

RESPONSE OF OVERWEIGHT RATS TO PIEPLANT AND MILK VETCH INTAKE CONSIDERING INFLAMMATION MARKERS AND SOME OBESITY EVIDENCE

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Abstract

The aim of this study was to assess the influence of Pieplant and milk vetch intake considering inflammation markers, visceral fats, Leptin hormone and antioxidant enzymes on male overweight rats. Histological changes of liver and kidneys, phenol fraction of Pieplant and milk vetch was also in the scope of this investigation. A total number of thirty five adult male rats of the strain Sprague Dawley were involved via distribution on 5 groups being: Control (-) fed on regular basal diet, Control (+) rats fed on HFD, overweight rats fed on 5% Pieplant diet, overweight rats fed on 5% milk vetch diet, and overweight rats given a diet with 5% of both plant material. After 28 days of feeding intervention rats were sacrificed. Obesity resulted in increases of visceral fats, peritoneal fats and internal organs weight. Leptin hormone, bilirubin fractions (direct and indirect) inflammation markers (Procalcitonin (PCT), C-reactive protein (CRP), and erythrocyte sedimentation rate), While antioxidant enzymes decreased and some changes occurred in the structure of liver and kidneys. Feeding on Pieplant and specially the milk vetch diets revised these parameters. This seems to be probably resulting from more phenol fraction in milk vetch compared to Pieplant diets. Combination of both plants revealed a synergism desirable action compared to each of two plants: Pieplant, milk vetch alone.

Keywords: Pieplant, milk vetch, Inflammation markers, Antioxidant enzymes Obesity markers

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1. INTRODUCTION

Actually Pieplant (famous name is rhubarb) is the name of perennial plants of the genus rheum, which have high economic value Sina Pieplant is edible (Yi, 2010) and also it is a purgative drug in China (Barceloux, 2009) since 3000 years BC.As reported by Jiao and Du (2000), Pieplant suggested to show erasing several health disorders including diabetic nephropathy, chronic renal failure, gastrointestinal bleeding. Phytochemical studies on Pieplant has proved that important components were included such as the phenolic compounds (Gao et al., 2011).

Milk vetch (famous as astraglus) is known from ancient years in china, where many forms of things were prepared from this plant (including tea, powder capsules and others). This plant contains important beneficial components such as flavonoids, glycosides and trace elements (Shahrajabian et al., 2019). As reported by these authors milk vetch was used in treatment of important diseases like diabetes mellitus, kidney diseases, leukemia as well as uterine cancer. It is used also for protection against several types of diseases (physical and emotional), being of anti aging properties. Milk vetch was thought to prevent bore less. It contains antioxidants and immune supporting agents.

Pieplant has a wide several conventional uses. Milk vetch is widely known for its antioxidant capabilities and is used to treat intestine, liver, kidney, and intestinal infections. It is also known traditionally for its restorative powers on diabetes. (Shabeer et al., 2009). Pieplant It also abundant in lignans, tannins, flavonoids, Coumarin, and norlignan components (Huang et al., 1998) (Huang et al., 1996).

2. PATIENTS AND METHODS

• Dried Plants:

Pieplant and milk vetch were purchased from an herb shop in Cairo.

• Rats:

A total number of thirty five adult Sprague Dawley male rats with a common body weight of 150 ± 10 g and an age of about 7 weeks have been collected from the Research Institute of Ophthalmology in Giza, Egypt.

• Standard Diet:

The standard diet was as follows AIN (1993) containing of casein (12%), maize oil (10%), mineral and vitamin mixture (4%), cellulose (5%),

choline chloride (0.2%), methionine (0.3%), and corn starch (67.5%) are all present in this composition.

for getting the fat rats feeding carried out on highfat diet (HFD), which composed as follows (Liu et al. 2004): Casein (25%), maize oil (1%), saturated fat (sheep tail fat) (19%), choline chloride (0.25%), vitamin mix (1%), salt mixture (3.5%), cellulose (5%), L-casein (0.18%), and corn starch (35.07%) are the main components of fat diet.

The process of Herbs and Other Feed Components: All ingredients were ground into a fine powder using an electric grinder and stored in darkstoppered glass tubes in a cool, dry environment until (**Russo**, 2011).

• Animal Species and Experimental Layout:

Normal and overweight rats were housed in twine cages below ordinary laboratory circumstances, and fed on basal weight loss plan for one week as acclimatisation duration at 25 Ć.

- Grouping of rats turned into as the subsequent:
- **Group** (1): Control (-) where normal rats consumed the main diet.
- **Group (2):** Control (+) where fattened rats consumed the main diet.
- **Group (3):** Overweight rats consumed 5% Pieplant diet.
- Group (4): Overweight rats consumed 5% milk vetch diet.
- Group (5): Overweight rats consumed 5% combination of Pieplant and milk vetch diets.

• Methods:

At the end of experiment post 12 hours of fasting rats sacrificed and Receiving blood samples into clean centrifuge tubes, allowing them to clot, and then centrifuging them, peritoneal fat determined using the following equation fat. Peritoneal fat pad (PEP %) = $\frac{Fat \text{ weight}}{Final \text{ weight}} \times 100$

CBC was measured in laboratory at Shebin El-Kom, Minufiya (Mera Laboratory), where values included levels of Procalcitonin (PCT), erythrocyte sedimentation rate (ESR), and C-reactive protein (CRP). Serum direct bilirubin, indirect bilirubin and total bilirubin were determined (Chary and Sharma, 2014). SOD, CAT and GPx were assayed by the approaches used by **Habig et al.** (1974), Luck (1974), and Kakkar et al. (1984), respectively. According to Guillaume and Björntorp's approach, Leptin Elisakit's Leptin hormone determination was made in 1996. Phenolic compounds fractions were assessed as outlined by **Ben-Hammouda et al.**, (1995).

After being sacrificed internal organs (liver, heart, lungs, spleen and kidneys) removed, washed, wiped gently to remove excess water and then weighed, Fixed liver and kidney with 10% natural formalin, embedded in paraffin wax and thin sections prepared to be examined by light microscope according to **Drury and Wallington** (1980).

ANALYTICAL STATISTICS:

(Armitage et al., 2002) Used a computerized Costat Programme to perform a one-way ANOVA with a completely randomized factorial design to statistically analyze the data. The Duncan's Multiple Range Test was used to separate the means when a substantial mean impact was found. $P \le 0.05$ was used to determine whether differences between treatments were significant.

3. RESULTS AND DISCUSSION

The results of **Table (1)** show the levels of peritoneal fat pad of fat rats as influenced by feeding on pie plant and milk vetch diets.

Table (1): Visceral test of fattened rats level as responding to feeing on Pieplant and milk vetch diets.

Parameters Groups	PFP %
Control -ve in G1.	3.20 ± 0.008^{e}
Control +ve in G2.	5.04 ± 0.002^{a}
Pieplant (5%) in G3.	3.50±0.006 ^c
Milk vetch (5%) in G4.	3.62 ± 0.003^{b}
G5: Both together (5%).	3.40 ± 0.005^{d}
LSD	0.02

Each column has values that are statistically different from the others ($P \le 0.05$).

It is evident that visceral fat was higher in control (+) than control (-). In fattened rats, visceral fat accumulated clearly. less the never nutritional intervention especially with milk vetch diets and in particular when fat rats received the mixture of both plants lowered significantly the level of

visceral fat. This indicates a synergistic action considering the two plants diets. This refers that both plants may be of value for fat rats, possibly because of active components in the plants, as will be shown from phenolic comments analysis.

(ng/dl)	PCT (ng/dl)	ESR (mmol/L)
0.6 ± 0.12^{e}	0.10 ± 0.01^{d}	3.5 ± 0.8 ^c
$4.5\pm0.2^{\text{ a}}$	$0.40 \pm 0.012^{\ a}$	$6.7\pm0.51^{\rm \ a}$
3.6 ± 0.23^{b}	0.30 ± 0.022 ^b	5.4 ± 0.62^{b}
$2.1\pm0.31^{\text{ c}}$	$0.15 \pm 0.017^{\ c}$	3.8 ± 0.81^{c}
1.3 ± 0.32^{d}	0.10 ± 0.023 ^b	$3.4 \pm 0.47^{\circ}$
0.5	0.06	1.2
	$\begin{array}{c} 0.6 \pm 0.12^{\text{e}} \\ 4.5 \pm 0.2^{\text{a}} \\ 3.6 \pm 0.23^{\text{b}} \\ 2.1 \pm 0.31^{\text{c}} \\ 1.3 \pm 0.32^{\text{d}} \\ 0.5 \end{array}$	(ng/dl) $0.6 \pm 0.12^{\circ}$ 0.10 ± 0.01^{d} 4.5 ± 0.2^{a} 0.40 ± 0.012^{a} 3.6 ± 0.23^{b} 0.30 ± 0.022^{b} $2.1 \pm 0.31^{\circ}$ $0.15 \pm 0.017^{\circ}$ 1.3 ± 0.32^{d} 0.10 ± 0.023^{b}

Table (2): Inflammation blood markers of overweight rats as responding to feeing on Pieplant and milk vetch diets.

Each column has values that are statistically different from the others (P \leq 0.05).

From results of **Table (2)** it is obvious that the health of rat deteriorate by fattening, while the lowers inflammation marker values found for fat rats fed on Pieplant an especially on milk vetch diets. Best treatments, however, was in case of the combination of both plants diets. It should be mentioned that according to (**Saltiel and Olefsky**,

2017) raising of inflammation markers is undesired and indicate the bad health case. According to these authors, values for normal and obese persons for CRP, PCT & ESR were 0.7- 2.6, 2-7.3 ng/l; 0.10.9 ng/ml and ESR 0-5, 7.8 mmol/l respectively (**Barati et al., 2008**).

Table (3): Internal organs weights of overweight rats as responding to feeing on Pieplant and milk vetch diets.

Parameters	Liver (g)	Heart (g)	Lungs (g)	Spleen (g)	Kidneys (g)
Groups	(5)	(5)	(5)	(5)	
Control -ve in G1	$6.7 \pm 0.01^{\circ}$	1.3±0.2 °	1.1 ± 0.2^{b}	$0.8{\pm}0.06^{\rm d}$	2.0 ± 0.2^{d}
Control +ve, G2	8.3±0.04 ^a	1.8 ± 0.03^{a}	2.3±0.08 ^a	1.7±0.02 ^a	2.8±0.03 ^a
Pieplant (5%) in G3.	7.8 ± 0.06^{a}	1.2±0.07 °	1.6±0.03 ^b	1.3±0.04 °	2.1 ± 0.07 ^{cd}
Milk vetch (5%) in G4.	8.2±0.08 ^a	1.6±0.08 ^b	2.1±0.04 ^a	1.5±0.07 ^b	2.5 ±0.05 ^b
G5: Both together (5%).	7±0.03 ^b	1.1±0.05 °	1.5 ± 0.06^{b}	1 ±0.03 °	2.2 ± 0.06^{cd}
LSD	0.35	0.12	0.18	0.103	0.15

Each column has values that are statistically different from the others (P \leq 0.05).

The results shown that due to inflammations the internal organs weights raised. Meanwhile the reverse was found for pie plant and specially milk vetch feeding diets. All internal organs showed a considerable weight gain as compared to the control negative (-) group., however, consuming tested plant diets, in particulate the combination of both Pieplant and milk vetch improved appreciably the weight showing pounced decreases.

 Table (4): Total bilirubin, direct bilirubin and indirect bilirubin of overweight rats as responding to feeing on

 Pieplant and milk vetch diets

Parameters Groups	T. Bil (ng/dl)	D. Bil (ng/dl)	Ind. Bil (ng/dl)
Control -ve in: G1	2.1 ± 0.1 ^c	1.0 ± 0.22^{d}	$1.1 \pm 0.06^{\mathrm{c}}$
Control +ve in: G2	5.3 ± 0.13^{a}	3.9 ± 0.10^{a}	$2.4\pm0.05^{\text{ a}}$
Pieplant (5%) in: G3.	$3.2 \pm 0.21^{\text{ b}}$	$2.5 \pm 0.31^{\text{ b}}$	0.7 ± 0.06^{b}
Milk vetch (5%) in: G4.	$1.9 \pm 0.32^{\circ}$	$1.3\pm0.24~^{\rm c}$	0.8 ± 0.08 ^b
G5: Both together (5%).	$1.2 \pm 0.31^{\text{ d}}$	1.1 ± 0.14^{d}	0.1 ± 0.07^{d}
LSD	0.7	0.6	0.11

Each column has values that are statistically different from the others (P \leq 0.05).

Bilirubin is found when red blood cells are broken down. Bilirubin is a part of the bile, which is made in the liver and sulfed in the gallbladder. The abnormal buildup of bilirubin cause jaundice. Bilirubin is a yellowish substance made dursy the body's normal person of breathing down of recells.

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If liver is healthy, it will remove most of the bilirubin from the body. Level of T. bilirubin in normal person is usually 1.2 ng/dl According to **Corash, (1983).** Fattiness raised bilirubin reactions

in (**Table 4**) indicating that liver functions is affected which was eliminate via Pieplant and specially milk vetch diets, and in particular the combination of both plants.

SERUM ANTIOXIDANT ENZYMES:

Parameters Groups	Superoxide dismutase (mmol/L protein)	Glutathione peroxidase (mg/mL protein)	Catalase (mmol/L)
Control -ve in: G1	64.40±0.007 ^a	0.80 ± 0.003^{a}	0.18±0.002 ^a
Control +ve in: G2	31.34±0.001 ^e	0.42±0.006 ^e	0.13±0.006 ^c
Pieplant (5%) in: G3.	31.83 ± 0.008^{d}	0.65 ± 0.002^{d}	0.14 ± 0.002^{bc}
Milk vetch (5%) in: G4.	37.70±0.003 °	$0.70\pm0.008^{\circ}$	0.15 ± 0.007^{b}
G5: Both together (5%).	63.30±0.004 ^b	0.77 ± 0.007^{b}	0.17±0.003 ^a
LSD	0.010	0.010	0.03

Table (5): SOD, GPx and CAT enzymes of overweight rats as responding to feeing on Pieplant and milk vetch diets.

Each column has values that are statistically different from the others (P \leq 0.05).

It was found that fattiness lower significantly the levels of SOD, GPX and CAT enzymes. Meanwhile, nutritional intervention using Pieplant and milk vetch diets, specially the last diet reversed such a day. Combination of both plant raised more the levels of the enzymes significantly indicating synergism action. On rats, **Jiangwei et al.**, (2011) and **He et al.**, (2012) confirmed the value of feeding on Pieplant and milk vetch for raising serum antioxidants.

Table (6): Leptin hormone of overweight rats as responding to feeing on Pieplant and milk vetch diets

Parameters	L H
Groups	(ng/ml)
Control -ve in : G1	3.45 ± 0.002^{e}
Control +ve in : G2	28.30 ± 0.007^{a}
Pieplant (5%) in: G3.	7.30 ± 0.002 ^c
Milk vetch (5%) in: G4.	7.40 ± 0.004^{b}
G5: Both together (5%).	7.01 ± 0.006^{d}
LSD	0.008

Each column has values that are statistically different from the others ($P \le 0.05$). LH=Leptin hormone.

The results of this study of **Table (6)** show that due to fattiness Leptin hormone was increased. Meanwhile by feeding on pie plant diet and specially the milk vetch diets Leptin hormone reduced. Maximum decrease recorded for the combination diets of both to lower. The last diet revealed very close value of Leptin hormone in comparison with normal rats. The value of feeding on Pieplant and milk vetch plants supported by conclusions of **Yu et al.**, (2003) and **Huang et al.**, (2017) working on fat rats.

Most of side effects due to fattiness are due to the oxidation stress and less of immunity Fernandez et al., (2011). At the same time phenolic compounds may reverse such a change because of the ant oxidation effect of mentioned compounds Savini et al., (2013). Therefore, in measuring phenolic compounds were determined in Pieplant and milk vetch plants (**Table 7**).

Groups	Pieplant	Milk vetch
Syringic	2.9	2.01
Pyrogallol	500.26	398.15
Gallic	12.15	36.81
Protocathechias	25.60	138.55
Catechol	17.32	55.03
4-Aminobenzoic	13.78	28.6

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Catechein	18.00	30.15
	18.00	50.15
Chlorogenic	17.82	80.27
P-OH-Benzoic	86.62	130.14
Epicatechen	76.19	86.51
Caffeic	7.84	21.30
Vanellic	25.17	33.16
Ferulic	4.26	7.24
Caffeine	140.31	150.32
Benzoic	68.54	80.41
Ellagic	167.11	247.11
Coumarin	12.17	18.34
Cinnamic	2.36	7.01
Total	1198.42	1551.15

From **table** (7)'s results, it was conceivable to see that the phenolic compounds was much more in milk vetch (about 30% more) as compared with

Histopathological Alterations:

In a microscopic slice of the healthy (control -) group 1 liver, the hepatic lobule's typical structure could be seen (Photo 1). However, the livers of the rats in group 2 (control + fat) exhibited symptoms of hepatocyte vacuolar degeneration (Photos 2 & 3). With the exception of a minimal central vein constriction and a mild swelling of the hepatocytes, rats fed group 3 Pieplant meals in their liver segments did not show any abnormalities (Photos 4 and 5). Additionally, rats in the Milk vetch diet-fed fat group (group 4) displayed modest hydropic

that of Pieplant which was in line with the more pronounced effect of milk vetch considering biochemical results of present work.

hepatocyte degeneration and slight hepatic sinusoids congestion in the liver (Photos 6 and 7). Hydropic hepatocyte degeneration, a small hepatic sinusoidal congestion, and a slight activation of Kupffer cells (Photo 9) were also seen.

It is clear that the Pieplant, Milk vetch, and particularly the different meals benefited. The histological form of the kidney and liver, resulting in was changed by weight gain and happened at the same time as the biochemical changes that occurred.

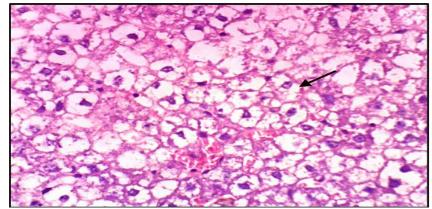


Photo (1): shows the typical histological structure of hepatic lobules in the liver of a group 1 (healthy rats) rat.

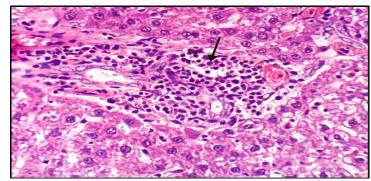


Photo (2): Hepatocyte vacuolar degeneration is visible in the rat liver from group 2 (fat rats) *Eur. Chem. Bull.* 2023,12(5), 4431-4439

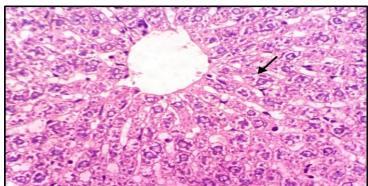


Photo (3): Rat liver from group 2 (fat animals) demonstrating inflammatory cell infiltration in the portal triad and vacuolar degeneration of hepatocyte

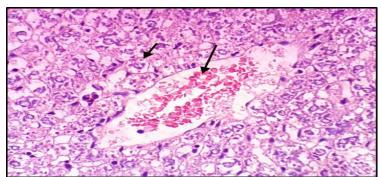


Photo (4): Hepatocytes are mildly hydropically degenerating in the liver of a group 3 (5% Pieplant) rat

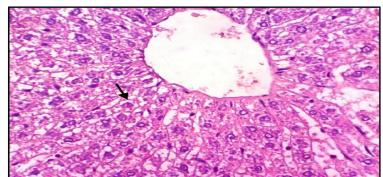


Photo (5): Rat liver from group 3 (5% Pieplant) demonstrating minor hydropic hepatocyte degeneration and slight central venous congestion

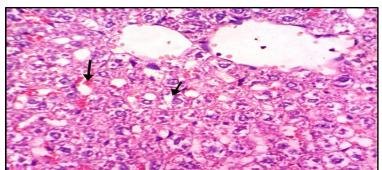


Photo (6): Rat liver from group 4 (milk vetch 5%) demonstrating mild hydropic hepatocyte degeneration.

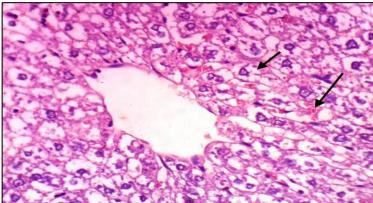


Photo (7): Rat liver from group 4 (milk vetch 5%) with mild hydropic hepatocyte degeneration and mild hepatic sinusoidal congestion

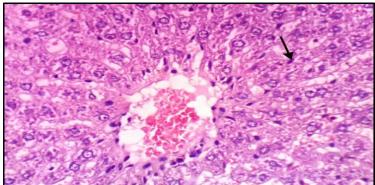
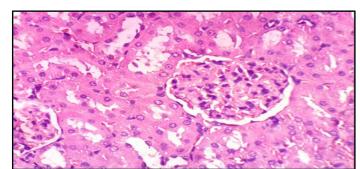


Photo (8): Rat liver from group 5 (mixed diets), showing hydropic hepatocyte degeneration and mild hepatic sinusoidal congestion



Liver of a rat from group 5 (mixed diets) demonstrating mild Kupffer cell activation in photo (9)

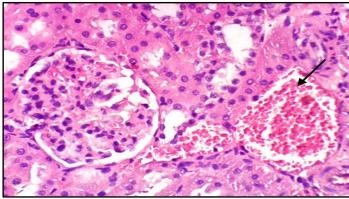


Photo (10): Photomicrograph of the renal parenchyma of a healthy rat from group 1 exhibiting the typical histological structure.

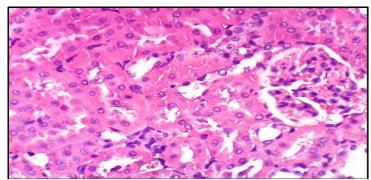


Photo (11): Congestion of renal blood vessels in a rat kidney from group 2 (overweight animals) (H & E X 400).

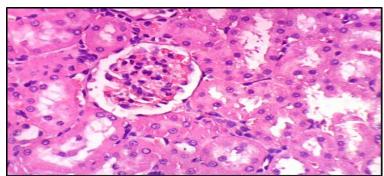
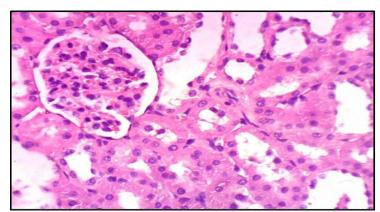


Photo (12): A rat kidney from group 3 (5% Pieplant) that has not had any histological changes (H & E X 400).



Rat kidney from group 4 (milk vetch 5%) in image 13 is shown without any histopathological changes (H & E X 400).

Rat kidney from group 5 (mixed diets), shown in photo (14) with no histopathological changes (H & E X 400).

4. REFERENCES

- American Institute of Nutrition (AIN) (1993): Purified diet for laboratory rodent; final report. J. Nutrition, 123:1939-1951.
- Armitage, P.G.; Berry and Mattews, J.M.S. (2002): Statistical Methods in M edical Research.4thEd.BlackwellScienceLtd.
- Barati, M.; Alinejad, F.;Bahar, M.A.; Tabrisi, M.S.; Shamshiri, A.R.; Bodouhi, N.O and Karimi, H.(2008): Comparison of WBC, ESR, CRP and PCT serum levels in septic and non-septic burn cases. Burns ; 34(6):770–774
- **Barceloux, D. G. (2009):** Rhubarb and oxalosis (Rheum species). Disease-a-Month, 6(55): 403-411.
- BenHammouda,M.;Kremer,R.J.;Miror,H.C.and Sarwar,M.(1995):Achemical basis in diuerential allelopathic pytential of soglum hybrids on wheat.J.Chem.Ecoli,21:775-786.
- Chary,T.M.and Sharma,H.(2004):Practical Biochemistry for Medical and Dental Students. Jaypee Brothers Medical Publishers(P)LTD, New Delhi.
- Corash, L. (1983): Platelet Sizing: Techniques, Biological Significance, and Clinical Applications. Current Topics in Haematology. New York: Alan R. Wise, Inc.

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- Drury, R.A. and Wallington, E.A. (1980): Cartons Histological Technique. 5th Ed., Oxford University Press, 250.
- Fernandez-Sanchez A.; Madrigal-Santillan E. and Bautista M.(2011):Inflammation, oxidative stress, and obesity. Int J Mol Sci ;1 2:3117–3132.
- Gao, L.L.; Xu, X.D.; Nan, H.J.; Yang, J.S. and Chen, S.L. (2011): Chemical constituents in Rheum tanguticum. Chinese Traditional and Herbal Drugs, 42(3): 443-446.
- Gholami, S.; Mirzaei, A.; Oryan, S. and Hossaeni, S.E. (2015): The Effect of Rhubarb Extracts on Lipid Profile and Oxidative Stress in Wistar Male Rats. International Medical Journal, 22 :(2).
- Guillaume, M. and Björntorp, P. (1996): Obesity in children, environmental and genetic aspects Hormone and Metabolic Research., 28(11): 573-581.
- Habig, W.H.; Pabst, M.J. and Jakoby, W.B. (1974): Glutathione S-transferases the first enzymatic step in mercapturic acid formation. Journal of Biological Chemistry. 249(22): 7130-7139.
- He, L. F.; Du, J. R. and Yu, L. (2012): Hepatoprotective effects of Rhubarb Choleretic capsule against alcoholic fatty liver in rats. Chinese Journal of General Practice, (11): 3.
- Huang, Y.C.; Tsay, H.J.; Lu, M.K.; Lin, C.H.; Yeh, C.W.; Liu, H.K. and Shiao, Y. J. (2017): Astragalus membranaceuspolysaccharides ameliorates obesity, hepatic steatosis, neuroinflammation and cognition impairment without affecting amyloid deposition in metabolically stressed APPswe/PS1dE9 mice. International Journal of Molecular Sciences, 18(12): 2746.
- Huang,Y.L.;Chen,C.C.;Hsu,F.L.and
- Chen, C.F. (1996): Anew lignin from Pieplant .J.Nat. Prod. 59:520-1.
- Huang, Y.L.; Chen, C.C.; Hsu, F.L. and Chen, C.F. (1 998): Tannins, Flavonol sulphates and a norlignan from Pieplant. J. Nat. Prod. 61:1194-7.
- Jiangwei, M.A.; Zengyong, Q. and Xia, X. (2011): Aqueous extract of Astragalus mongholicus ameliorates high cholesterol diet induced oxidative injury in experimental rat models. Journal of Medicinal Plants Research, 5(5): 855-858.

- Jiao, D.H. and Du, S.J. (2000): The Study of rhubarb. Shanghai Science & Technology Press, Shanghai, 291.
- Kakkar, P.; Das, B. and Viswanathan, P.N. (1984): A modified spectrophotometric assay of superoxide dismutase. Ind. J. Biochem. Biophys., 21:130-132.
- Liu, M.; Shen, L.; Liu, Y.; Woods, S.C.; Seeley, R.J.; D'Alessio, D. and Tso, P. (2004):Obesity induced by a high-fat diet down regulates apolipoprotein A-IV gene expression in rat hypothalamus. American Journal of Physiology-Endocrinology and Metabolism, 287(2): 366-370.
- Luck, H. (1974): Catalase. In: Methods of Enzymatic Analysis, Vol. II, edited by J. Bergmeryer & M. Grabi, Academic Press, New York, P.P. 885-890.
- **Russo, E. (2001):** Handbook of Psychotropic Herbs: A Scientific Analysis of Herbal Remedies for Psychiatric Condition. The Howrth Herbal Press, Inc.
- Savini, I.; Catani, M.V. and Evangelista, D.(2013):Obesity-associated oxidative stress: Strategies finalized to improve redox state. Int J Mol Sci ; 14:10497–10538.
- Saltiel, A.R. and Olefsky, J.M.(2017): Inflammatory mechanism linking obestity and metabolic disease. J. Clin. Investig., 127, 1–4.
- Shahrajabian, M. H.; Sun, W. and Cheng, Q. (2019): Astragalus, an ancient medicinal root in traditional Chinese medicine, a gift from Silk Road. International Journal of Agriculture and Biological Sciences, 3(06): 27-38.
- Shabeer, J.; Srivastava, R.S. and

Singh,S.K.(2009): Antidiabetic and antioxidant effect of various fractions of Pieplant in Alloxan diabetic rats.J.Ethnopharmacol;12:34-8.

- Yi, H.Y. (2010): Application of Molecular Techniques in The Research of Germplasm Resources of Rheum. Biotechnology Bulletin, 12.
- Yu, C.; Xiao-yi, Q. and Hui-ming, J. (2003): Effects of Rhubarb Compound of on Adipocyte Leptin and C/EBP# alpha# Expression in Obese Rats. Chinese Journal of Basic Medicine in Traditional Chinese Medicine.; 9(4): 27-3.