anti-hypertensive⁵, Anti-hyperlipidemia⁶, anthelmintic⁷, anti-oxidant⁸ even MC extract exhibited greater antioxidant activity compared to Vit-A and C. A skin cream containing MC extract demonstrated a significant moisturizing smoothness and antiwrinkle effect in clinical trials⁹. Hypoglycemic property, improve fertility by preventing testicular and epididymal damages¹⁰, and nutritional value^{11,12}.

MC fruit are rich in carotenoid content; the lycopene content is 380 μ g/gm of seed membrane, which is 10-fold that of a well-known source of lycopene¹³, relative to mass MC fruit contains 70 times the lycopene found in SL and up to ten times the β -carotene found in carrot². MC fruit is an excellent source of dietary supplements rich in multi-

phytochemicals¹⁴. Vitamin C, vitamin E, omega-3 and omega-6 fatty acids¹⁵, Carotenoids¹⁶ such as lutein, zeaxanthin, and cryptoxanthin, β -carotene, cis- and trans- β -carotene, cis- and trans-lycopene, fiber, Protein, and calcium². are all important phytonutrients found in MC fruit. Carotenoids from MC are more bioaccessible than those from carrots and SL¹⁷. Another fruit of CL, this is a member of the Cucurbitaceae family¹⁸. Lycopene, beta-carotene, vitamin C, citrulline, polyphenols like phenolic acids, flavonoids, stilbenes,¹⁹ and lignans, carbohydrate (sucrose, glucose, and fructose)²⁰, and ascorbic acid²¹ are all found in CL.

CL juice exhibits anti-inflammatory^{22,23} anti-hyperlipidemic²³, antioxidant²⁴ anti-hyperglycemic activities^{24,25}. Extracts of pulp have a gastroprotective, antiulcerative effect²⁶, reduces atherosclerosis²⁷ and have analgesic properties²⁸. They also exhibit anti-urolithiatic and diuretic activity²⁹. Citrulline (content of CL juice) supplements relieve fatigue or muscle soreness and lower the rating of perceived exertion (RPE)³⁰. CL flesh extract increases sexual potency or sustains erections³¹. Supplementation of CL juice prevents increased post-exercise BP in females³². Daily CL consumption is useful for atherosclerosis (decreased vascular cell adhesion molecule-1)³³. ethanolic extract of CL rind is cytotoxic and has been evaluated taking 7 human cancer cell lines: A549, Caco-2, H1299, HCT116, Hep2, HepG2, and MCF-7³⁴. CL juice has been shown to have hepatoneuroprotective³⁵, and improve muscle endurance-support³⁶.

The other content of my product is SL, which belongs to the family Solanaceae³⁷. Lycopene, β -Carotene, Phenol, Flavonoids, Vitamin C³⁸, Lutein, Phytoene, Phytofluene³⁹, γ -Carotene and Vitamin A⁴⁰ are the main components of SL. The fruit of SL has proven therapeutic effectiveness in patients with cardiovascular dysfunction, obesity, and diabetes⁴¹. SL juice has anticarcinogenic⁴², antioxidant and antiplatelet aggregation properties^{43,44}. Reduce the risk of cardiovascular diseases by reducing the concentration of inflammatory molecules (adhesion molecules) related to atherosclerosis⁴⁵. The SL fruit has nutritional and medicinal value as an antioxidant; it helps reduce cholesterol, prevents prostate cancer, reduces heart disease risk, improves liver health by detoxifying, and improves vision⁴⁶.

Reduced gastric acidity is clinically achieved by the SL dietary supplement in gastroesophageal reflux disease⁴⁷. SL juice is effective in reducing systolic and diastolic blood pressure^{48,49}, it decreases LDL cholesterol⁵⁰ and benefit male infertility by improving sperm motility⁵¹. consumption of SL juice modifies clinical asthma outcomes⁵² and reduce blood pressure in hypertensive pregnant women⁵³.

2. Materials and methods

Chemicals

2,2-diphenyl-1-picrylhydrazyl (DPPH), ethanol, 0.5 mM Nitroblue Tetrazolium (NBT), 100 M phenazine methosulphate, Tris-HCl buffer (pH 8.0), 50 mM phosphate buffer (pH 7.4), 2 mM H₂O₂, and nicotinamide adenine dinucleotide (NADH), phosphate buffer (0.2 M, pH 6.6) potassium ferricyanide, trichloroacetic acid, ferric chloride, sodium nitroprusside, Griess reagent, and Ascorbic acid.

Plant Material

All three-plant materials were procured from *IdoBio (Xi'an) Phytochem Co., Ltd., Xi'an, Shaanxi, China*, and carotenoid from *Research-Lab Fine Chem Industries, Mumbai, India*.

Statistics: The statistical significance were determined by one way ANOVA using GraphPad Prism 8 at 95% confidence interval. All the tests were performed in triplicate and graph were plotted using the mean value.

A) DPPH radical scavenging activity

The assay was carried out in accordance with the method of Blios M.S. 1958⁵⁴ with minor modification. DPPH antioxidant activity is performed by taking increasing concentrations (10, 20, 30, 40, 50, 60, 70, 80, 90, 100 $\mu\text{g/ml}$) of plant products and β -carotene. 1 ml of a 0.1 mM solution of DPPH in ethanol was taken from a freshly prepared stock solution and added to this 1 ml of sample solution in distilled water. Ascorbic acid is used as a reference standard. 30 minute later absorbance were measured at 517 nm. Lower absorbance indicates higher scavenging activity. The radical scavenging activity of the sample was expressed as a percentage inhibition activity, which was calculated as follows:

Where A_0 represents the absorbance of a blank (control) containing no sample, and A_t represents the absorbance of the sample. Every test was performed in triplicate, and a graph

$$\% \text{ inhibition} = \left(\frac{A_0 - A_t}{A_0} \right) \times 100$$

was plotted using the mean values.

B) Superoxide radical scavenging activity

Radicals of the superoxide anion (SO) are produced in a mixture of 2.0 ml of Tris-HCl buffer (16 mM, pH 8.0) with 2.0 ml of nitroblue tetrazolium (NBT, 0.3 mM) and 2.0 ml of nicotinamide adenine dinucleotide solution (NADH, 0.936 mM). Then, 1 ml of the sample solution (10, 20, 30, 40, 50, 60, 70, 80, 90, 100 $\mu\text{g/ml}$) was added to this combination. 2.0 ml of phenazine methosulfate solution (PMS, 0.12 mM) was then added to the mixture to start the reaction, which was then incubated for 5 minutes at 250 °C. Absorbance was measured at 560 nm using a blank consisting of 2.0 ml Tris-HCl buffer, 2.0 ml NBT, 2.0 ml NADH solution, 4.0 ml water, and 2.0 ml PMS solution. Ascorbic acid was treated as a reference in the same way that sample solutions were. The result was expressed as a percentage inhibition, which was calculated as follows.⁵⁵

Where A_0 is absorbance of blank (control) not containing sample, A_t is absorbance of sample.

C) Hydrogen peroxide scavenging activity

$$\% \text{ inhibition} = \left(\frac{A_0 - A_t}{A_0} \right) \times 100$$

The free radical scavenging ability of the sample and β -carotene was determined according to the method explained by Ruch, R.J., with some modifications. Sample solutions (10-100 $\mu\text{g/ml}$) were made with 50 mM phosphate buffer (pH 7.4), to which 0.6 ml of 2 mM H_2O_2 solution was added, and the final volume was made up to 2 ml by adding 50 mM phosphate buffer. incubated for 40 minutes, and absorbance was measured at 230 nm. Ascorbic acid is

used as a reference standard.⁵⁶

The scavenging activity of the sample was calculated as a percentage inhibition using the following equation:

$$\% \text{ inhibition} = \left(\frac{A_0 - A_t}{A_0} \right) \times 100$$

Where A_0 represents the absorbance of a blank (control) containing no sample and A_t represents the absorbance of the sample. Every test was performed in triplicate, and a graph was plotted using the mean values.

D) Ferrous reducing antioxidant power -FRAP

2 ml of samples of various concentrations (10, 20, 30, 40, 50, 60, 70, 80, 90, and 100 $\mu\text{g/ml}$) were mixed with 2 ml of phosphate buffer (0.2 M, pH 6.6) and 2 ml of potassium ferricyanide (10 mg/ml). This mixture is incubated at 50° C for 20 minutes. cooled to room temperature, and to this 2 ml of trichloroacetic acid (100 mg/l) was added. After centrifuging at 3000 rpm for 10 minutes, 1 ml of the supernatant was mixed with 1 ml of distilled water and 0.2 ml of 0.1% (w/v) ferric chloride and incubated for 10 minutes. The absorbance of the sample was measured at 700 nm by a spectrophotometer against a blank. A positive control is ascorbic acid. Higher reducing power results in greater absorbance.⁵⁷

E) Nitric oxide radical-scavenging activity

0.5 ml of sample with various concentrations from 10 $\mu\text{g/ml}$ to 100 $\mu\text{g/ml}$ were added to 2 ml of 10 mM sodium nitroprusside in phosphate buffered saline. and incubated at room temperature for 150 minutes. A volume of 0.5 ml of the Griess reagent (1% sulphanilamide, 2% orthophosphoric acid, and 0.1% naphthylethylenediamine dihydrochloride) was added, and the mixture was incubated at 25° C for 5 min. scavengers inhibit the generation of nitric oxide radicals from sodium nitroprusside in phosphate buffered saline. The diazotization of nitrite with sulphanilamide, followed by coupling with naphthylethylenediamine, results in a light pink to deep purple coloured chromophore in the reaction mixture, which is measured at 546 nm. The positive and negative controls were prepared in the same manner as the sample, with the exception that the solvent was used as the negative control and ascorbic acid was used as the positive control.^{58,59} The decreased absorbance as a percentage of inhibition was calculated using the following formula:

Where A_0 represents the absorbance of a blank (control) containing no sample and A_t represents the absorbance of the sample. Every test was performed in triplicate, and a graph was plotted.

$$\% \text{ inhibition} = \left(\frac{A_0 - A_t}{A_0} \right) \times 100$$

3. Results and Discussion

Results are represented as the mean of tests performed in triplicate. All the results were expressed as mean SD, calculated by one-way ANOVA, and were found statistically significant at $P < 0.05$. When compared to ascorbic acid, the GF, WM, TF, and BC have potent antioxidant activities. Overall, the results show that GF has the highest antioxidant potential for all types of free radicals tested in this study; GF's ability to scavenge free

radicals was greater than that of TF and WM. The free radical scavenging activity increased as the concentration of the test sample increased. The correlation of responses with the concentration ($\mu\text{g/ml}$) was significant at $P < 0.0001$.

For all approaches, the GF IC_{50} was consistently found to be between 48.72 and 55.95. The R square and IC_{50} values are shown in the following table.

Table 1. IC_{50} values of MC, CL, SL and BC by DPPH, Superoxide, H_2O_2 , and Nitric Oxide Activity

Sample	IC_{50}			
	DPPH	Superoxide	H_2O_2	Nitric Oxide
MC	51.11	48.72	55.95	54.30
CL	80.91	80.83	67.52	84.18
SL	59.02	63.08	53.08	59.86
BC	41.15	51.37	39.60	50.81
AA	32.72	34.90	30.15	41.95

DPPH radical scavenging activity

The ability of sample to remove stable DPPH ion was calculated as percentage inhibition. All sample has acceptable ion preventing ability out of this the lowest scavenging ability was found for WM and the highest for GF. IC_{50} and percentage inhibition of DPPH ion represented in table. Ascorbic acid used as standard. Out of all the test samples highest antioxidant activity was observed by GF at the dose of 100 $\mu\text{g/ml}$.

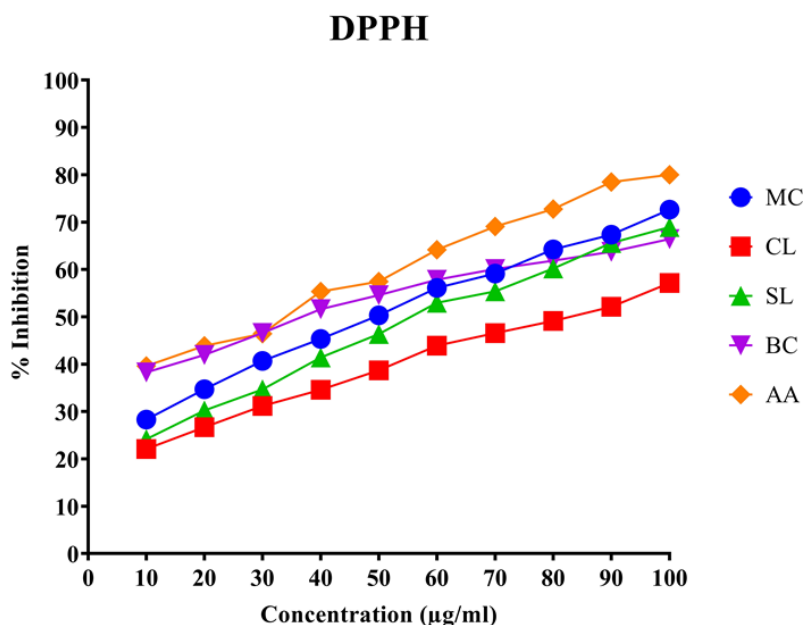


Fig. 1. DPPH radical scavenging activity of the samples (MC, CL, SL and BC), Data are expressed as mean \pm SD ($n = 3$, $p < 0.0001$) for all samples.

Superoxide radical scavenging activity

The discoloration of the mixture brought on by the removal of superoxide causes the absorbance to drop as sample concentration rises. 10 Sample concentration at 110-100 $\mu\text{g/ml}$

assayed in triplicate and SD and IC_{50} calculated. In this test the lowest inhibition activity from 10 to 100 $\mu\text{g/ml}$ seen for WM was from 21.94 to 57.16%. For beta carotene the lowest inhibition was 33.28% and highest 63.62%.

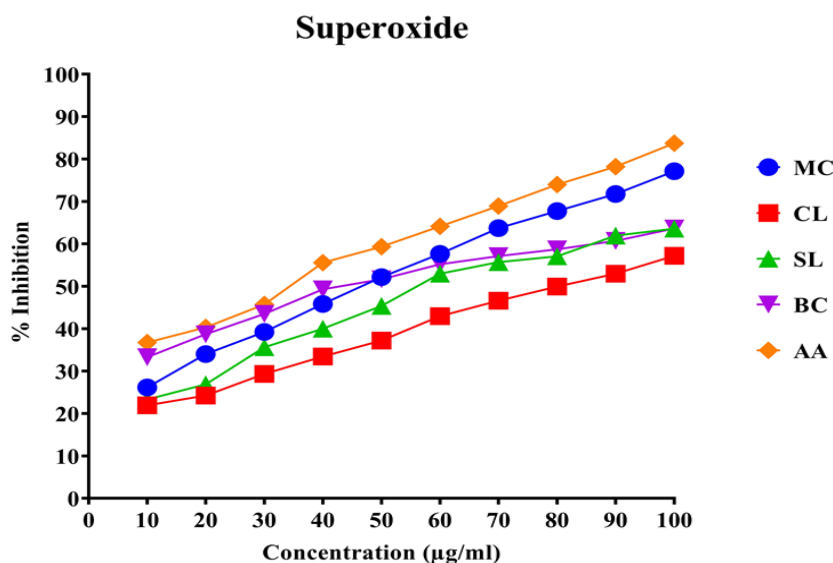


Fig. 2. Superoxide radical scavenging activity of the samples (MC, CL, SL and BC), Data are expressed as mean \pm SD ($n = 3$, $p < 0.0001$) for all samples.

Hydrogen peroxide scavenging activity

for all the test sample and the compound of samples showed increase inhibition activity as the concentration increased from 10 to 100 $\mu\text{g/ml}$. the highest activity was found for GF, 71.87% Inhibition of Hydrogen peroxide. For WM it was lowest comparing to other test samples. From all the test the H_2O_2 radical scavenging activity was highest 76.68% for BC.

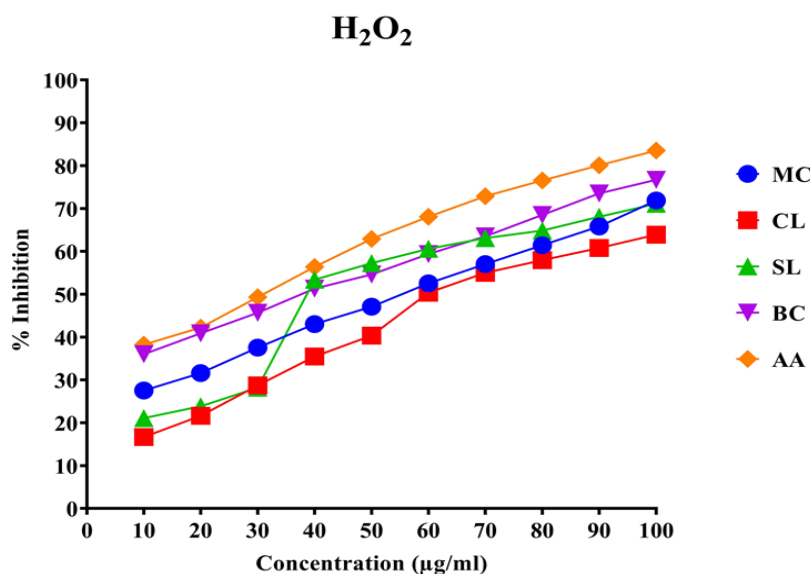


Fig. 3. Hydrogen peroxide scavenging activity of the samples MC, CL, SL and BC, Data are expressed as mean \pm SD ($n = 3$, $p < 0.0001$) for all samples.

Ferrous reducing antioxidant power – FRAP

Higher absorbance indicates greater radical scavenging activity, in this test the absorbance was gradually increase by increasing concentration with a significance of $P < 0.0001$ in relation to concentration versus absorbance. The highest absorbance was 0.099 for BC after TF (0.095). for GF and WM absorbance was 0.081 at 100 $\mu\text{g/ml}$.

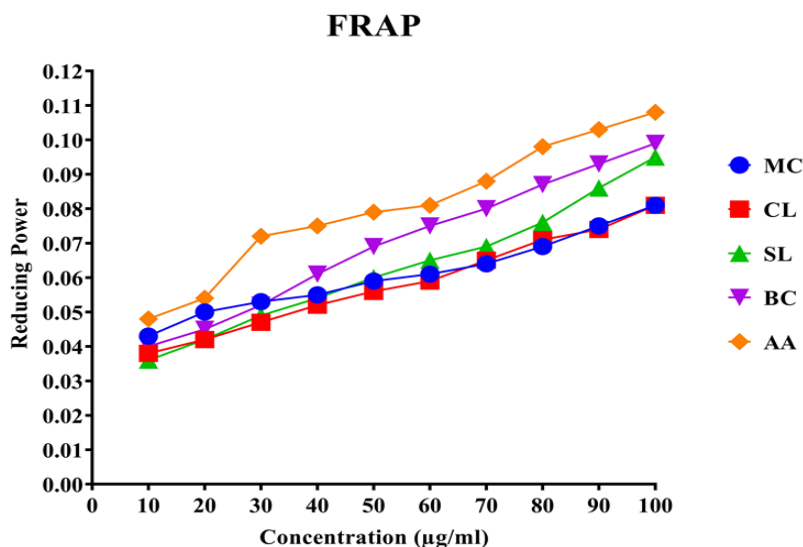


Fig. 4. Ferrous reducing antioxidant power of the samples MC, CL, SL and BC, Data are expressed as mean \pm SD ($n = 3$, $p < 0.0001$) for all samples.

Nitric oxide radical-scavenging activity

Nitric oxide scavenging potential was again lowest for the WM (55.08) than GF (68.55), TF (62.66) and BC (67.55) at the concentration of 100 $\mu\text{g/ml}$. The highest potential reported by GF ranging from 29.20 to 68.55. the scavenging property of BC was close to ascorbic acid.

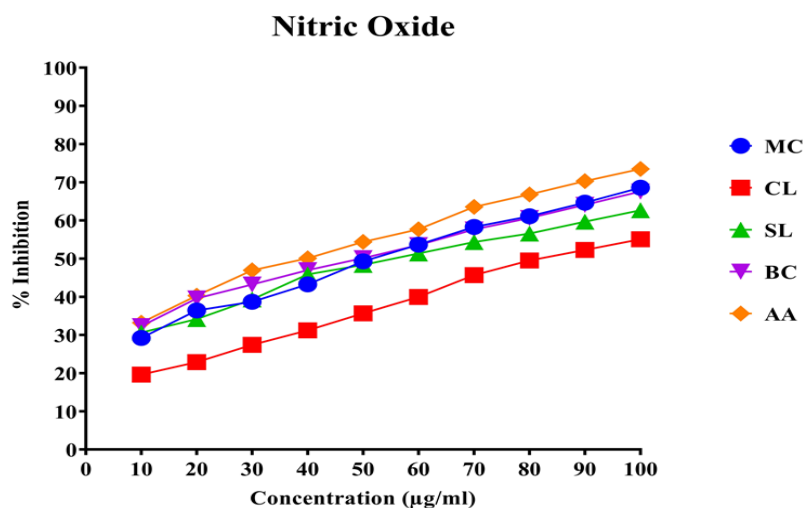


Fig. 5. Nitric oxide radical-scavenging activity of the samples MC, CL, SL and BC, Data are expressed as mean \pm SD ($n = 3$, $p < 0.0001$) for all samples.

4. Conclusion

All test results indicated that DPPH, H₂O₂, superoxide, FRAP, and nitric oxide had potential free radical inhibiting properties. The parts of the plant selected are rich in carotenoid pigments, which play an important role as antioxidants in biological fluids. The polyherbal formulation can be efficiently used as a potent antioxidant. All the results are statistically significant. The selection of plant parts was based on an extensive literature study to make the formulation for oral submucous fibrosis. The investigation's earlier findings, which included the identification of phytoconstituents, along with the therapeutic results, show that it has great potential for a wide range of clinical applications. It can be used successfully to treat oral submucous fibrosis, with added clinical and nutritional benefits.

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References

1. Dilani Wimalasiri, Chaitali Dekiwadia, Siat Yee Fong, Terrence J. Piva and Tien Huynh. 2020. "Anticancer activity of *Momordica cochinchinensis* (red gac) aril and the impact of varietal diversity." *BMC Complementary Medicine and Therapies* 20 (365): 1-15.
2. Lim, T.K. 2012. *Momordica cochinchinensis* Vol. 2, in *Edible Medicinal And Non-Medicinal Plants*, by T.K. Lim, 369-381. Dordrecht: Springer. doi:10.1007/978-94-007-1764-0_48.
3. Kiwon Jung, Young-Won Chin, Yoon Hee Chung, Yang Hae Park, Hunseung Yoo, Dong Sun Min, Bongyong Lee, and Jinwoong Kim. 2013. "Anti-gastritis and wound healing effects of *Momordicae Semen* extract and its active component." *Immunopharmacology and Immunotoxicology* 35 (1): 126-132.
4. Hung-Tse Huang, Yu-Chi Lin, Li-Jie Zhang, Chia-Ching Liaw, Hsin-Yen Chen, Ming-Tung Hsueh & Yao-Haur Kuo. 2019. "Anti-Inflammatory and anti-proliferative oleananetype-type triterpene glycosides from the vine of *Momordica cochinchinensis*." *Natural Product Research* 35 (16): 2707-2714.
5. Gulladawan Jan-on, Dr.Upa Kukongvirivapan, Boontium Kongsaktragoon, Dr.Poungnat Pakdeechote. 2015. "Effect of *Momordica cochinchinensis* Extract on Hemodynamics and Oxidative Stress in L-NAME-induced Hypertensive Rats." 34th the national graduate research conference. Khon Kaen: Khon Kaen University, Thailand. 1274-1280
6. Mitsuru Satsukawa, Tetsuhiro Kawamoto, Natsumi Yamanaka, Byung-Yoon Cha, Je-Tae Woo and Noriko Ogawa. 2016. "Fatty Liver Inhibitory Effect of Freeze-Dried Gac (*Momordica cochinchinensis*) Aril in Rats Fed a High-Fat Diet." 63 (1): 44-50.
7. Md. Mosiqr Rahman, Ashik Ahmed, Saibal Saha Sunny, Md. Samiul Huq Atanu, Abdullah Faruque and Md. Sohel Rana. 2014. "In-vitro Evaluation of Cytotoxic and Anthelmintic Activity of *Luffa acutangula*, *Luffa aegyptiaca* and *Momordica*

cochinchinensis." *British Journal of Pharmaceutical Research* 4 (2): 267-277.

8. Miran Jang, Gun-Hee Kim. "Antioxidant activity and hplc analysis of lycopene, β -carotene and α -tocopherol from geuk (*Momordica cochinchinensis* Spreng) fruit" *Journal of International Scientific Publications: Agriculture and Food* 2 (1): 430-438.

9. Pakapun Leevutinun, Panvipa Krisadaphong, and Amorn Petsom. 2015. "Clinical evaluation of Gac extract (*Momordica cochinchinensis*) in an antiwrinkle cream formulation." *J. Cosmet. Sci* 66 (3): 175–187.

10. Apichakan Sampanang, Supatcharee Arun, Wannisa Sukhorum, Jaturon Burawat, Somsak Nualkaew, Chanwit Maneenin, Bungorn Sripanidkulchai, Sittichai Iamsaard. 2017. "Antioxidant and Hypoglycemic Effects of *Momordica cochinchinensis* Spreng. (Gac) Aril Extract on Reproductive Damages in Streptozotocin (STZ)-Induced Hyperglycemia Mice." *Int. J. Morphol.* 35 (2): 667-675.

11. L. K. Bharathi, H. S. Singh, S. Shivashankar, A. N. Ganeshamurthy, P. Sureshkumar. 2013. "Assay of Nutritional Composition and Antioxidant Activity of Three Dioecious *Momordica* Species of South East Asia." *Proc. Natl. Acad. Sci.* 84 (1): 31-36.

12. Hanh Phan-Thi, Yves Waché. 2019. "Behind the Myth of the Fruit of Heaven, a Critical Review on Gac (*Momordica cochinchinensis* Spreng.) Contribution to Nutrition." *Current Medicinal Chemistry* 26 (24): 4585-4605.

13. Hiromitsu AOKI, Nguyen Thi Minh KIEU, Noriko KUZE, Kazue TOMISAKA, and Nguyen Van CHUYEN. 2002. "Carotenoid Pigments in GAC Fruit (*Momordica cochinchinensis* SPRENG)." *Biosci. Biotechnol. Biochem* 66 (11): 2479–2482.

14. Ali Abdulqader, Faisal Ali, Amin Ismail, and Norhaizan Mohd Esaa. 2018. "Gac (*Momordica cochinchinensis* Spreng.) fruit and its potentiality and superiority in-health benefits." *Journal of Contemporary Medical Sciences* 4 (4): 179–186.

15. Xuan T. Tran, Sophie E. Parks, Minh H. Nguyen, Paul D. Roach, Tuyen C. Kha. 2017. "Changes in physicochemical properties of Gac fruit (*Momordica cochinchinensis* Spreng.) during storage." *Australian Journal of Crop Science* 11 (4): 447-452.

16. Benjawan Thumthanaruk, Natta Laohakunjit, and Grady W. Chism. 2021. "Characterization of spray-dried Gac aril extract and estimated shelf life of Beta-carotene and lycopene." *PeerJ* 9:e11134: 1-16. doi:10.7717/peerj.11134.

17. Judith Müller-Maatsch, Jasmin Sprenger, Judith Hempel, Florence Kreiser, Reinhold Carle, Ralf M. Schweiggert. 2017. "Carotenoids from gac fruit aril (*Momordica cochinchinensis* [Lour.] Spreng.) are more bioaccessible than those from carrot root and tomato fruit." *Food Research International* 99 (2): 928-935.

18. Makaepa M. Maoto, Daniso Beswa, Afam I. O. Jideani. 2019. "Watermelon as a

potential fruit snack." *International Journal of Food Properties* 22 (1): 355–370. doi:10.1080/10942912.2019.1584212.

19. Imen Tlili, Chafik Hdidder, Marcello Salvatore Lenucci, Ilahy Riadh, Hager Jebari, Giuseppe Dalessandro. 2011. "Bioactive compounds and antioxidant activities of different watermelon (*Citrullus lanatus* (Thunb.) Mansfeld) cultivars as affected by fruit sampling area." *Journal of Food Composition and Analysis* 24 (3): 307-314. doi:10.1016/j.jfca.2010.06.005.

20. Giulia Bianchia, Anna Rizzoloa, Maurizio Grassi, Lia Provenzi, Roberto Lo Scalzo. 2018. "External maturity indicators, carotenoid and sugar compositions and volatile patterns in ‘Cuoredolce®’ and ‘Rugby’ mini-watermelon (*Citrullus lanatus* (Thunb) Matsumura & Nakai) varieties in relation of ripening degree at harvest." *Postharvest Biology and Technology* 136: 1-36. doi:10.1016/j.postharvbio.2017.09.009.

21. Shameena Beegum PP, Vidya Ram Sagar, Abhijit Kar, Eldho Varghese, Surender Singh, Harshwardhan Choudhary. 2022. "Identification and Quantification of Physicochemical and Bioactive Components from Sugar Baby Variety of Watermelon (*Citrullus lanatus*)." *Agricultural research* 11: 410–420. doi:10.1007/s40003-021-00583-7.

22. Cheol-Hyun Kim, Min-Kyung Park, Sung-Kyu Kim, Young-Hee Cho. 2014. "Antioxidant capacity and anti-inflammatory activity of lycopene in watermelon." *International Journal of Food Science and Technology* 49: 2083–2091. doi:10.1111/ijfs.12517.

23. Sabar Messaoudi, Soraya Tebibel, Aya Khadidja Beladjila, Fatima Khelifi Touhami, Zahia Kabouche. 2019. "Anti-hyperlipidemic, Anti-inflammatory and Antioxidant Activities of *Citrullus lanatus* ." *World Journal of Environmental Biosciences* 8 (1): 100-106.

24. O. A. Oseni, O. E. Odesanmi, F. C. Oladele. 2015. "Antioxidative and antidiabetic activities of watermelon (*Citrullus lanatus*) juice on oxidative stress in alloxan-induced diabetic male Wistar albino rats." *Nigerian Medical Journal* 56 (4): 272-277. doi:10.4103/0300-1652.169707.

25. Basiru Olaitan Ajiboye, Moturayo Tawakalt Shonibare, Babatunji Emmanuel Oyinloye. 2020. "Antidiabetic activity of watermelon (*Citrullus lanatus*) juice in alloxan-induced diabetic rats." *Journal of Diabetes & Metabolic Disorders* 19: 343–352. doi:10.1007/s40200-020-00515-2.

26. Francis S. Oluwole, Morufu E. Balogun, Adedeji G. Temitope. 2013. "Antisecretory Effects of Watermelon (*Citrullus lanatus*) Juice in Male Albino Rats." *Annual Review & Research in Biology* 3 (4): 358-366.

27. Aruna Podur, Debra L. Rateri, Shubin K. Saha, Sibum Saha, Alan Daugherty. 2013.

"Citrullus lanatus 'sentinel' (watermelon) extract reduces atherosclerosis in LDL receptor-deficient mice." *Journal of Nutritional Biochemistry* 24: 882–886.

28. 28 Mohammad Ashraful Islam, Saimon Shahriar, Tanveer Hossain, K. M. Yasif Kayes Sikdar, A. S. M. Monjur-Al-Hossain, Md. Raihan Sarkar and Mohammad. 2022. "In vitro Antioxidant and In vivo Analgesic Activities of Citrullus lanatus Rind and Flesh Extract: A Comparison ." *Bangladesh Pharmaceutical Journal* 25 (1): 67-72. doi:10.3329/bpj.v25i1.57842.

29. Waqar Ahmed Siddiqui, Muhammad Shahzad, Arham Shabbir, Ali Ahmad. 2018. "Evaluation of anti-urolithiatic and diuretic activities of watermelon (Citrullus lanatus) using in vivo and in vitro experiments." *Biomedicine & Pharmacotherapy* 97: 1212–1221. doi:10.1016/j.biopha.2017.10.162.

30. Hye Chang Rhim, Sung Jong Kim, Jewel Park, Ki-Mo Jang. 2020. "Effect of citrulline on post-exercise rating of perceived exertion, muscle soreness, and blood lactate levels: A systematic review and meta-analysis." *Journal of Sport and Health Science* 9: 553-561. doi:10.1016/j.jshs.2020.02.003.

31. Phukphon Munglue, Sajeera Kupittayanant and Pakanit Kupittayanant. 2014. "Effect of Watermelon (Citrullus lanatus) Flesh Extract on Sexual Behavior of Male Rats." *CMUJ NS Special Issue on Food and Applied Bioscience* 13 (1): 519-527.

32. Kara Blohm, Joshua Beidler, Phil Rosen, Jochen Kressler & Mee Young Hong. 2019. "Effect of acute watermelon juice supplementation on post-submaximal exercise heart rate recovery, blood lactate, blood pressure, blood glucose and muscle soreness in healthy non-athletic men and women." *International Journal of Food Sciences and Nutrition* 71 (4): 482-489. doi:10.1080/09637486.2019.1675604.

33. R. Andrew Shanely, Jennifer J. Zwetsloot, Thomas J. Jurrissen, Lauren C. Hannan, Kevin A. Zwetsloot, Alan R. Needle, Anna E. Bishop, Guoyao Wu, Penel. 2020. "Daily watermelon consumption decreases plasma sVCAM-1 levels in overweight and obese postmenopausal women." *Nutrition Research* 76 : 9-19. doi:10.1016/j.nutres.2020.02.005.

34. Heba A. El Gizawy, Alaadin E. El-Haddad, Yasmin M. Attia, Sally A. Fahim, Mai M. Zafer and Amr M. Saadeldeen. 2022. "In Vitro Cytotoxic Activity and Phytochemical Characterization (UPLC/T-TOF-MS/MS) of the Watermelon (Citrullus lanatus) Rind Extract." *molecules* 27 (8): 1-19. doi:10.3390/molecules27082480.

35. Omolola R. Oyenih, Blessing A. Afolabi, Ayodeji B. Oyenih, Olusegun J. Ogunmokun, Oluwafemi O. Oguntibeju. 2016. "Hepato- and neuro-protective effects of watermelon juice on acute ethanol-induced oxidative stress in rats." *Toxicology Reports* 3: 288–294. doi:10.1016/j.toxrep.2016.01.003.

36. Raajih Isma'il Al-Faruqi and Muhammad Ichwan. 2021. "Effect of watermelon (*Citrullus vulgaris*) rind juice in muscle endurance in healthy non-athlete volunteers" *AIP Conference Proceedings* 2342 120001-9. doi: 10.1063/5.0045732
37. Lim, T.K. 2012. *Solanum lycopersicum*. Vol. 6, in *Edible Medicinal And Non-Medicinal Plants*, by T.K. Lim, 339–363. Dordrecht: Springer. doi:10.1007/978-94-007-5628-1_41.
38. Riadh Ilahy, Chafik Hdider, Marcello S. Lenucci, Imen Tlili, Giuseppe Dalessandro. 2011. "Phytochemical composition and antioxidant activity of high-lycopene tomato (*Solanum lycopersicum* L.) cultivars grown in Southern Italy." *Scientia Horticulturae* 127: 255–261. doi:10.1016/j.scienta.2010.10.001.
39. Daniela Erba, M. Cristina Casiraghi, Albert Ribas-Agusti', Rafaela Ca'ceres, Oriol Marfa', Massimo Castellari. 2013. "Nutritional value of tomatoes (*Solanum lycopersicum* L.) grown in greenhouse by different agronomic techniques." *Journal of Food Composition and Analysis* 31: 245–251. doi:10.1016/j.jfca.2013.05.014.
40. Concepción Sánchez-Moreno, Lucía Plaza, Begonia de Ancos, M. Pilar Cano. 2006. "Nutritional characterisation of commercial traditional pasteurised tomato juices: carotenoids, vitamin C and radical-scavenging capacity." *Food Chemistry* 98: 749–756. doi:10.1016/j.foodchem.2005.07.015.
41. Alam P, Raka MA, Khan S, Sarker J, Ahmed N, Nath PD, Hasan N, Mohib MM, Tisha A, Taher Sagor MA,. 2019. "A clinical review of the effectiveness of tomato (*Solanum lycopersicum*) against cardiovascular dysfunction and related metabolic syndrome." *Journal of Herbal Medicine* 16. doi:10.1016/j.hermed.2018.09.006.
42. RC Agrawal, Rachana Jain, Wasim Raja, M Ovais. 2009. "Anticarcinogenic Effects of *Solanum lycopersicum* Fruit Extract on Swiss Albino and C57 Bl Mice." *Asian Pacific journal of cancer prevention* 10: 379-382.
43. Eduardo Fuentes, Reinhold Carle, Luis Astudillo, Luis Guzmán, Margarita Gutiérrez, Gilda Carrasco, and Iván Palomo. 2013. "Antioxidant and Antiplatelet Activities in Extracts from Green and Fully Ripe Tomato Fruits (*Solanum lycopersicum*) and Pomace from Industrial Tomato Processing" *Evidence-Based Complementary and Alternative Medicine* 2013: 1-9. doi:10.1155/2013/867578.
44. Asim K. Dutta-Roy, Lynn Crosbie, Margaret J. Gordon. 2001. "Effects of tomato extract on human platelet aggregation in vitro." *Platelets* 12 (4): 218-227. doi:10.1080/09537100120058757.
45. Mariel Colmán-Martínez, Miriam Martínez-Huélamo, Palmira Valderas-Martínez, Sara Arranz-Martínez, Enrique Almanza-Aguilera, Dolores Corella, Ramón Estruch, and Rosa M. Lamuela-Raventós. 2017. "trans-Lycopene from tomato juice attenuates inflammatory biomarkers in human plasma samples: an intervention trial." *Molecular Nutrition & Food Research* 61 (11): 1-29. doi:10.1002/mnfr.201600993.

46. Dawid, Jafer. 2016. "The Role of Tomato Products for Human Health (Solanum lycopersicum)- A Review ." *Journal of Health, Medicine and Nursing* 33: 66-74.
47. Ciro Langella, Daniele Naviglio, Marina Marino, Armando Calogero, Monica Gallo. 2018. "New food approaches to reduce and/or eliminate increased gastric acidity related to gastroesophageal pathologies." *Nutrition* 54: 26-32. doi:10.1016/j.nut.2018.03.002.
48. Yusuf Sabilu, Nuziyati, Andi Faisal Fachlevi, Syawal Kamiluddin Saptaputra and Healthy Hidayanty. 2017. "Tomato Juice (Lycopersicum commune) Reduces Blood Pressure in Elderly Hypertensive Indonesians in Kulisusu, North Buton." *Asian Journal of Clinical Nutrition* 9 (3): 111-117
49. Siti Fadlilah, Adi Sucipto and Mohamad Judha. 2020. "Cucumber (Cucumis sativus) and tomato (Solanum lycopersicum) juice effective to reduce blood pressure." *GSC Biological and Pharmaceutical Sciences* 10 (01): 001–008. doi:10.30574/gscbps.2020.10.1.0246.
50. Marja-Leena Silaste, Georg Alftan, Antti Aro, Y. Antero Kesa'niemi and Sohvi Ho'rkkö. 2007. "Tomato juice decreases LDL cholesterol levels and increases LDL resistance to oxidation." 98: 1251–1258. doi:10.1017/S0007114507787445.
51. Yu Yamamoto, Koichi Aizawa, Makiko Mieno, Mika Karamatsu, Yasuko Hirano, Kuniko Furui, Tatsuya Miyashita, Kazumitsu Yamazaki, Takahiro Inakuma, Ikuo Sato, Hiroyuki Suganuma, Teruaki Iwamoto. 2017. "The effects of tomato juice on male infertility." *Asia Pac J Clin Nutr* 26 (1): 65-71. doi:10.6133/apjcn.102015.17.
52. Lisa G. Wood, Manohar L. Garg, Heather Powell, & Peter G. Gibson. 2008. "Lycopene- rich treatments modify noneosinophilic airway inflammation in asthma: Proof of concept." *Free Radical Research* 42 (1): 94-102. doi:10.1080/10715760701767307.
53. Theresia Anita, Agus Suwandono, Ida Ariyanti, Noor Pramono, Suryati Kumorowulan. 2017. "Effect of consuming tomato (lycopersium commune) juice in lowering blood pressure in pregnant mothers with hypertension." *Belitung Nursing Journal* 3 (6): 707-711.
54. BLOIS, MARSDEN S. 1958. "Antioxidant Determinations by the Use of a Stable Free Radical." *Nature* 181: 1199–1200. doi:10.1038/1811199a0.
55. Ielciu, I., B. Sevastre, N.-K. Olah, A. Turdean, E. Chișe, R. Marica, I. Oniga, et al. 2021. "Evaluation of Hepatoprotective Activity and Oxidative Stress Reduction of Rosmarinus officinalis L. Shoots Tincture in Rats with Experimentally Induced Hepatotoxicity." *Molecules* 26 (1737): 1-15. doi:10.3390/molecules26061737.
56. Ruch RJ, Cheng S-J, Klaunig JE: Prevention of cytotoxicity and inhibition of intercellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogenesis* 1989, 10:1003–1008.
57. Naima Saeed, Muhammad R Khan, Maria Shabbir. 2012. "Antioxidant activity, total

phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L." *BMC Complementary and Alternative Medicine* 2012 12 (221): 1-12.

58. Shruti Shukla, Archana Mehta, Jinu John, Siddharth Singh, Pradeep Mehta, Suresh Prasad Vyas. 2009. "Antioxidant activity and total phenolic content of ethanolic extract of *Caesalpinia bonducella* seeds." *Food and Chemical Toxicology* (47): 1848–1851. doi:doi:10.1016/j.fct.2009.04.040.

59. Nazir Ahmad Wani, Sharmila Tirumale. 2018. "Evaluation of antioxidant properties of different extracts of *Chaetomium cupreum* SS02." *Bulletin of Faculty of Pharmacy, Cairo University* 56: 191–198. doi:10.1016/j.bfopcu.2018.08.001.