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Determination of antimicrobial resistance patterns and detection of ESBL and AMPC: Producing uropathogenic *E. coli*

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Background: Uropathogenic E. coli's multidrug resistance has become a potential threat to global health. The purpose of the current study is to evaluate the antimicrobial against Extended Spectrum - Lactamases (ESBL) and AmpC producer *E. coli* isolates.

Methodology: All clinical isolates of *Escherichia coli*, isolated from UTI patients were evaluated for susceptibility to a panel of antimicrobials and were analysed for the ESBL and AmpC using Vitek 2.

Results and conclusions: *Total 1911 no of urine samples were screened out of these 782 were having the significant bacteriuria. In this 226 sample have shown the growth of E.Coli.* Maximum no of patients are of age group more than 60 years (35.41%).52.21% were ESBL producers. The AST of ESBL *E.Coli* showed isolates were sensitive to Tigecycline (100%) and were 99.15% sensitive to Imipenem and Meropenem followed by 97.46% to Amikacin. The 37 no wee identified the AmpC. Further research is required to detect ESBL and AmpC enzyme production simultaneously in such isolates. It is necessary to detect ESBL and AmpC-producing E. coli early to prevent treatment failure. Continuous surveillance and prudent antibiotic use, as well as the implementation of effective infection control measures, are essential.

Keywords: Uropathogenic E.Coli, ESBL, AmpC, MDR

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Antimicrobials are often used in hospitals, with up to 70% of patients in the intensive care unit (ICU) and 35% of adult ward patients, respectively, receiving one on any given day¹. The patient is at danger of getting multidrug resistant (MDR) pathogens due to this heavy antibiotic burden. Exogenous crosscontamination, or the transmission of MDR organisms from other patients or the healthcare environment, or selection pressure exerted on the patient's own microbiome, which gives mutant strains a competitive edge².

ESBLs are resistant to all beta-lactams with the exception of cephamycins and carbapenems. Therapeutic failure has been described with third-generation cephalosporin with an in vitro pattern of intermediate or greater sensitivity, thus once ESBLs production is confirmed, the strain must be considered resistant to all beta-lactams, except for carbapenems and cephamycins. However, cefoxitin, cefotetan and cefamandole are not recommended as treatment options due to the risk of developing antibiotic resistance during the course of the treatment³.

Carbapenems are the treatment of choice for severe infections, since they seem to be the only ones capable of maintaining bactericidal activity for 24 hours against high inocula, with the exception of ertapenem. In case of septic shock, meropenem and imipenem are mostly recommended. In mild cases, piperacillin/tazobactam and amoxicillin/ clavulanic acid or fluoroquinolones may be considered⁴. ESBLs-producing E. coli is the most frequently isolated infectious agent in this group.

Since its discovery in 1983, Extended Spectrum -Lactamases (ESBL)-producing organisms have become a significant issue in the field of infectious diseases⁵.Since they can totally hydrolyze oximino-beta lactams like third generation cephalosporins, which are typically used to treat hospital acquired infections, the ESBL and AmpC generating E. coli have posed a serious hazard to hospitalized patients. This plasmid-mediated resistance may spread more easily due to the use of broad-spectrum oral antibiotics and likely subpar infection control procedures. Ambulatory patients with chronic illnesses constitute another patient population that may host ESBL-producing microbes in addition to identified populations at risk⁵⁻⁹. On the chromosomes of several Enterobacteriaceae and a few other organisms are encoded the therapeutically significant cephalosporins known as AmpC -lactamases. Where they have a role in the resistance to most penicillins, cephalothin, cefazolin, cefoxitin, and -lactamase inhibitor/-lactam combos. Clavulanic acid, an ESBL inhibitor, had no effect on the activity of AmpC -lactamase. While they were given Group 1 in Bush et al functional categorization system, AmpC enzymes are classified as Class-C in the Ambler structural classification of beta lactamases. Members of the Enterobacteriaceae family can have either chromosomal or plasmid-mediated AmpC -lactamase genes. AmpC overproduction and outer membrane porin mutations can lessen sensitivity to carbapenems, especially in plasmid-mediated AmpC producers¹⁰⁻¹².

Urinary tract infection is one of the most common infections, mostly caused by gram negative bacteria and out of which E Coli is most common isolate. E coli isolates are becoming

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more ESBL & AmpC Beta lactamase producing Uropathogens. The present study was undertaken to study antibiotic resistance patterns, including ESBLs & Amp C beta lactamases in urinary isolates of E Coli.

Material and methods

After approval of Institutional Ethics committee the present study was conducted at a tertiary care hospital. A total of 1911clean catch midstream urine samples were collected in a sterile container for the period Jan 2022 to Oct 2022. Urine samples were transported immediately to Department of Microbiology for processing. Then the urine samples were inoculated on MacConkey's and Blood agar plates by using calibrated loop delivering 0.001 ml of sample and incubated at 37°C aerobically for 24 hrs. For gram negative bacilli more than 10⁵ colonies per ml of single organism were considered significant.

All isolates were identified and tested for susceptibility by the Vitek 2 system (bioMerieux; Inc., Durham, NC, USA) using the Gram-negative strain cards AST N280, AST N281, AST N405, AST N406. The isolates were tested for susceptibility for the following antimicrobial agents: amikacin, cefepime, cefoperozone, ciprofloxacin, colistin, gentamicin, imipenem, meropenem, piperacillin/tazobactam, tigecycline, and trimethoprim sulfamethoxazole.

All results were interpreted using the Advanced Expert System (software version VT2-R04.03). Using the Vitek 2 system according to the manufacturer's recommendations. Whilst the Vitek 2 Advanced Expert SystemTM (AES), which automatically compares minimum inhibitory concentrations (MICs) of antibiotics to a large database and suggests possible mechanisms of resistance, tends to favour the most conservative options to ensure safe patient treatment, it may thereby add to the over-calling of ESBLs. An isolate was considered an ESBL producer if the phenotypic interpretation by the AES of the Vitek 2 included ESBL with or without decreased outer membrane permeability and not an ESBL if only wild type or β -lactamases other than ESBLs were suggested by AES.

Results

Summarization and analysis of data was carried out by using software statistical package for social sciences (SPSS-20 version). Data was condensed in the form of tables. Statistics like percentages and mean were computed. Chi square test was applied to study the association. Chi square test were said to be significant when probability was less than 0.05.

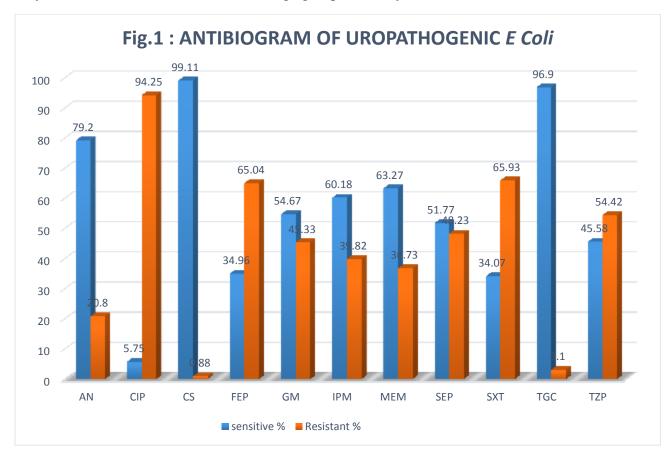
The Department of microbiology received total 1911 no of urine samples during Jan. 2022 to Oct. 2022. Out of these total urine samples, 782 no of sample were positive for one or two organisms having the significant bacteriuria. In this 226 sample have shown the growth of *E.Coli*.

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Sr. NO.	Age Group (Yrs.)	Male (%)	Female (%)	Total (%)
1	<20	6(5.22)	8(7.21)	14(6.19)
2	21 - 40	32(2.83)	33(29.73)	65(28.76)
3	41 - 60	28(24.35)	39(35.14)	67(29.64)
4	>60	49(42.60)	31(27.92)	80(35.41)
5	Total	115(100)	111(100)	226(100)
$\chi^2 = 6.088, df = 3, P = 0.1074$				

Table1: Age and Sex distribution of Cases.

The age and sex distribution of UTI cases having growth of *E.Coli* are shown in table1. Maximum no of patients are of age group more than 60 years (35.41%) followed by age group of 41to 60, followed by 21to 40 age group. Maximum number of males are of age group more than 60 years while maximum females are of age group 41to 60 years.



AN-AMIKACIN CS- COLOSTIN GM- GENTAMICIN CIP-CIPROFLOXIN FEP- CEFEPIME IPM- IMIPENEM

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MEP- MEROPENEM SXT- TRIMETHOPRIM SULFAMETHOXAZOLE TZP- PIPERACILLIN/TAZOBACTUM

SEP-CEPEROZONE TGC- TIGECYCLINE

	ESBL Positive(118)			ESBL Negative(108)		
Antibiotic	Sensitive (%)	Resistant (%)	Intermedi ate (%)	Sensitive (%)	Resistant (%)	Intermedia te (%)
Amikacin	115 (97.46)	3(2.54)	0 (00.00)	64(59.26)	44(40.74)	2(1.85)
Ciprofloxacin	8 (6.78)	101(85.59)	9(7.63)	5(4.63)	103(95.37)	3(2.78)
Colistin	118 (100)	0(0.00)	0(00.00)	106(98.15)	2(01.85)	0(00.00)
cefepime	61(51.69)	57(48.31)	0(00.00)	19(17.59)	89(82.41)	0(00.00)
Gentamicin	78 (66.10)	40(33.90)	0(00.00)	45(41.67)	63(58.33)	1(0.93)
Imipenem	117 (99.15)	1(0.85)	1(00.85)	20(18.52)	85(78.70)	13(12.04)
Meropenem	117 (99.15)	0(0.00)	6(05.08)	26(24.07)	82(75.93)	0(00.00)
Cefoperozone	99 (83.90)	13(11.02)	0(00.00)	18(16.67)	90(83.33)	1(0.93)
Trimethoprim/ sulfamethoxazole	48 (40.68)	70(59.32)	0(00.00)	29(26.85)	79(73.15)	0(00.00)
Tigecycline	118 (100)	0(0.00)	0(00.00)	101(93.52)	7(06.48)	6(5.56)
Piperacillin/tazobact um	87 (73.73)	25(21.19)	6(05.08)	17(15.74)	91(84.26)	0(00.00)

Table2: Antimicrobial susceptibility testing pattern

Fig. 1 & Table No 2 shows the Antimicrobial Susceptibility testing pattern in all E.coli isolates. There were 118 ESBL positive *E.Coli*. The ESBL positive *E.Coli* isolates all the isolates were sensitive to Tigecycline (100%) and were 99.15% sensitive to Imipenem and Meropenem followed by 97.46% to Amikacin. for Cefoperozone 83.90% sensitivity while for Piperacillin/Tozobactum 73.73% isolates were sensitive. ESBL *E.coli* showed 51.61% & 40.68% sensitivity to Cefepime and Trimethoprim/Sulfamethoxazole respectively.

The out of 226 *E.Coli* grown in the UTI cases, the AES identified the 118 samples as the extended spectrum B lactamases. Out of the remaining 108 *E.Coli*, the Carbapenemase Resistant Enterobacteriaceae (CRE) stains were 84. The difference between *E.Coli* and ESBL & CRE E.Coli is statistically significant (Table No. 3)

Organism	Non ESBL	ESBL	Total
E.Coli	24	118	142
E.Coli CRE	84	00	84
Total	108	118	226
$\chi^2 = 146.069, df = 1, P = < 0.005$			

Table No. 3: ESBL producing E.Coli.

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Organism	Non AmpC	AmpC	Total
E.Coli	105	37	142
E.Coli CRE	84	00	84
Total	189	37	226
$\chi^2 = 20.192, df = 1, P = < 0.003$			

Table No. 4: AmpC producing *E.Coli*

Table 4 shows the distribution of AmpC producing *E.Coli*. The 37 no of UTI cases have identified the AmpC out of 226 *E.Coli*. The AmpC *E.Coli* & *E.Coli* CRE, the difference is statistically significant.

The maximum number of ESBL *E.Coli* were found in the age group of more than 65 years (45 nos.) followed by 33 nos. in age group of 41 to 60 years and 32 isolates of ESBL *E.Coli* in the age group of 21 to 40 years and very less no (08) isolates in the age group of below 20 years (Table5). The sex wise distribution of ESBL *E.Coli* is equally distributed there are 59 females and 59 in males.

Sr. NO.	Age Group (Yrs.)	Non ESBL	ESBL	Total (%)
1	<20	6	8	14
2	21 - 40	33	32	65
3	41 - 60	34	33	67
4	>60	35	45	80
5	Total	108	118	226

Table 5: Age distribution of ESBL & non ESBL Cases.

Discussion

The prevalence of ESBL-producing Enterobacteriaceae has risen alarmingly in the current period of expanding usage of broad-spectrum antibiotics¹³. ESBLs and AmpC -lactamase producing *E. coli* are currently a global problem for hospitalized patients. The prevalence of ESBLs and AmpC production among urine isolates varies substantially globally, geographically, and through time, and is subject to fast change.

In the present study in 226 urine sample showed *E.Coli* growth. The ESBL production was observed in 52.21% of *E. Coli*. The similar results 50-80 % ESBL production have been reported in other studies also¹⁴⁻¹⁸. Antimicrobial selection varies from region to region, and so does ESBL production.

In present study 37 isolate were detected as AmpC producing E.Coli out of total 226 E.Coli (16.37%).Bakshi R^6 et al showed the 66.6% isolates by AmpC disc method. In the same study they have reported Susceptibility of ESBLs and AmpC producers E. coli to fosfomycin,

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imipenem, nitrofurantoin and amikacin were found to be 100%, 98%, 83.8% and 72% respectively¹⁸. In the present study our ESBL isolates were Imipenem & Meropenem 99% sensitive, Amikacin 97.46% Piperacillin/Tozobactum 73.73% sensitive to these antibiotics. Similarly other studies have also reported susptibility of ESBL producers to imipenem and amikacin at 100% & 68% respectively^{9,19}.

In present study out of 226 *E.Coli*, the AES identified the 118 samples as the extended spectrum B lactamases. Out of the remaining 108 *E.Coli*, the Carbapenemase Resistant Enterobacteriaceae (CRE) stains were 84(77.78%) which is also the alarming sign for treating the patients in tertiary care hospital. The increasing drug resistance of bacteria is the major cause of treatment failure of urinary tract infection cases in hospitals.

References

- Versporten A, Zarb P, Caniaux I, Gros MF, Drapier N, Miller M, Jarlier V, Nathwani D, Goossens H, Koraqi A, Hoxha I. Antimicrobial consumption and resistance in adult hospital inpatients in 53 countries: results of an internet-based global point prevalence survey. The Lancet Global Health. 2018 Jun 1;6(6):e619-29.
- Arulkumaran N, Routledge M, Schlebusch S, Lipman J, Conway Morris A. Antimicrobial-associated harm in critical care: a narrative review. Intensive care medicine. 2020 Feb;46(2):225-35.
- López-Pueyo MJ, Barcenilla-Gaite F, Amaya-Villar R, Garnacho-Montero J. Antibiotic multiresistance in critical care units. Medicina Intensiva (English Edition). 2011 Jan 1;35(1):41-53.
- Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. Infectious Diseases Society of America 2022 Guidance on the Treatment of Extended-Spectrum βlactamase Producing Enterobacterales (ESBL-E), Carbapenem-Resistant Enterobacterales (CRE), and Pseudomonas aeruginosa with Difficult-to-Treat Resistance (DTR-P. aeruginosa). Clinical Infectious Diseases. 2022 Apr 19.
- 5. Gupta V, Yadav A, Joshi RM. Antibiotic resistance pattern in uropathogens. Indian journal of medical microbiology. 2002 Apr 1;20(2):96-8.
- 6. Babypadmini S, Appalaraju B. Extended spectrum β-lactamases in urinary isolates of Escherichia coli and Klebsiella pneumoniae-prevalence and susceptibility pattern in a tertiary care hospital. Indian Journal of medical microbiology. 2004 Jul 1;22(3):172-4.
- Lautenbach E, Patel JB, Bilker WB, Edelstein PH, Fishman NO. Extended-spectrum βlactamase-producing Escherichia coli and Klebsiella pneumoniae: risk factors for infection and impact of resistance on outcomes. Clinical infectious diseases. 2001 Apr 15;32(8):1162-71.
- 8. Jabeen K, Zafar A, Hasan R. Comparison of double disc and combined disc method for the detection of extended spectrum beta lactamases in enterobacteriaceae. Journal of Pakistan Medical Association. 2003;53(11):534.

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- El-Hady SA, Adel LA. Occurrence and detection of AmpC β-lactamases among Enterobacteriaceae isolates from patients at Ain Shams University Hospital. Egyptian Journal of Medical Human Genetics. 2015;16(3):239-44.
- 10. Tulara NK. Nitrofurantoin and fosfomycin for extended spectrum beta-lactamases producing Escherichia coli and Klebsiella pneumoniae. Journal of global infectious diseases. 2018 Jan;10(1):19.
- 11. Grover N, Sahni AK, Retd SB. Therapeutic challenges of ESBLS and AmpC betalactamase producers in a tertiary care center. Medical Journal Armed Forces India. 2013 Jan 1;69(1):4-10.
- 12. Sah BS, Aryal M, Bhargava D, Siddique A. Drug resistance pattern of bacterial pathogens of Enterobacteriaceae family. Tribhuvan University Journal of Microbiology. 2017;4:15-22.
- 13. Paterson DL, Bonomo RA. Extended-spectrum β-lactamases: a clinical update. Clinical microbiology reviews. 2005 Oct;18(4):657-86.
- 14. Azap ÖK, Arslan H, Şerefhanoğlu K, Çolakoğlu Ş, Erdoğan H, Timurkaynak F, Senger SS. Risk factors for extended-spectrum β-lactamase positivity in uropathogenic Escherichia coli isolated from community-acquired urinary tract infections. Clinical microbiology and infection. 2010 Feb 1;16(2):147-51.
- 15. Chervet D, Lortholary O, Zahar JR, Dufougeray A, Pilmis B, Partouche H. Antimicrobial resistance in community-acquired urinary tract infections in Paris in 2015. Medecine et maladies infectieuses. 2018 May 1;48(3):188-92.
- 16. Sood S, Gupta R. Antibiotic resistance pattern of community acquired uropathogens at a tertiary care hospital in Jaipur, Rajasthan. Indian journal of community medicine. 2012 Jan 1;37(1):39-44.
- 17. Yashavanth R, Ronald R, Anita K, Narendra N, Faseela T, Rai YK, AJIMS M. Uropathogens and their antimicrobial susceptibility pattern in a tertiary care hospital. Journal of Evolution of Medical and Dental Sciences. 2012 Oct;1(4):467.
- 18. Bakshi R, Sehgal VK, Kansal P, Kaur S. Detection of extended-spectrum beta lactamases and AmpC beta lactamases producing uropathogenic Escherichia coli in a tertiary care hospital. Int J Med Dent Sci. 2019;8:1783-92.
- 19. Madhavi VN, Subbulu P. Prevalence and antibacterial susceptibility testing pattern of bacterial pathogens causing urinary tract infections in community. Journal of Evolution of Medical and Dental Sciences. 2016 Apr 14;5(30):1528-32.